

A

Mechanobiology

Morgan Huse – 11/20/25

Trichet et al., 2012

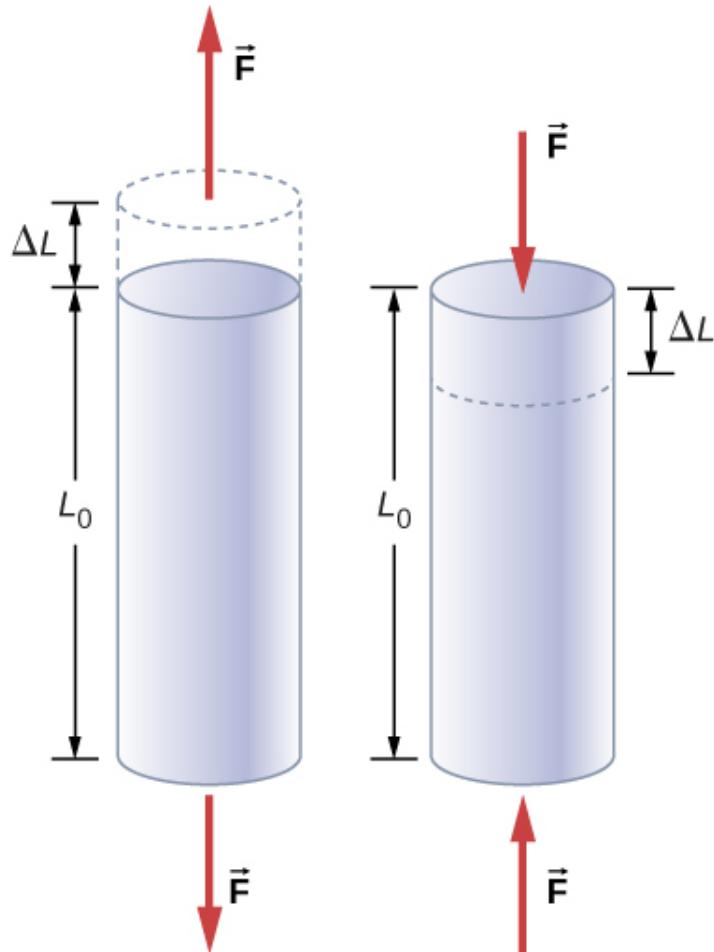
Lecture outline

- 1) The mechanical properties of cells.
- 2) Mechanical force exertion by cells.
- 3) Mechanotransduction: pathways and functions.
- 4) Cell motility.
- 5) Cell extrinsic forces.
- 6) Cell-cell interactions.

Lecture outline

- 1) The mechanical properties of cells.
- 2) Mechanical force exertion by cells.
- 3) Mechanotransduction: pathways and functions.
- 4) Cell motility.
- 5) Cell extrinsic forces.
- 6) Cell-cell interactions.

Stress and strain



Stress (force)
 $\sigma = F/A$

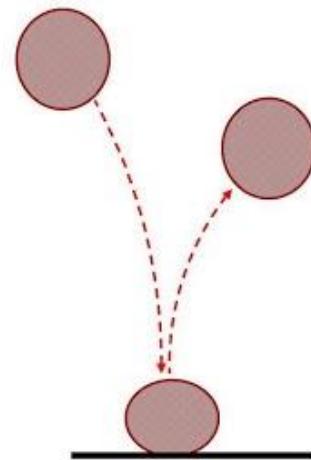
Strain (deformation)
 $\epsilon = \Delta L/L$

Modulus $E = \sigma/\epsilon$

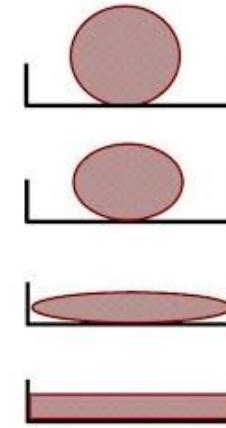
Cells are viscoelastic

Viscous materials resist shear flow linearly with time when stress is applied

Elastic materials strain when stress is applied and immediately return to their original state once stress is removed

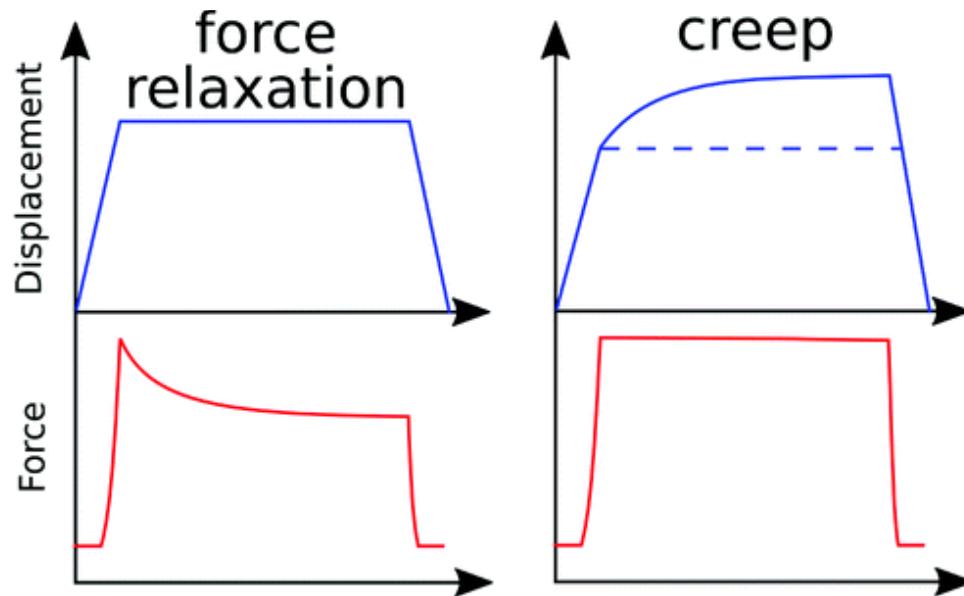


T is short [< 1s]



T is long [24 hours]

<http://www.iq.usp.br/>

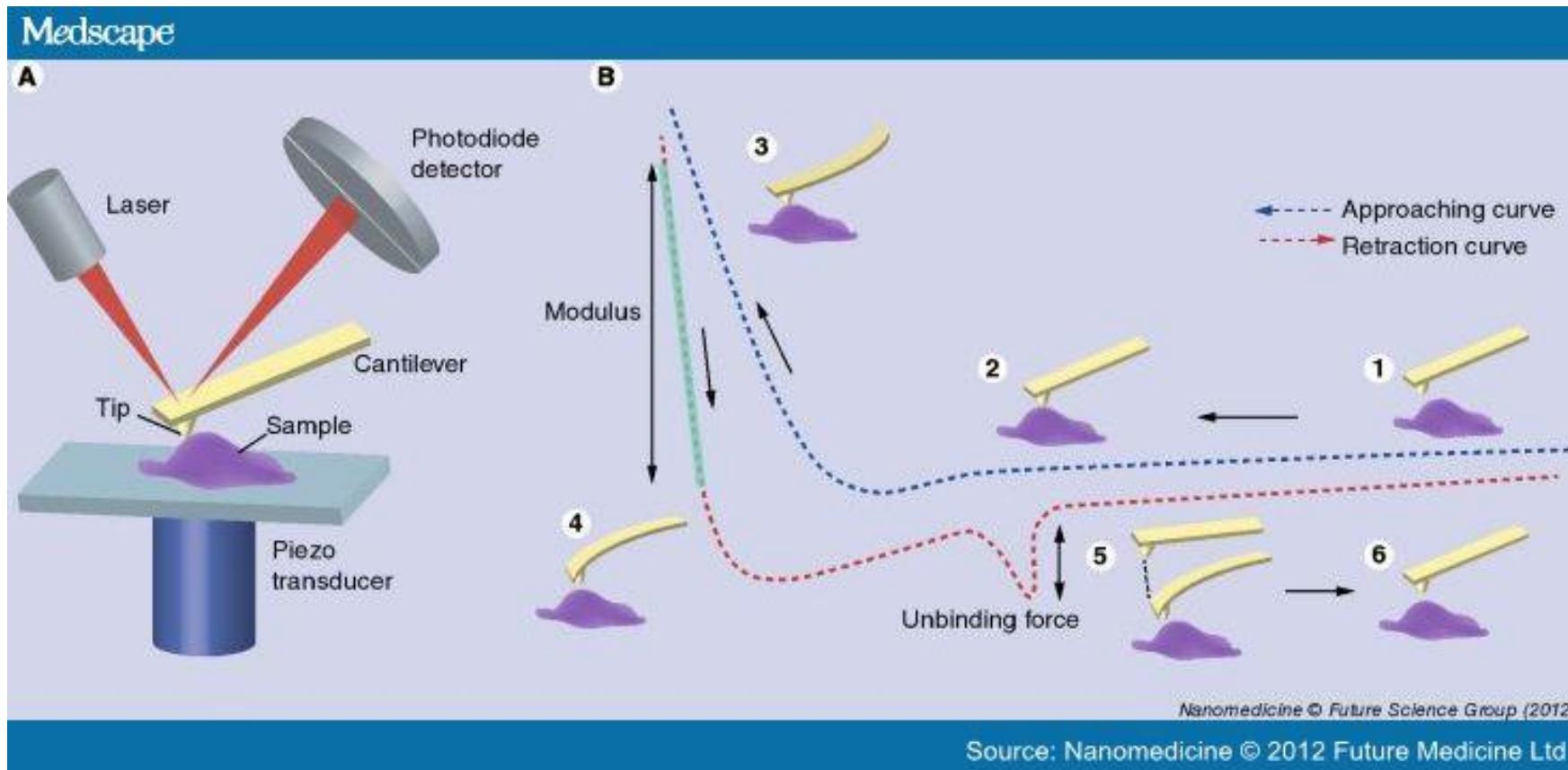


Properties of viscoelastic cells

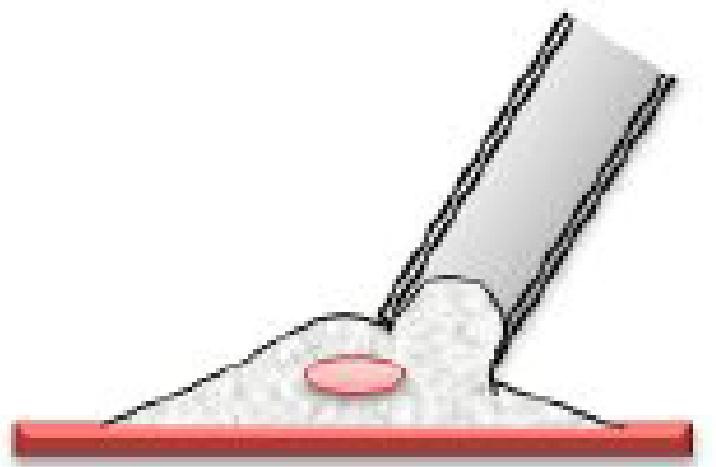
Biophysical properties – things we can measure

Cell Stiffness/rigidity/deformability: Resistance to transient deformation

Atomic force microscopy (AFM)

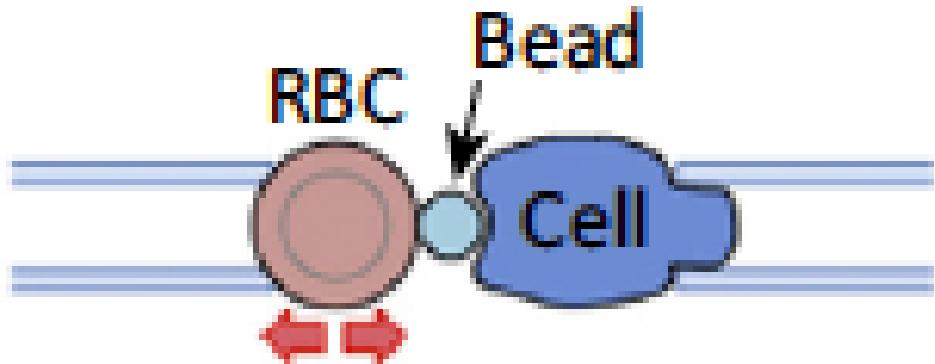


Other ways to measure deformability



Micropipette aspiration

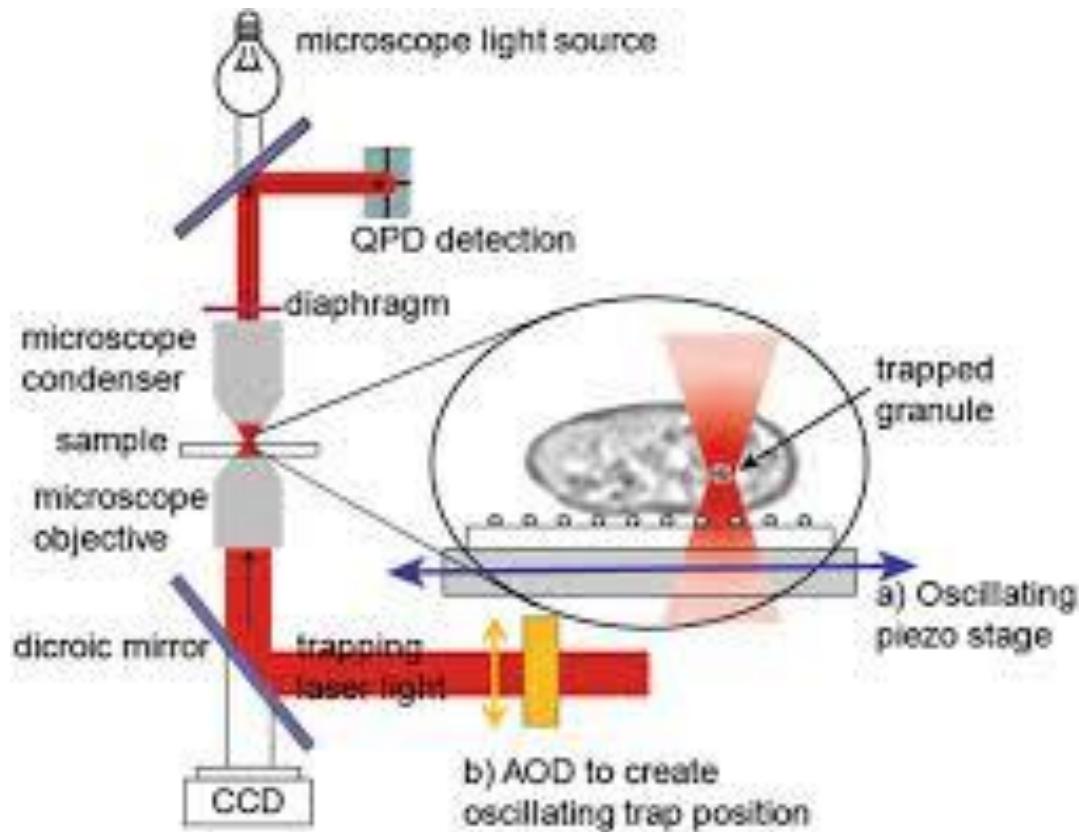
Bhat et al, 2012



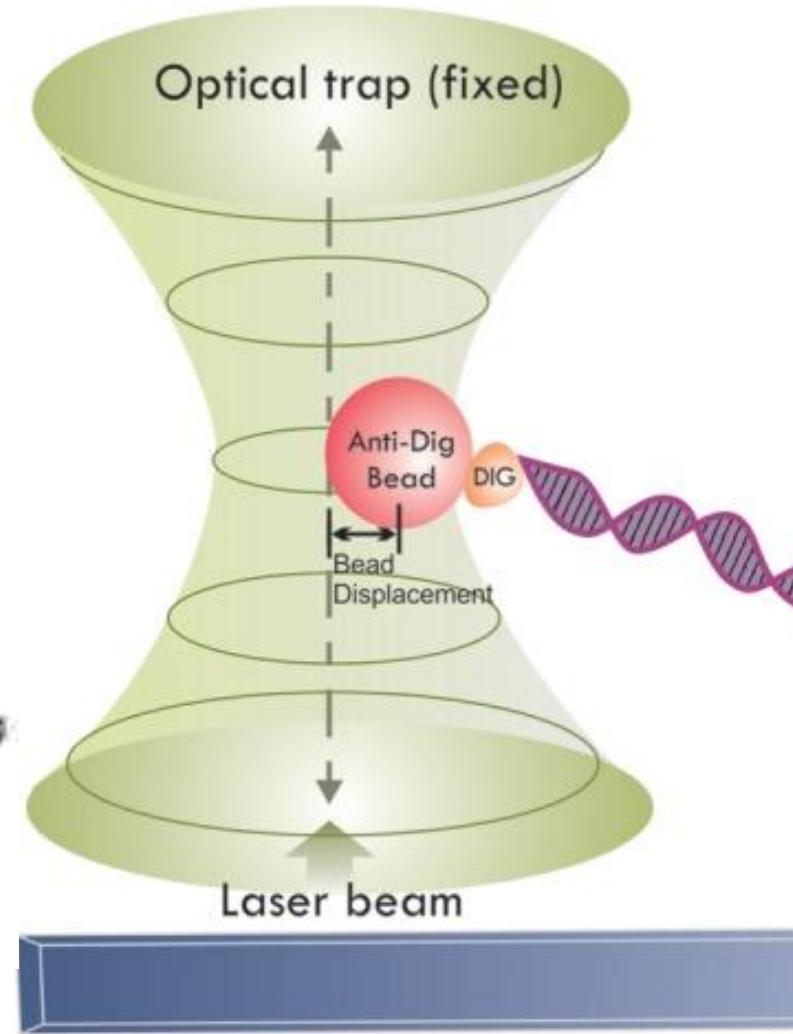
Biomembrane force probe

Basu et al, 2017

Optical trap



Ritter et al, 2017

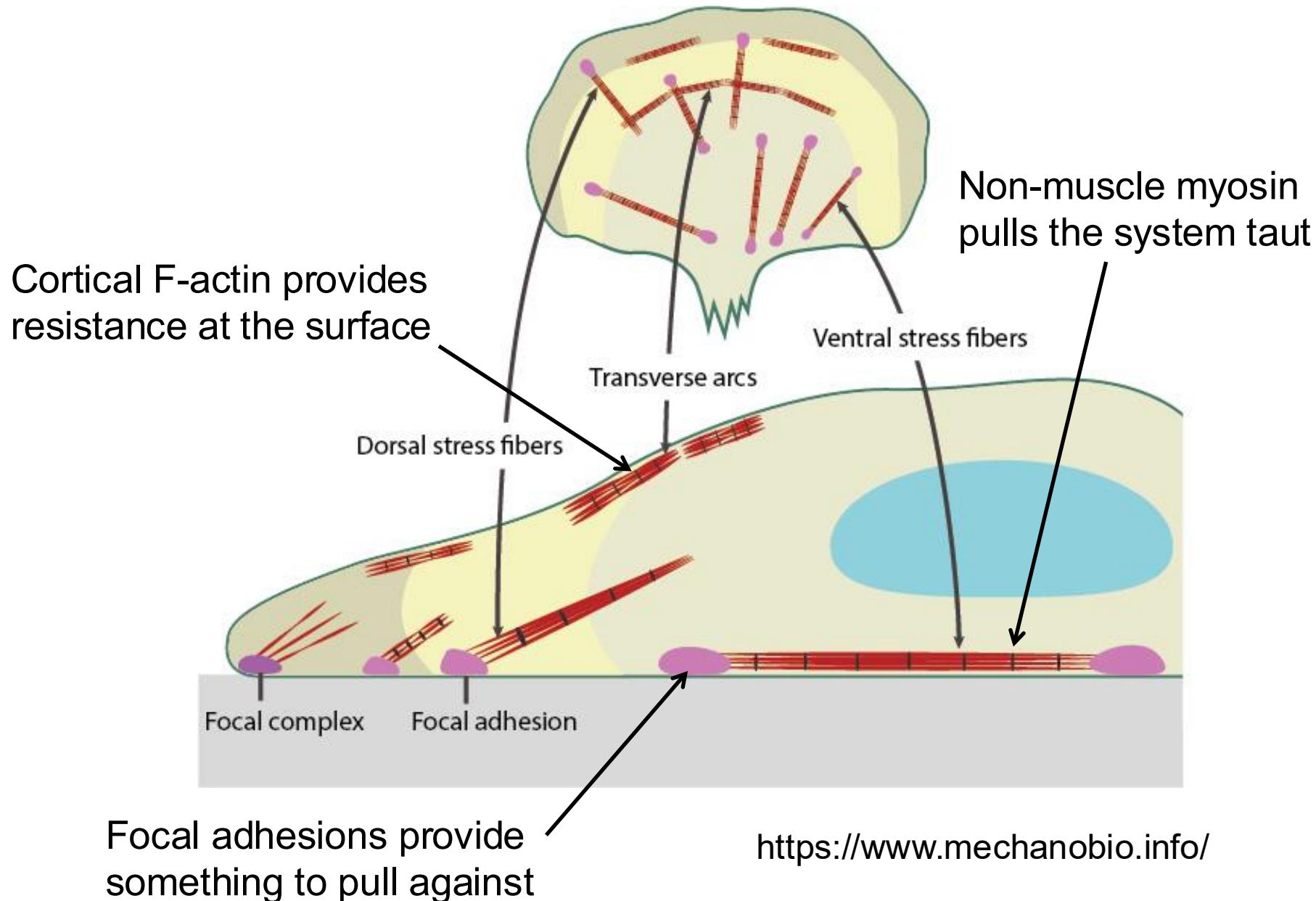


Wang et al, 2013

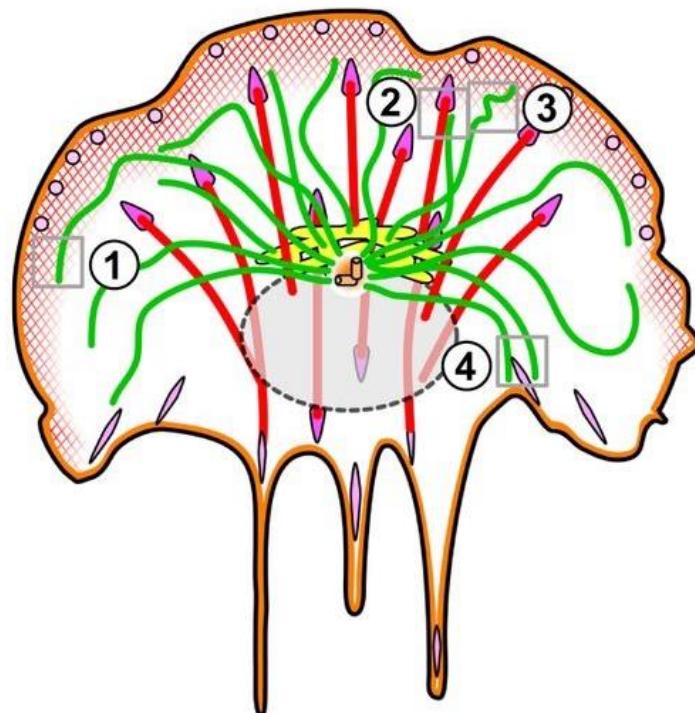
How stiffness is maintained

- 1) Cytoskeleton
- 2) Osmotic pressure
- 3) Cell membrane

Mechanical properties of the cytoskeleton – actomyosin system



Mechanical properties of the cytoskeleton – microtubules

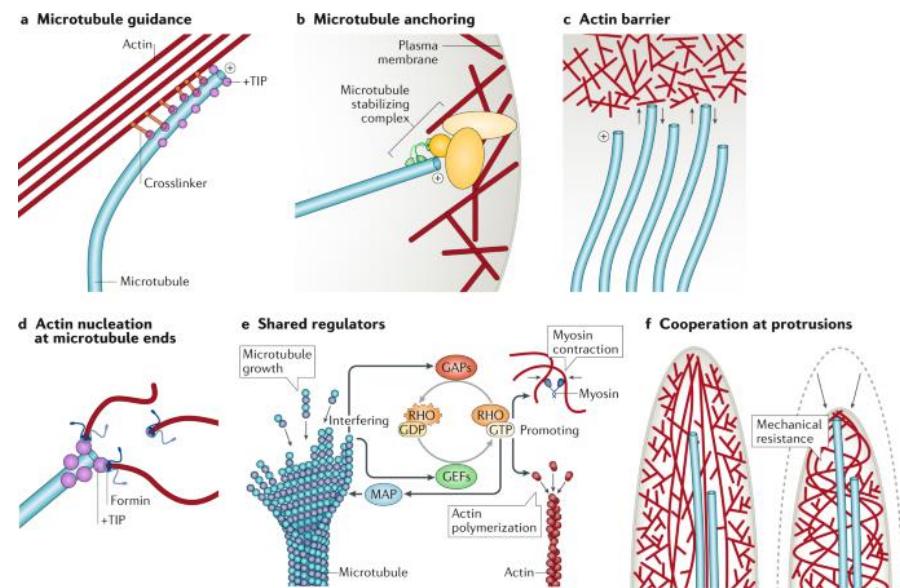


- ①  Retrograde F-actin flow pushes MTs backwards
- ②  FA targeting by MTs: interaction with F-actin and cortical complex
- ③  MT buckling indicates compression
- ④  Trailing adhesion targeting by MTs

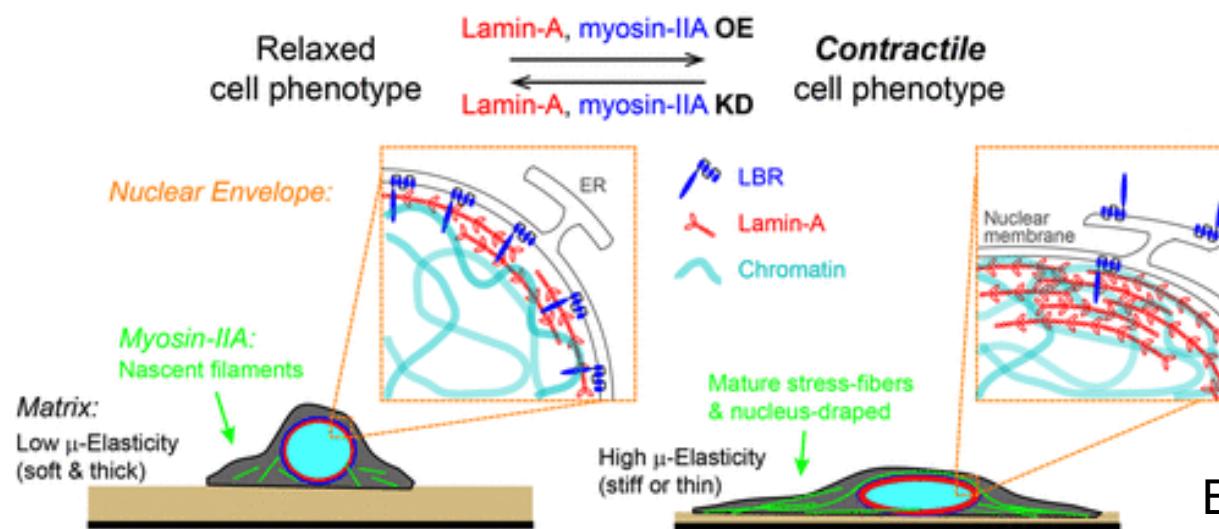
Microtubules resist compression

Provide safe space for cellular organelles

Microtubules are coupled to cortical F-actin in different ways.



Mechanical properties of the cytoskeleton – lamins



Buxboim et al, 2017

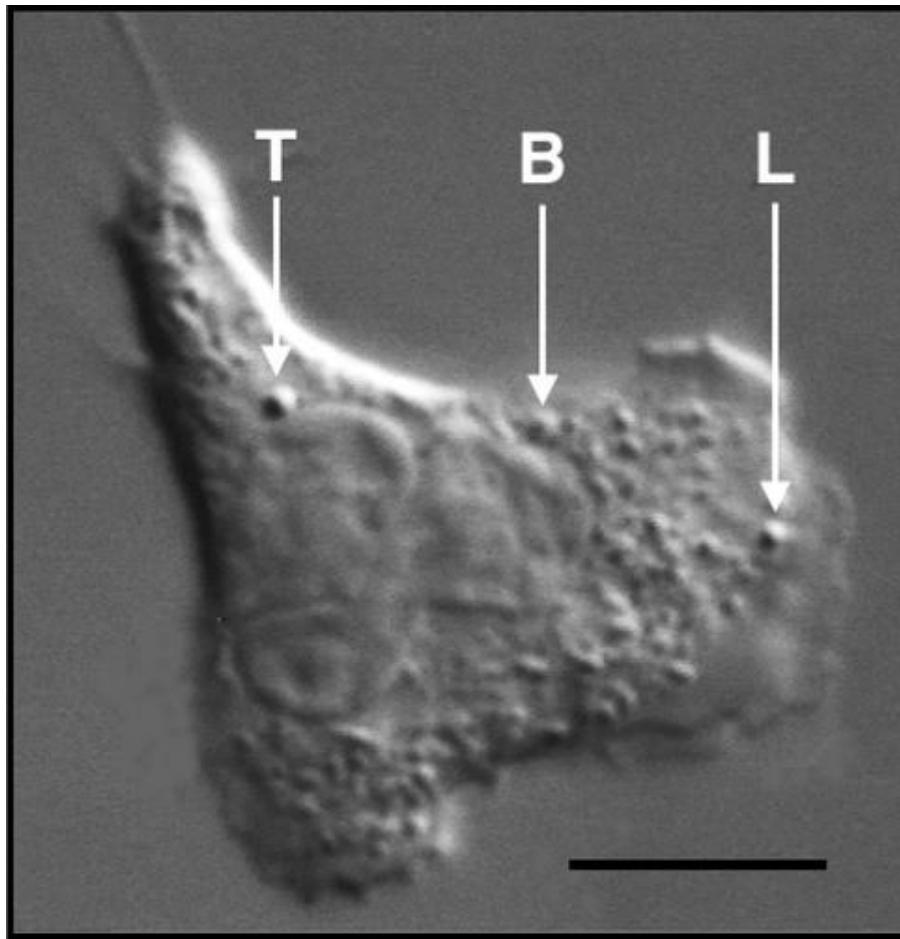
Lamins allow nucleus to resist compression



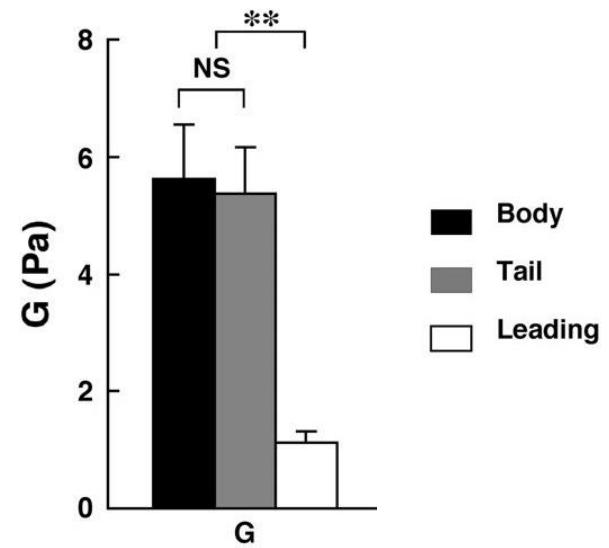
<http://bam.lab.mcgill.ca/>

Lamins are coupled to the cytoplasmic cytoskeleton

Cellular tension is heterogeneous



Tanai et al, 2004

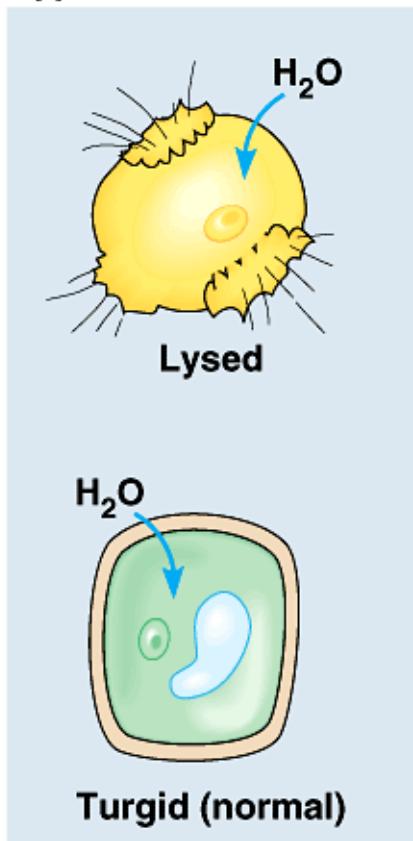


Elastic modulus Stress/strain

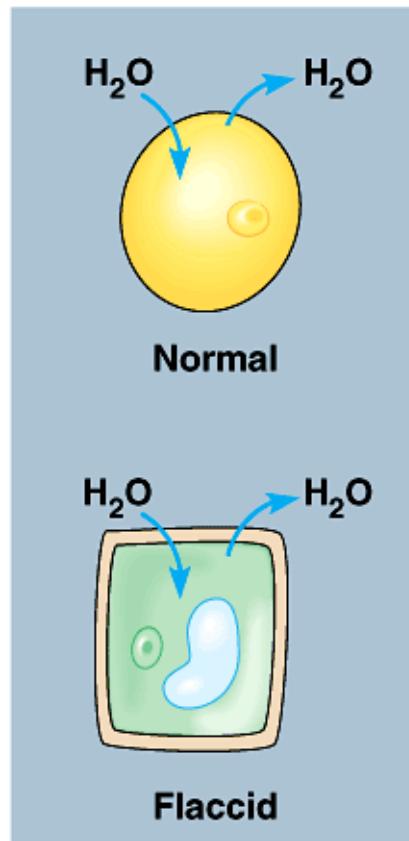
Stress = force per unit area
Strain = deformation

Osmotic pressure

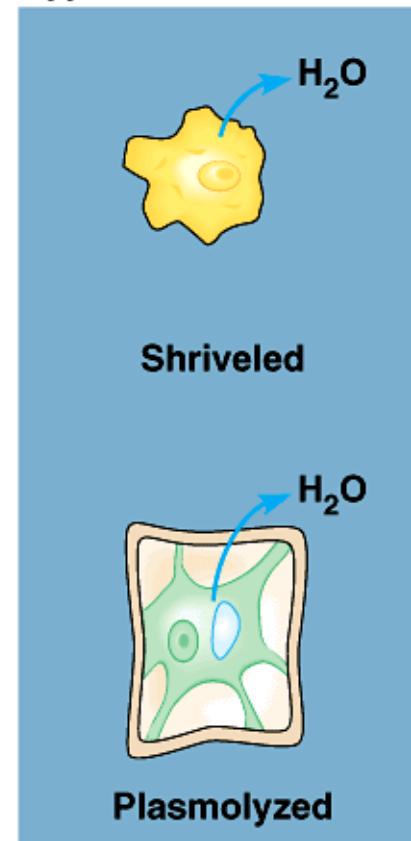
Hypotonic solution



Isotonic solution



Hypertonic solution



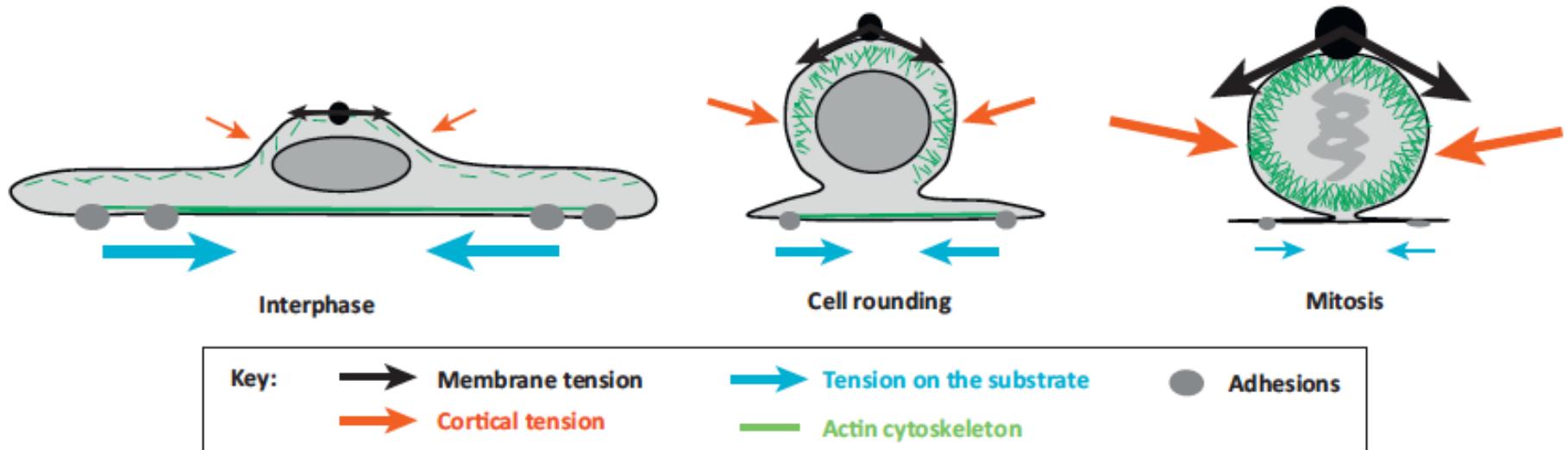
Animal cell

Plant cell

Copyright © Pearson Education, Inc., publishing as Benjamin Cummings.

Osmotic pressure is actively maintained by ion transport.
Cells will adjust to changes in tonicity

Membrane tension



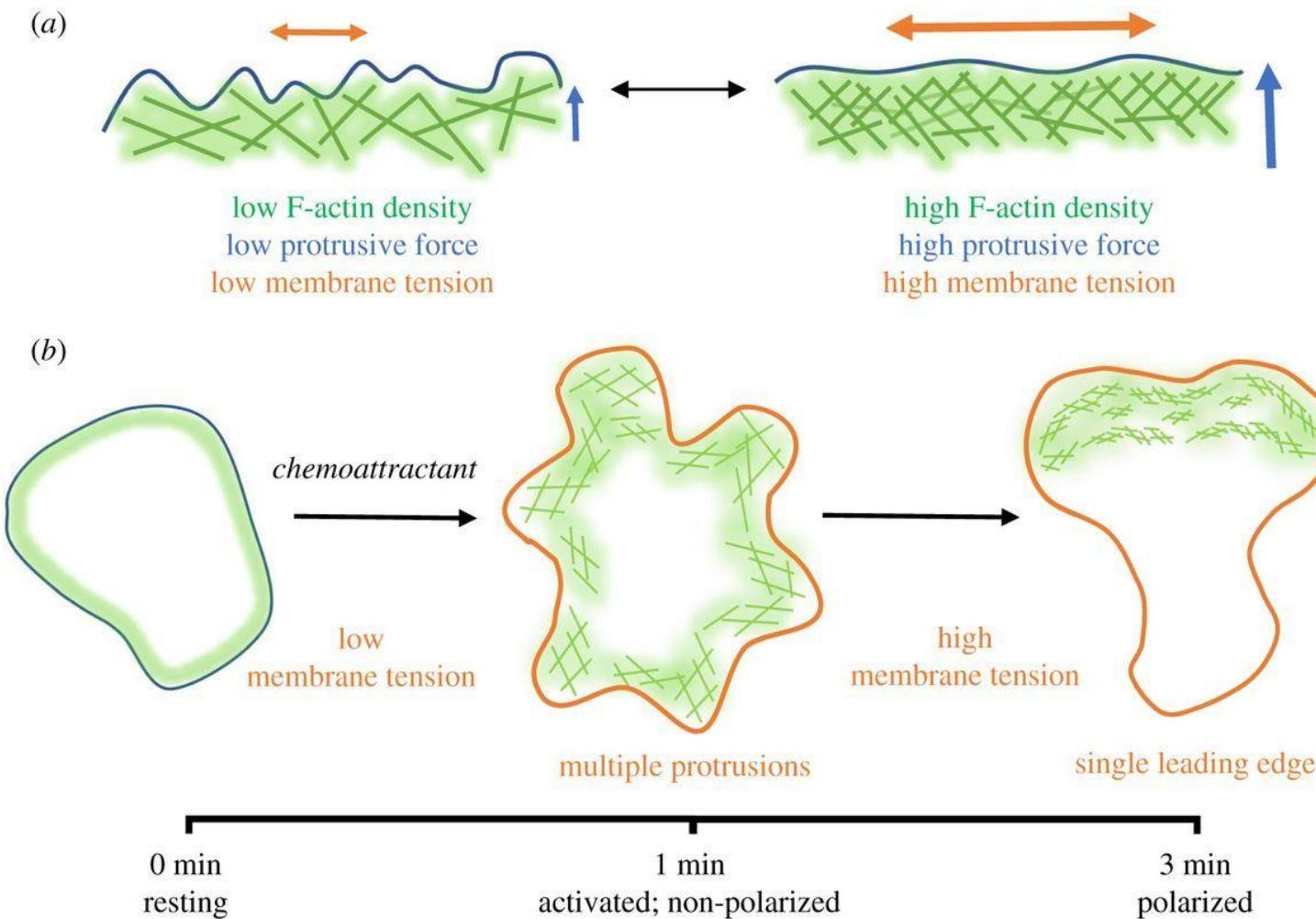
TRENDS in Cell Biology

Gauthier et al, 2012

Membrane tension =
in plane tension (osmotic pressure) + membrane-cytoskeleton adhesion

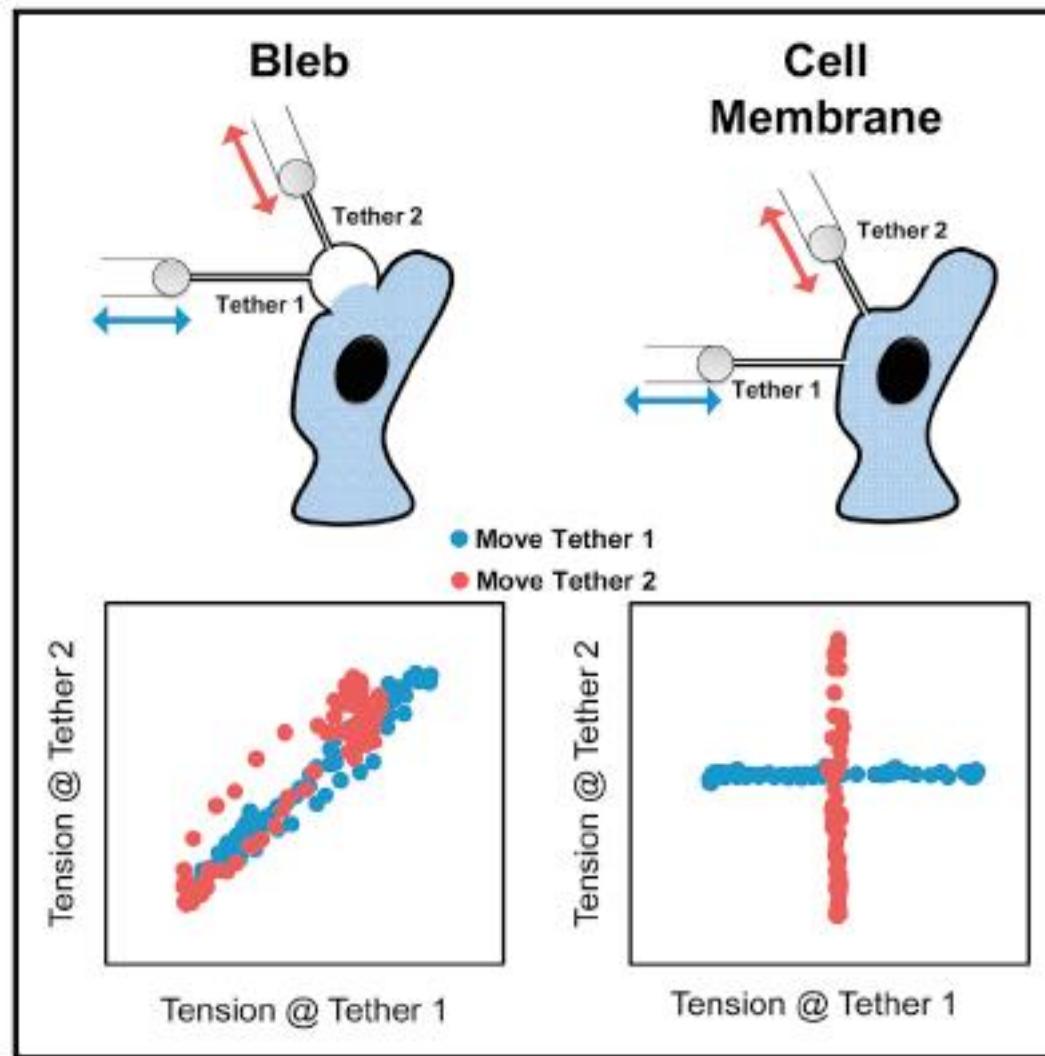
Initially thought to be uniform across the cell surface because tension distributes quickly across bilayers

Membrane tension - a long distance regulator?



Membrane tension

However . . .



Shi et al, 2018

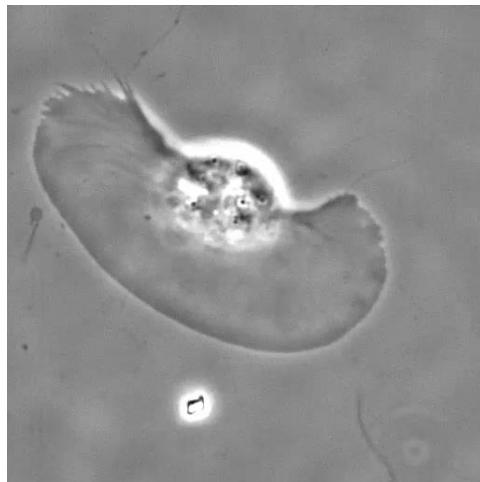
Lecture outline

- 1) The mechanical properties of cells.
- 2) Mechanical force exertion by cells.
- 3) Mechanotransduction: pathways and functions.
- 4) Cell motility.
- 5) Cell extrinsic forces.
- 6) Cell-cell interactions.

The dynamic cell

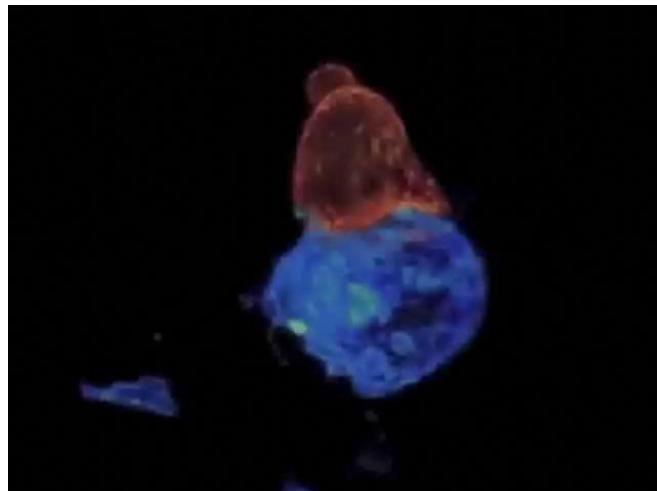
Cells also move – these movements necessarily transmit forces

Fish keratocyte



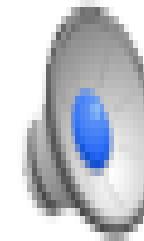
Barnhart et al, 2010

T cell – tumor cell



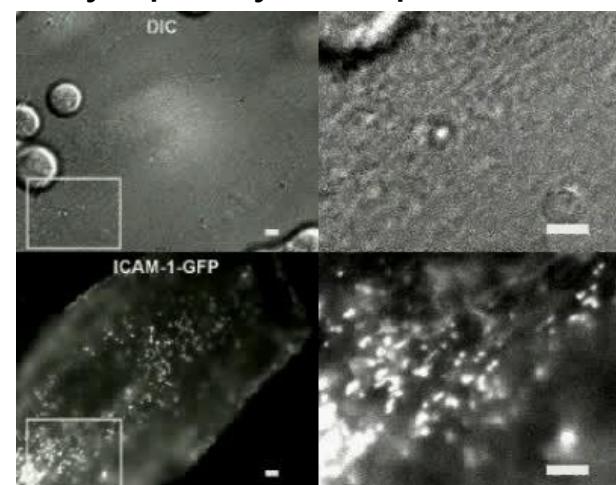
Ritter et al, 2015

Drosophila embryo



Fernandez-Gonzalez et al, 2009

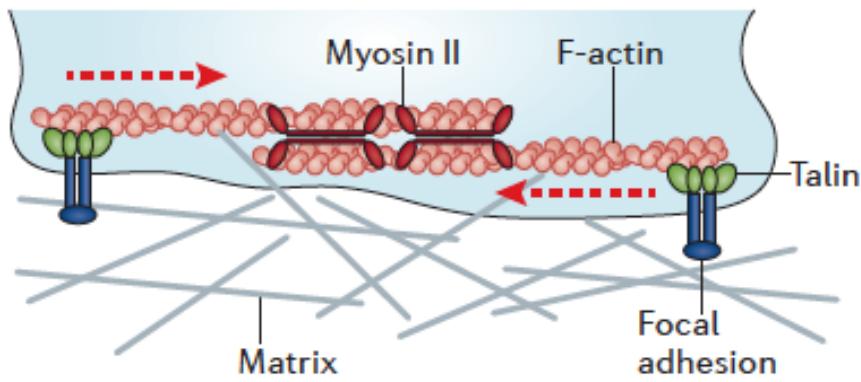
Lymphocyte diapedesis



Carman et al, 2007

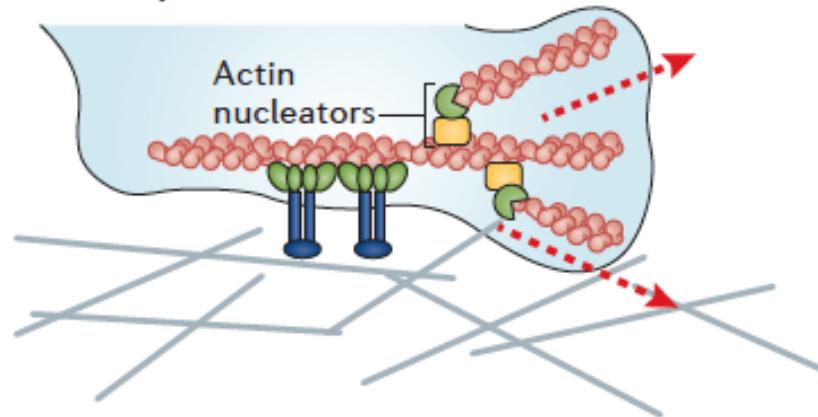
How cells generate force – the F-actin cytoskeleton

a Myosin contractility



Contractile force

b Actin protrusion



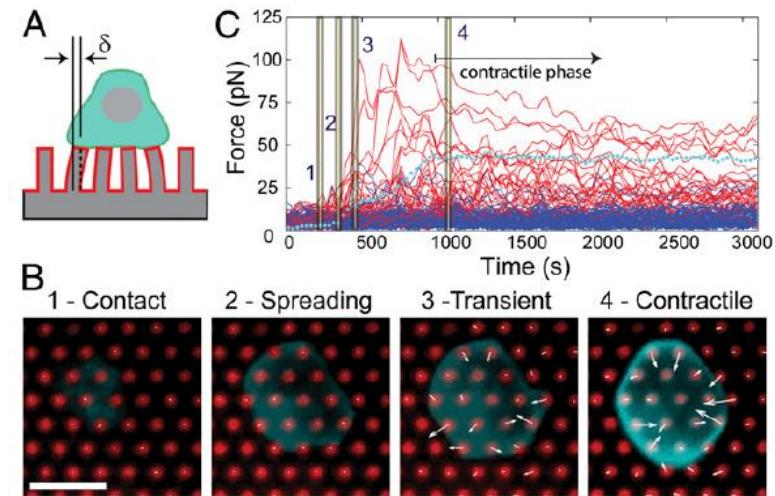
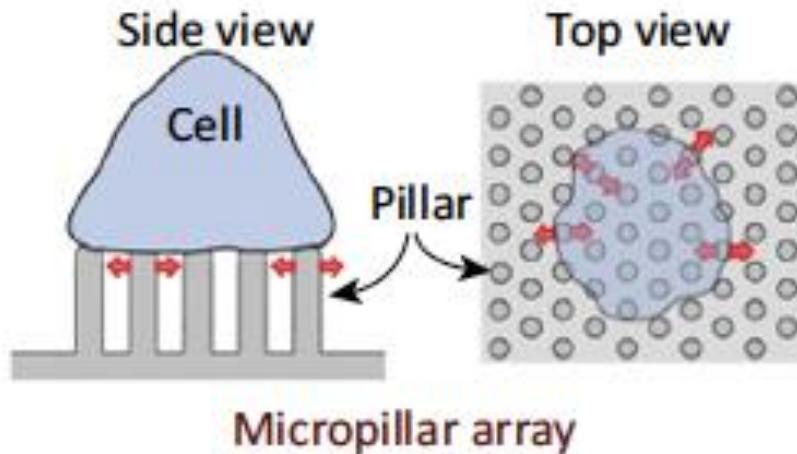
Protrusive force

Huse, 2017

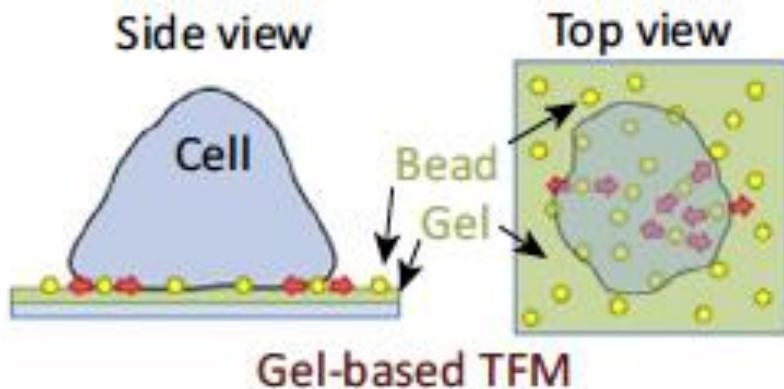
Useful for motility, manipulation of other cells, signaling

Measuring cellular forces

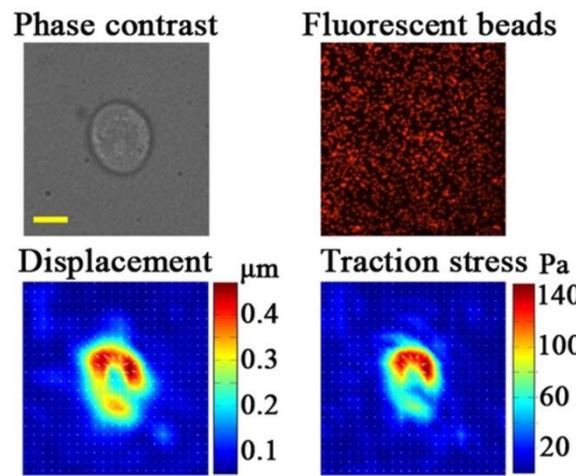
Traction force microscopy



Bashour et al., 2014



Basu and Huse, 2017



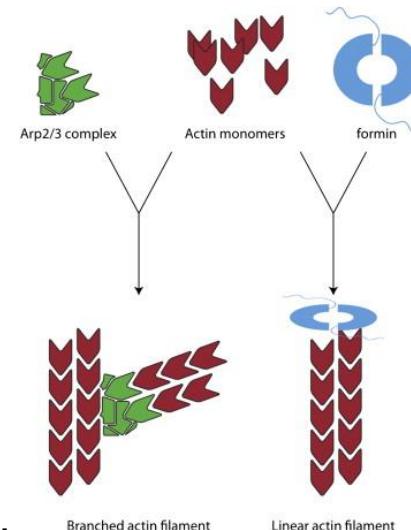
Wang et al., 2018

Components – F-actin polymerization

Actin polymerization drives cellular protrusion

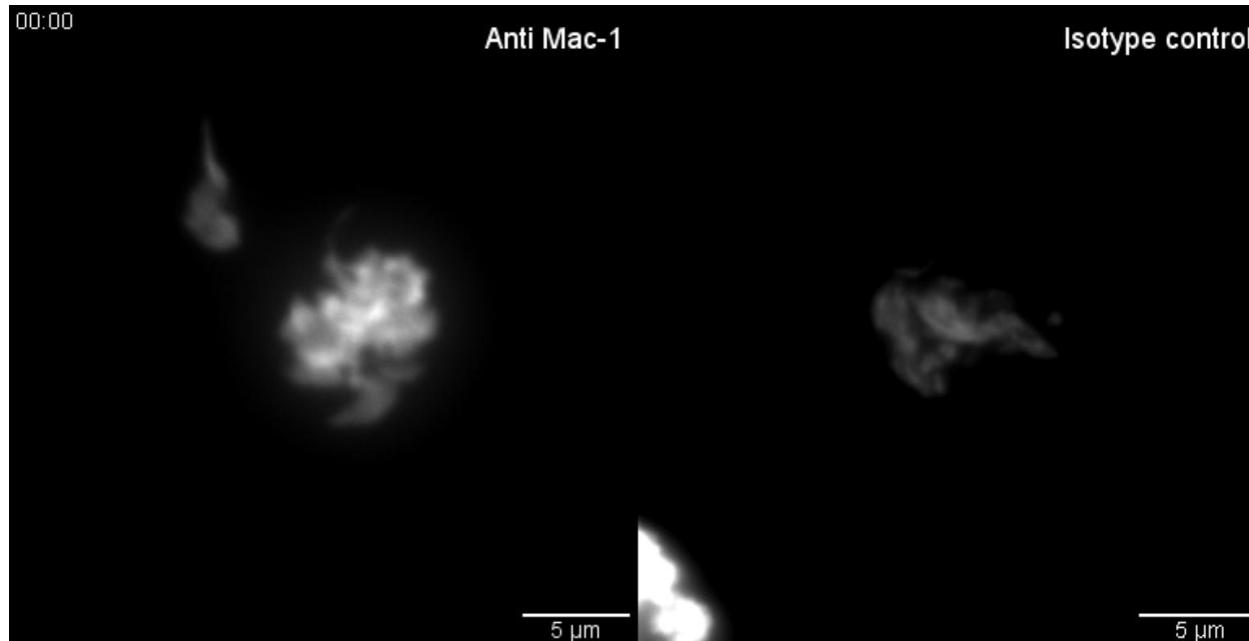
Arp2/3 complex generates branched arrays.

Formins induce linear arrays and bundles.



Protrusion generally coupled to retraction - treadmilling

Insall and Machesky, 2009



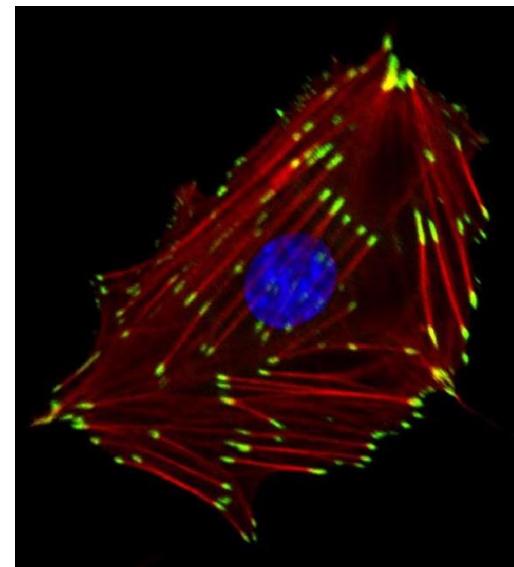
Jaumouille et al., 2019

Components – Focal adhesions

An in vitro phenomenon.

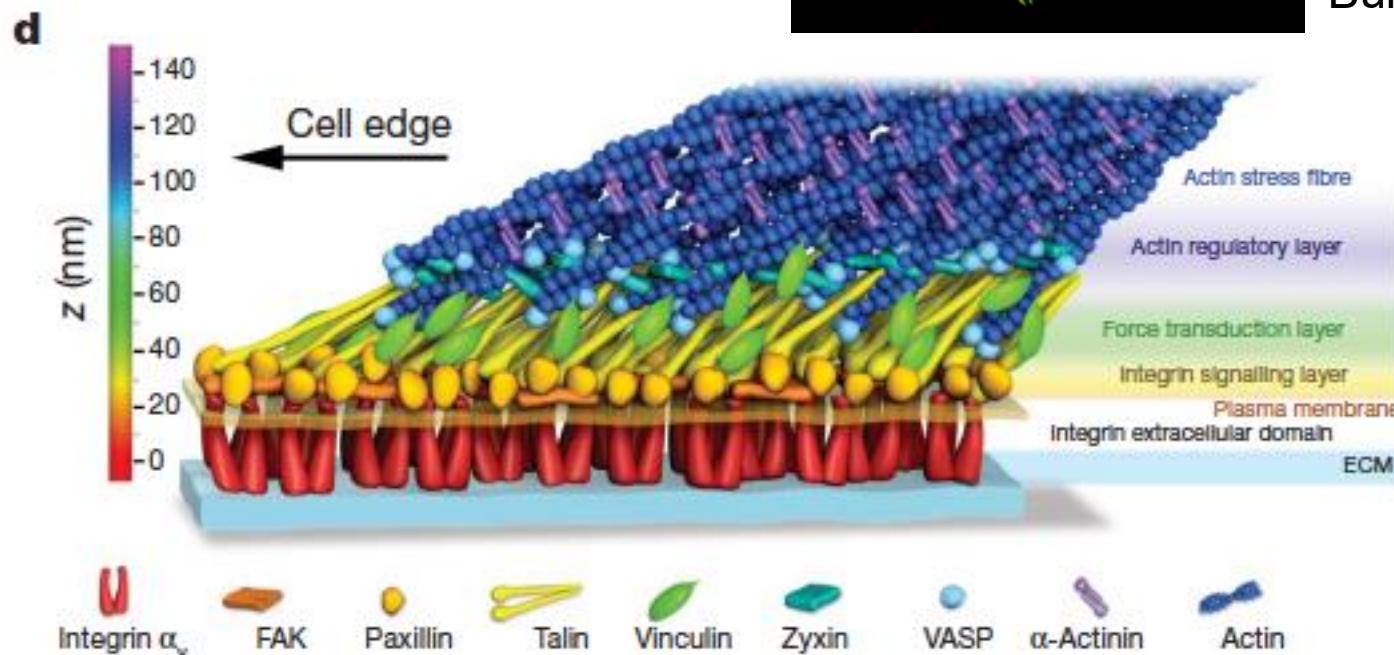
However, adhesion-like structures are observed in vivo.

A source of signals in addition to adhesive strength.



F-actin
Vinculin

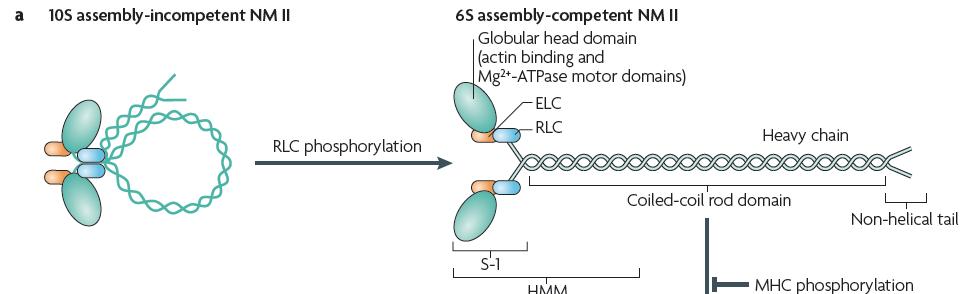
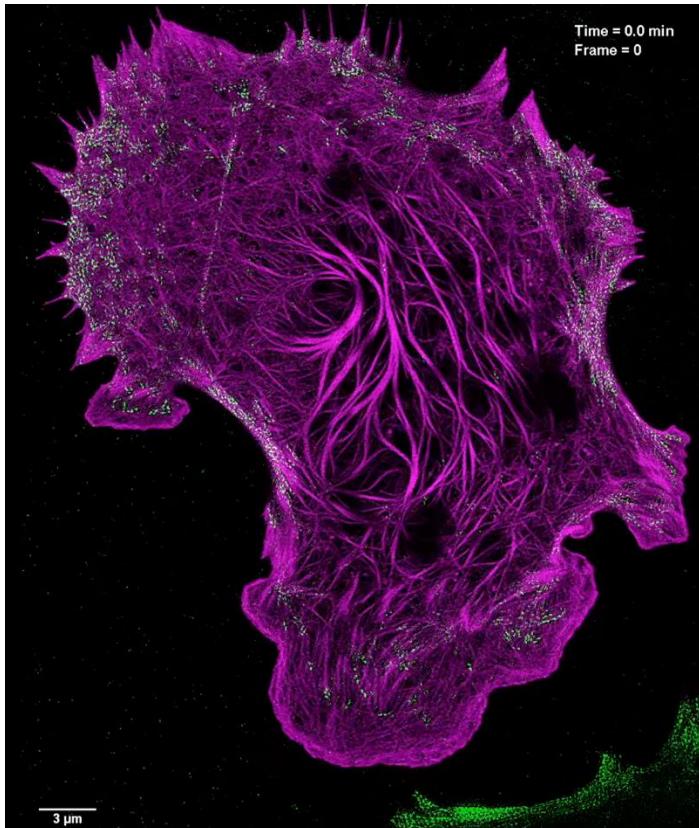
Burridge, 2017



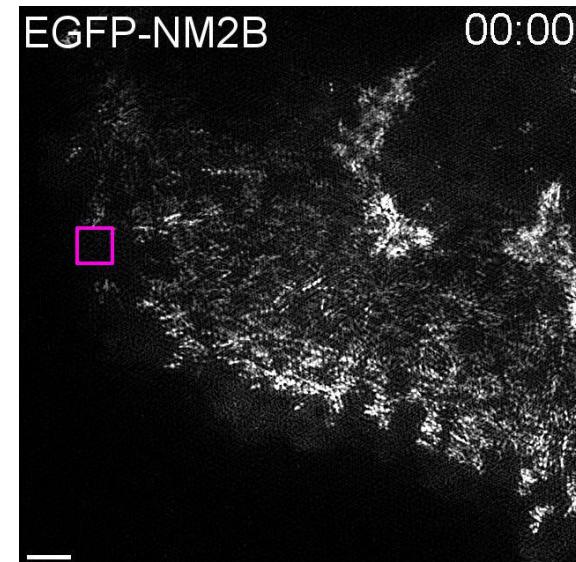
Kanchanawong et al., 2010

Components – Myosin

Analogous to role in skeletal muscle.



Vincente-Manzanares
et al., 2009



Beach et al., 2017

Lecture outline

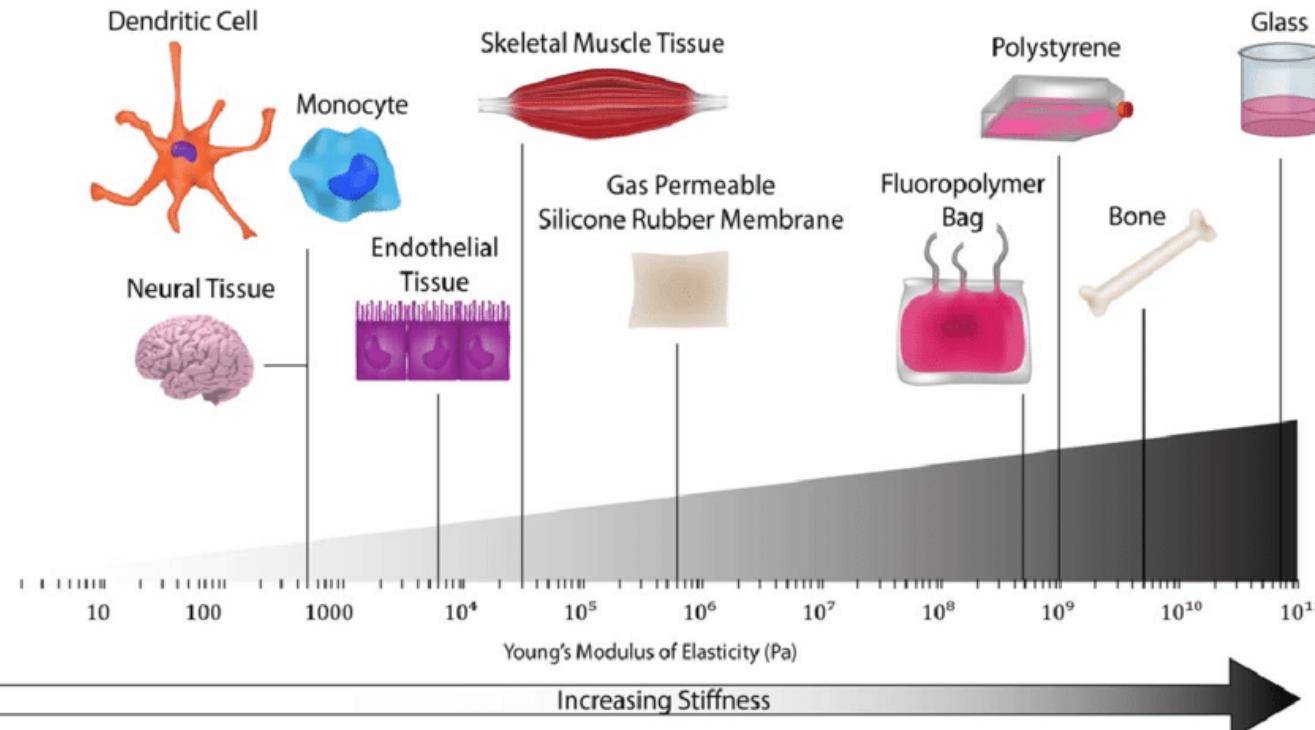
- 1) The mechanical properties of cells.
- 2) Mechanical force exertion by cells.
- 3) **Mechanotransduction: pathways and functions.**
- 4) Cell motility.
- 5) Cell extrinsic forces.
- 6) Cell-cell interactions.

Mechanobiology and signal transduction

Mechanosensing: Cellular responsiveness to mechanical stimuli

Mechanotransduction: Conversion of mechanical stimulus into chemical signal

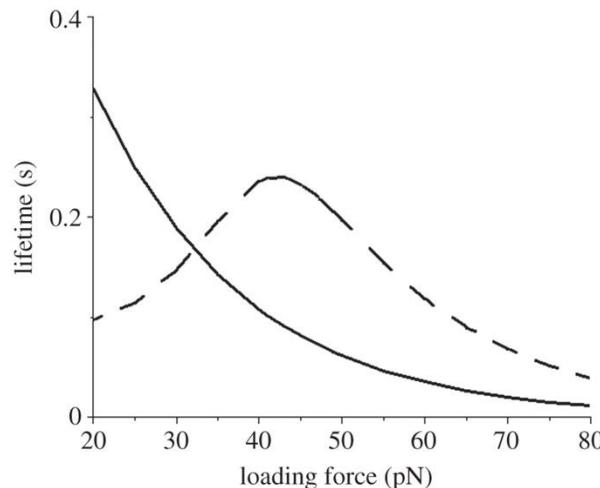
Enables cells to acquire mechanical information about their environment and respond accordingly. Important because mechanical environments *in vivo* vary widely.



Mechanotransduction – ligand binding by integrins

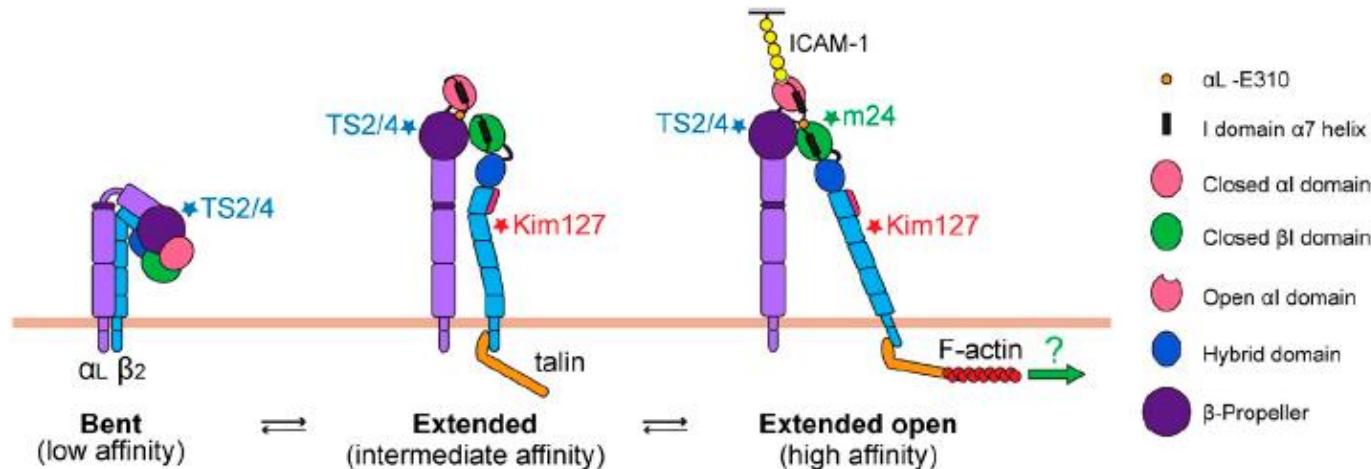
Catch bond – strength increases until optimal applied force.

Slip bond – strength decreased under applied force.



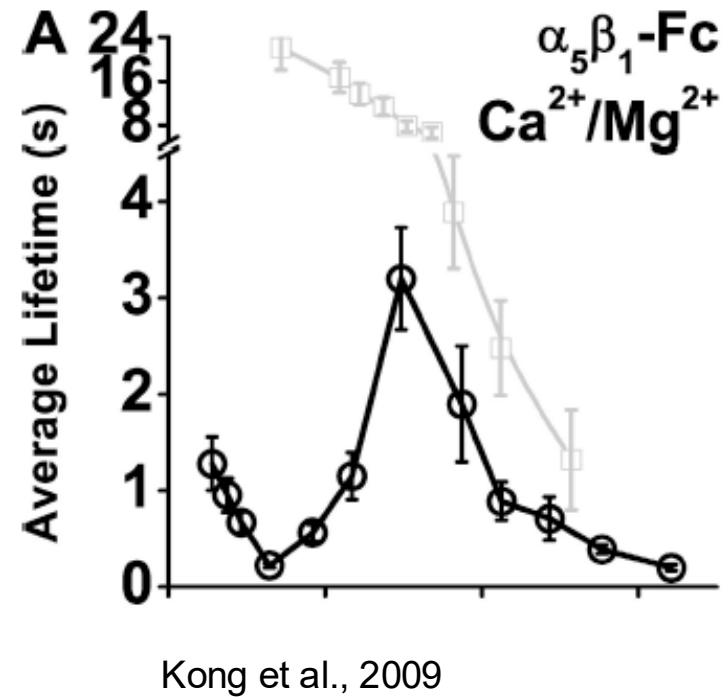
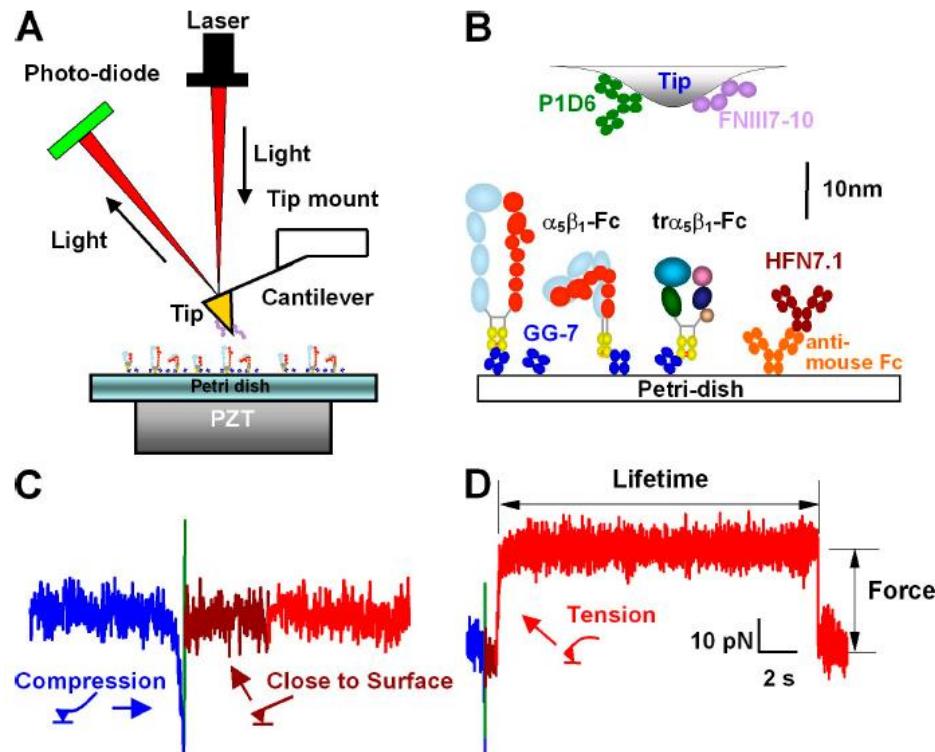
Sun et al., 2011

LFA1 forms catch bonds with ICAM



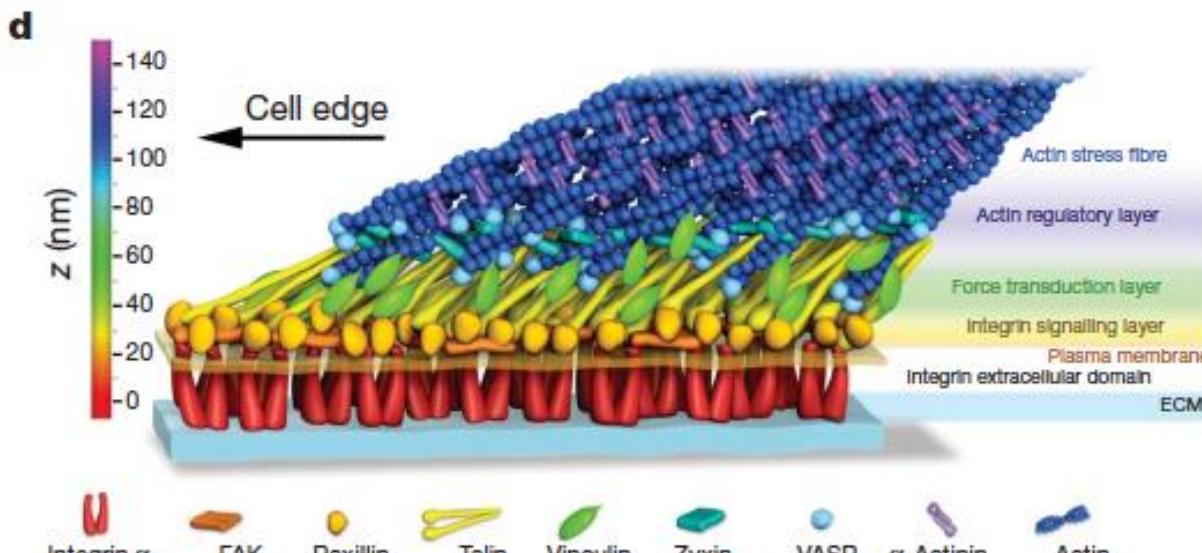
Comrie et al., 2015

Mechanotransduction – ligand binding by integrins



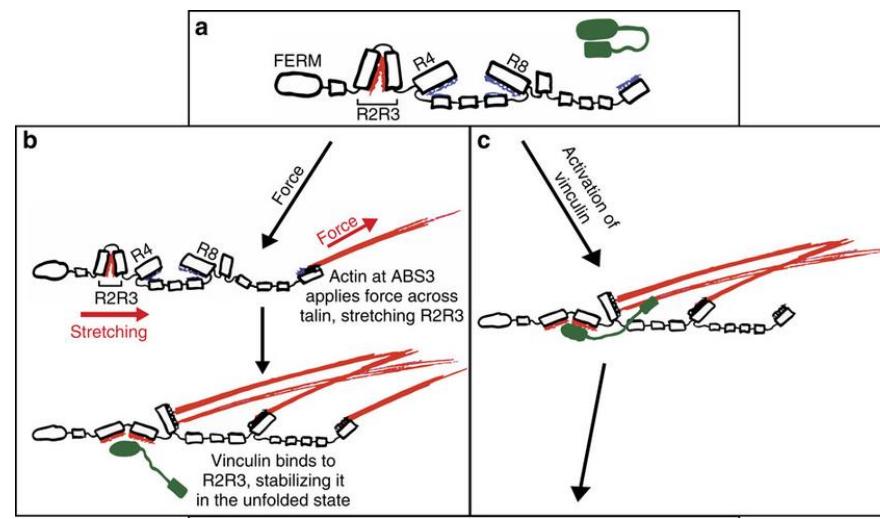
Mechanotransduction – adhesion assembly

Focal adhesion architecture



Kanchanawong et al., 2010

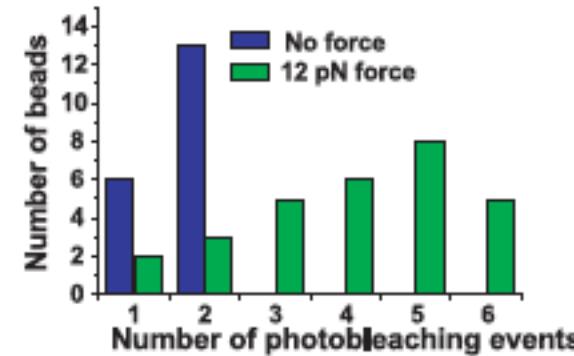
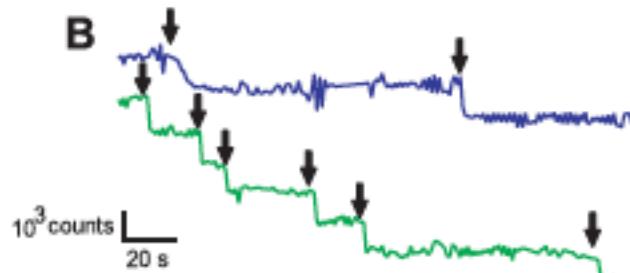
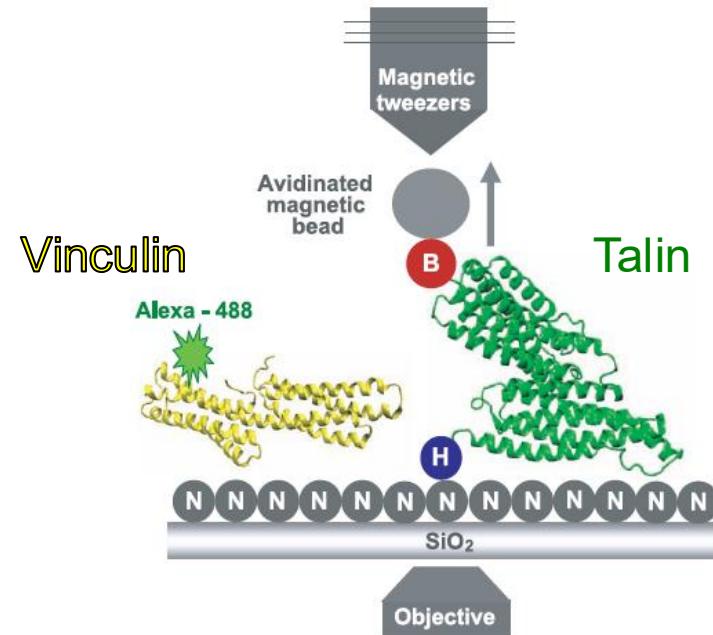
Talin and other adhesion proteins undergo force dependent transitions



Mechanotransduction – adhesion assembly

Optical trap set up with purified proteins.

Measure binding by photobleaching of bound vinculin.

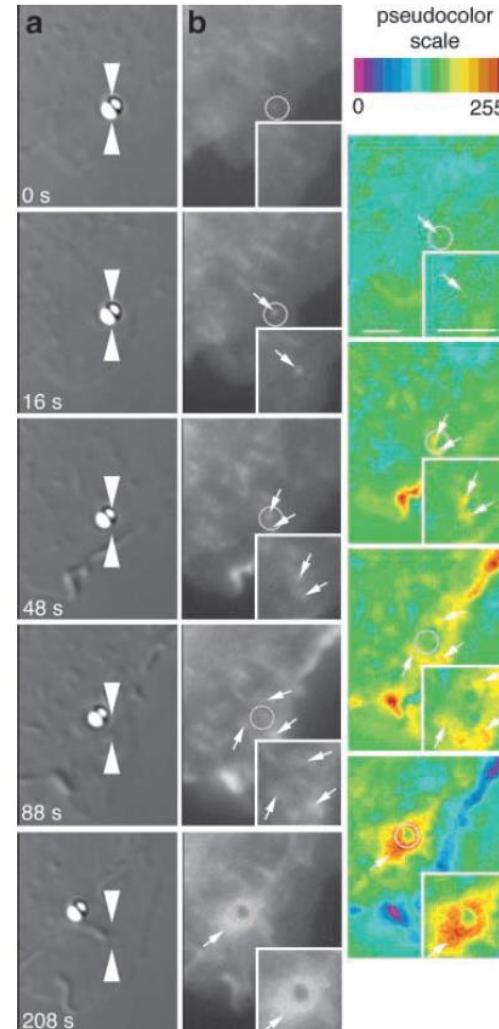


Mechanotransduction – adhesion size

Adhesions themselves are force dependent. Grow in size with pulling force.

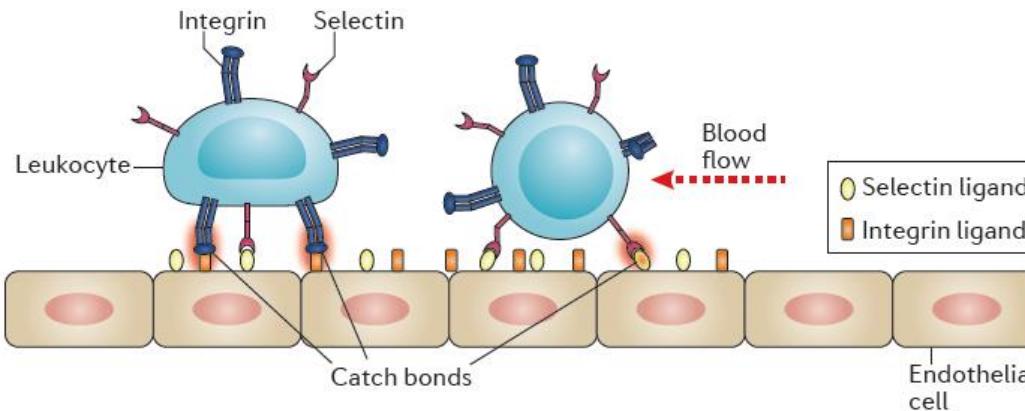
Fibronectin covered bead, placed into contact with a cell, pulled using optical trap.

GFP-vinculin as adhesion probe.

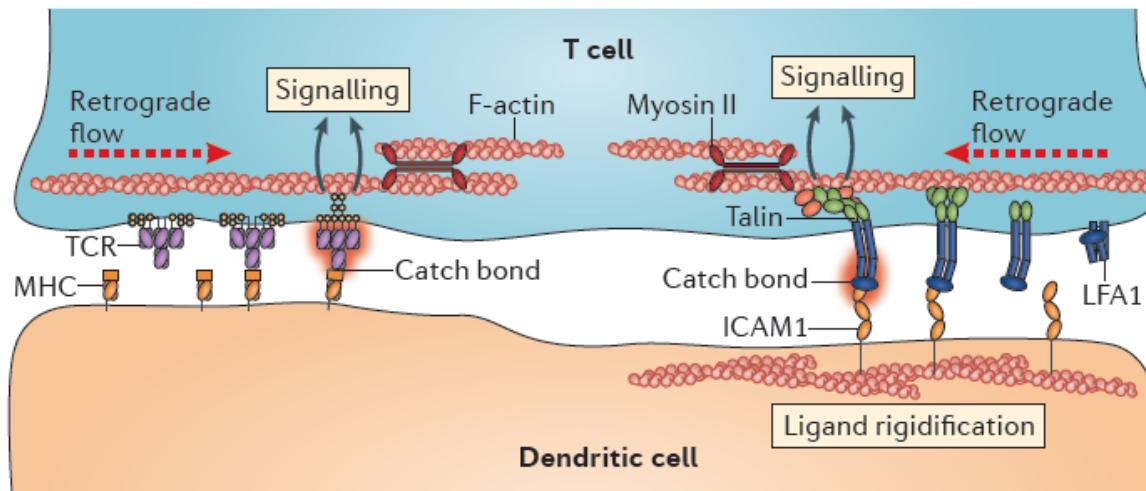


Other mechanosensitive catch bond receptors

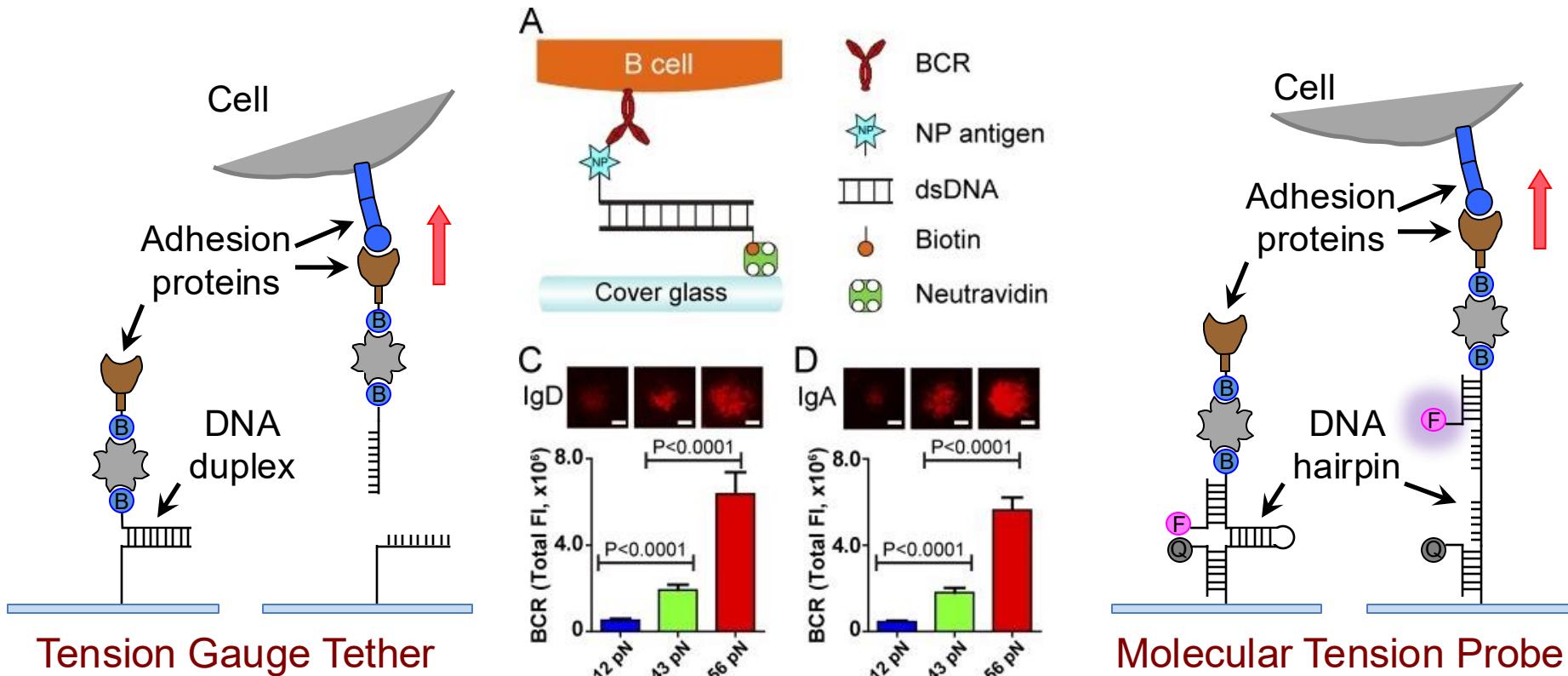
Selectins during leukocyte rolling



Antigen receptors in immune synapse

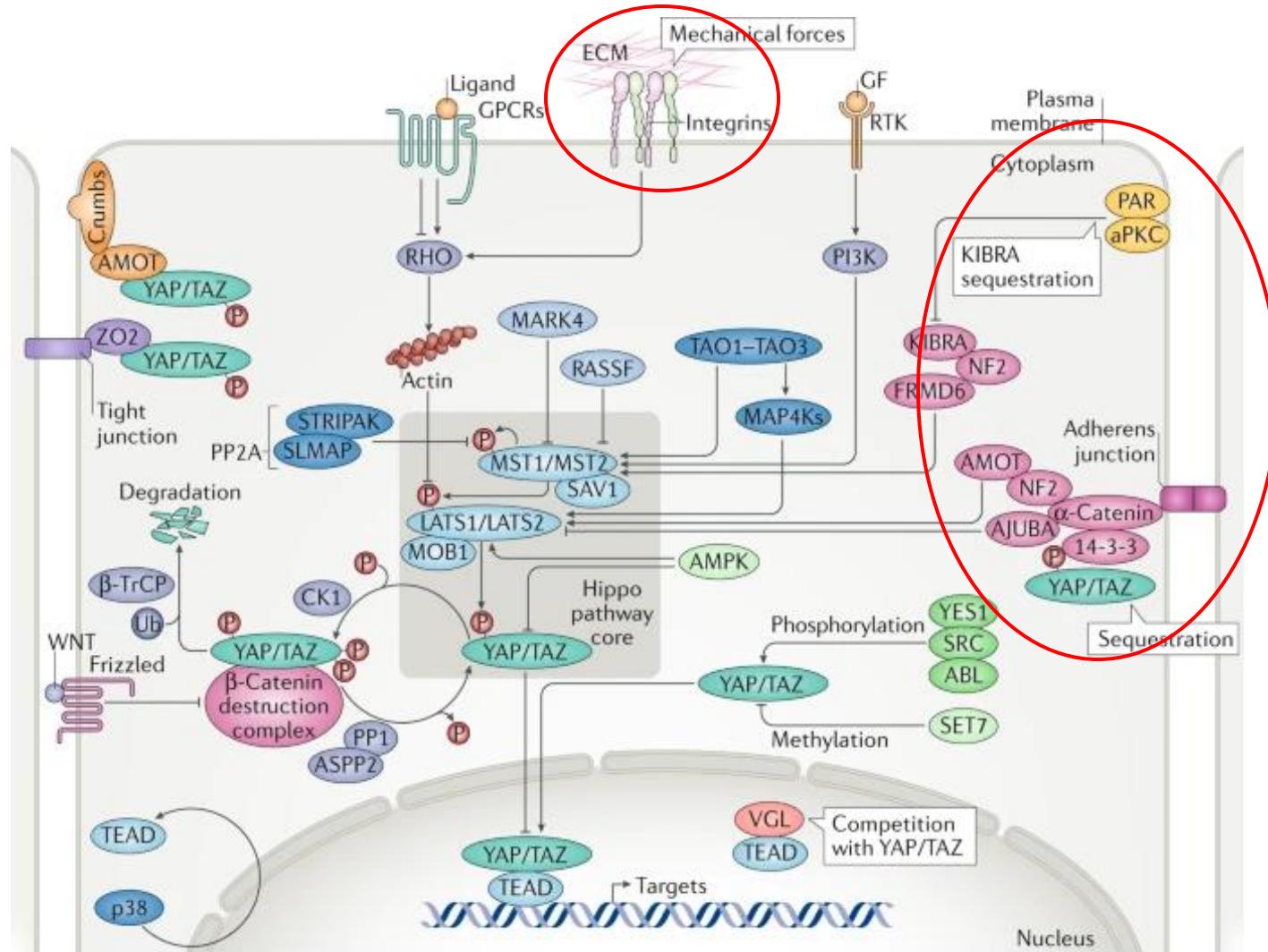


Sustained force required for full receptor activation

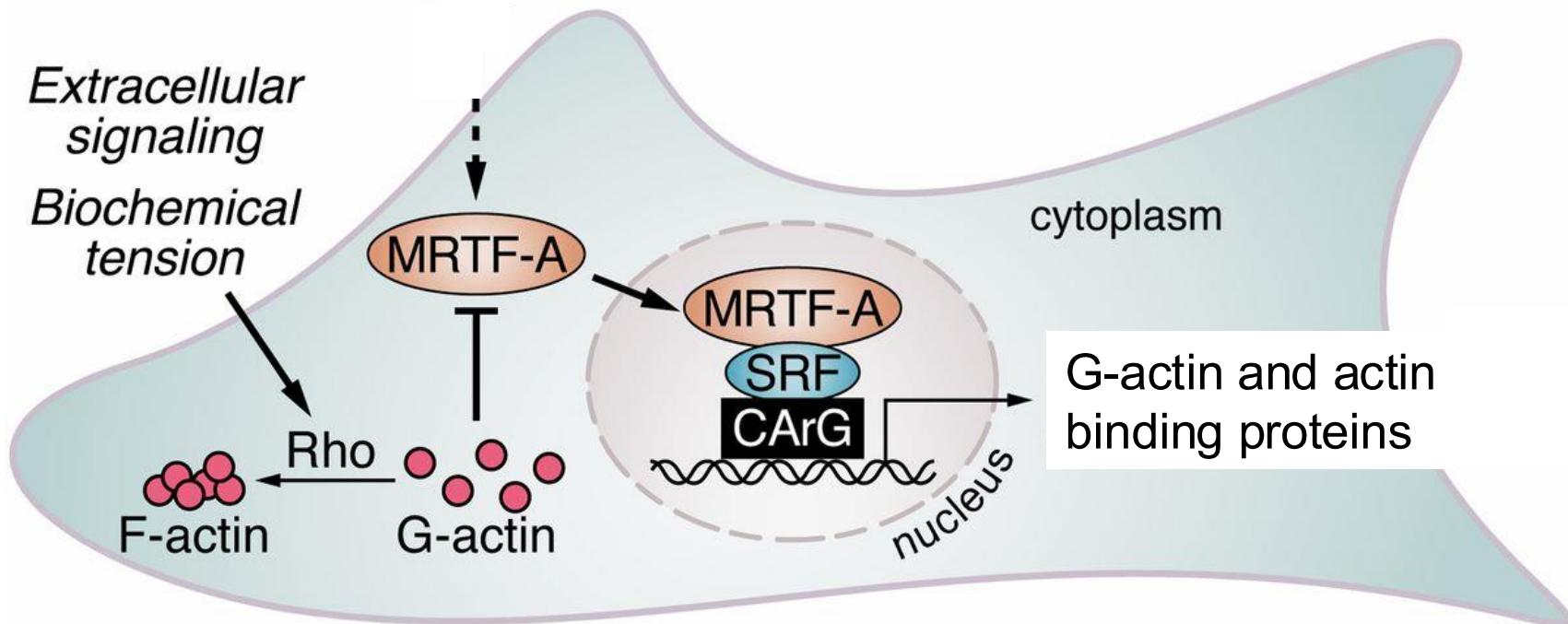


Mechanotransduction to the nucleus – YAP/TAZ

Different adhesions, different results



Mechanotransduction to the nucleus – MRTF

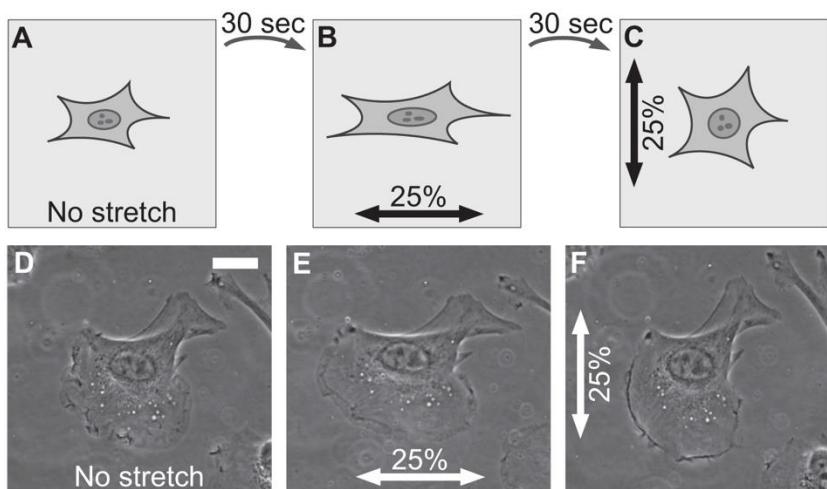


Velasquez et al., 2013

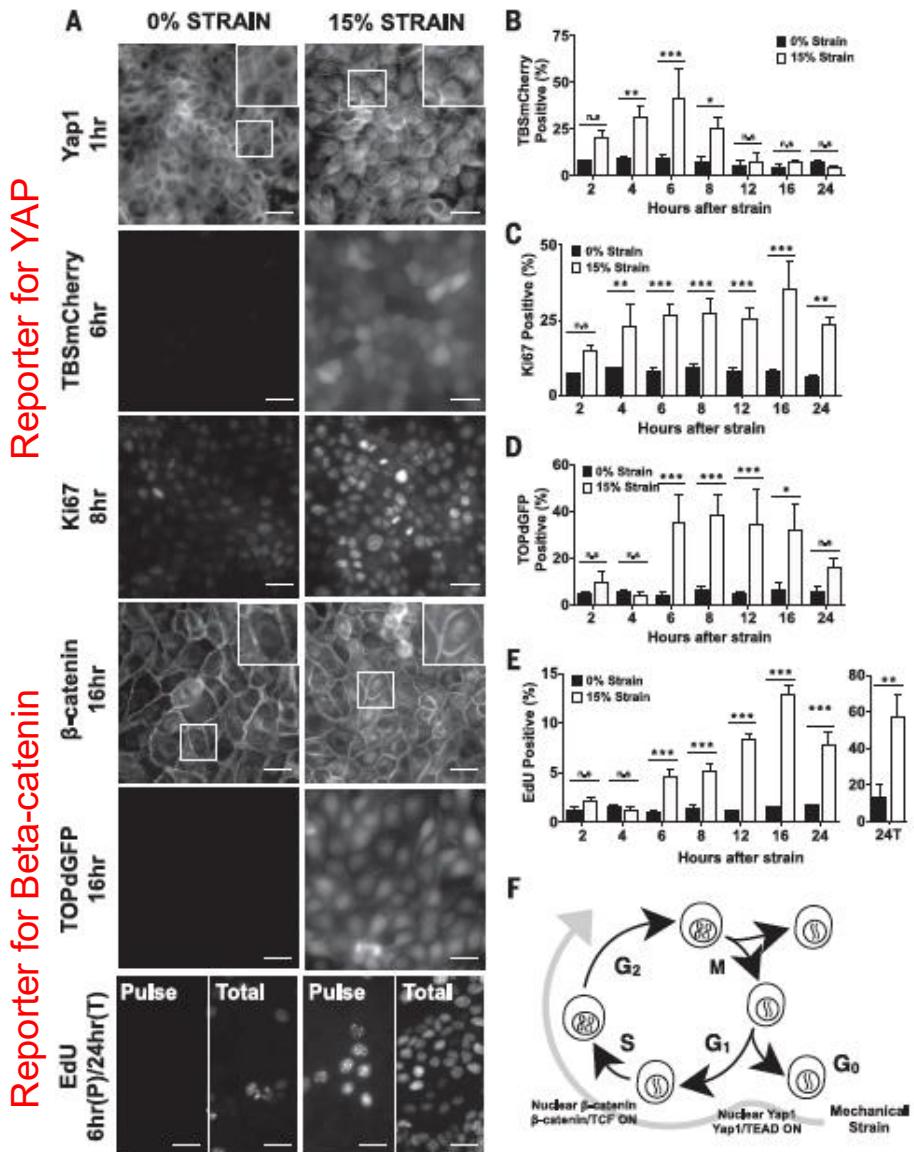
Biological consequences of mechanotransduction

Epithelial sheets on stretchable substrates.

Stretch promotes proliferation via YAP and β -catenin.



Tremblay et al, 2014



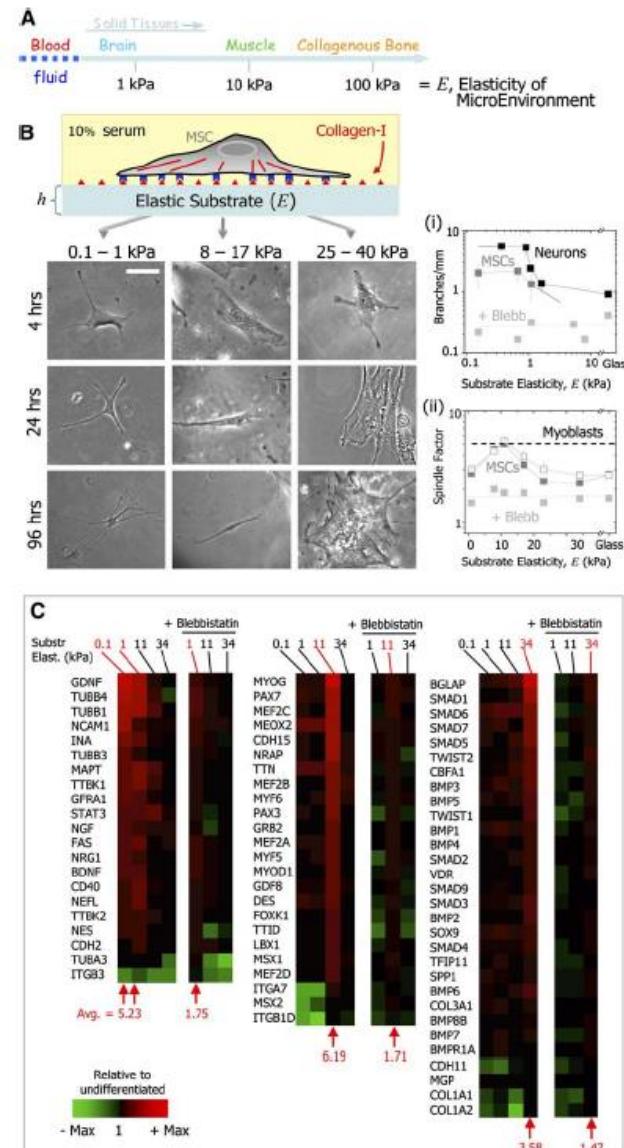
Benham-Pyle et al, 2015

Biological consequences of mechanotransduction

Mesenchymal stem cells.

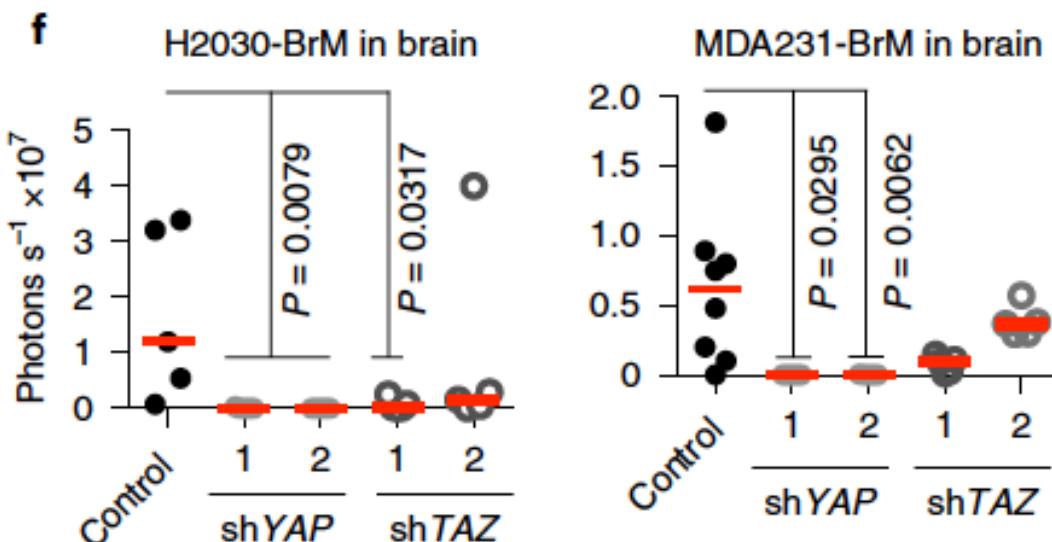
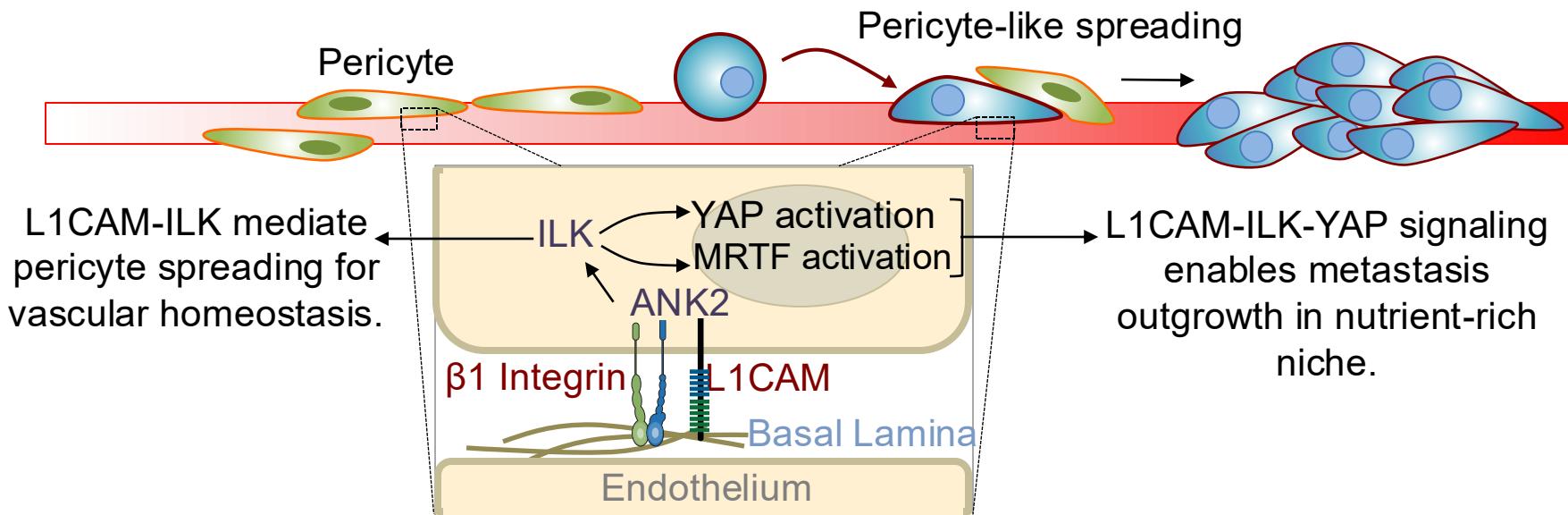
Cultured on hydrogels of different stiffness.

Stiffness determines whether the cells differentiate into neurons, muscle, or bone.



Engler et al, 2006

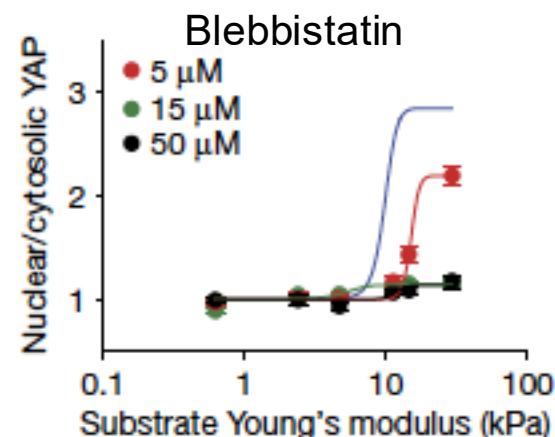
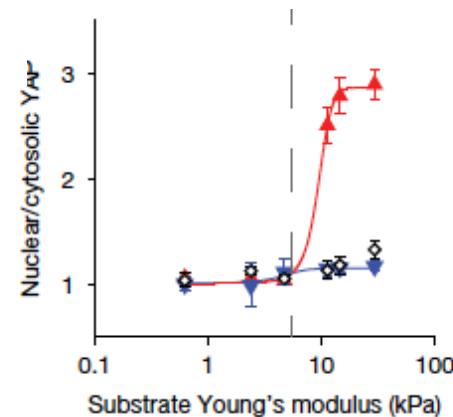
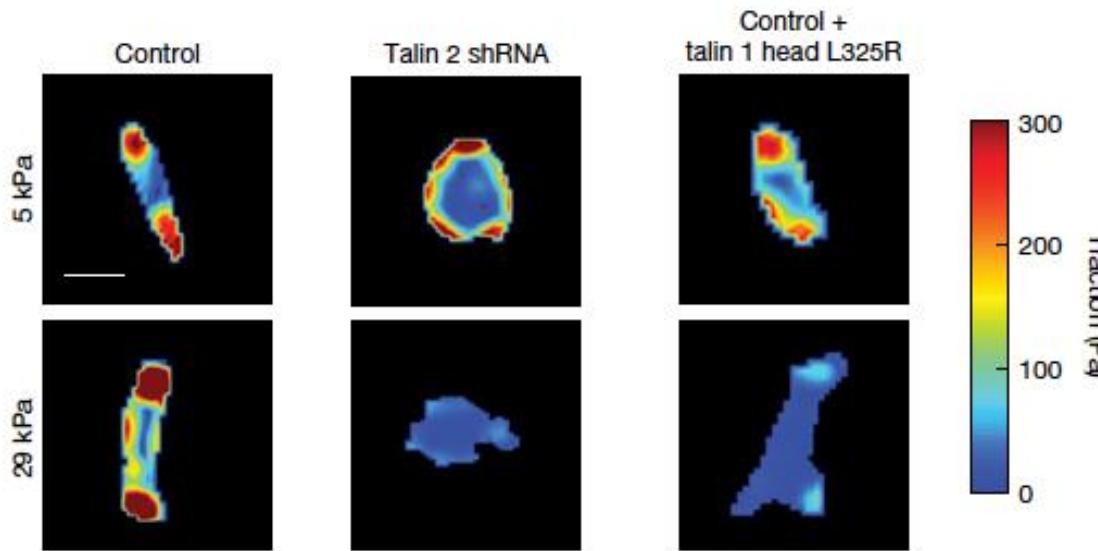
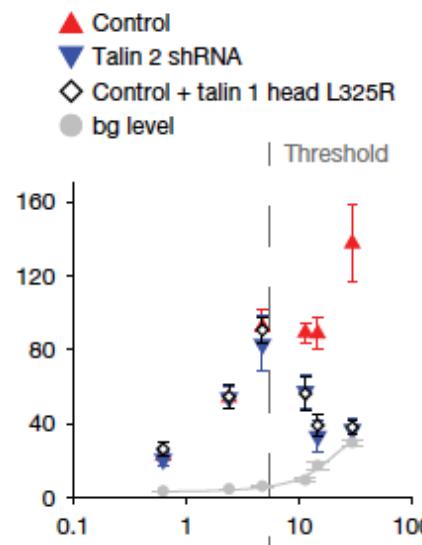
Biological consequences of mechanotransduction



Feedback between force exertion, mechanotransduction

TFM on hydrogels

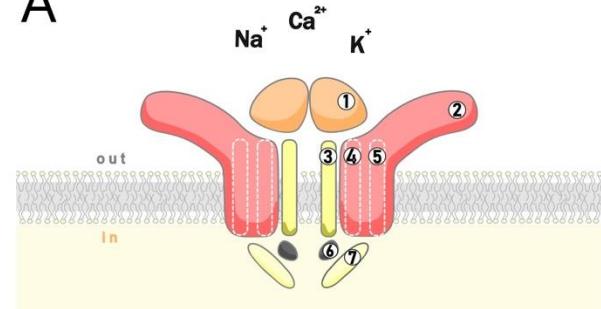
Plus/minus talin shRNA



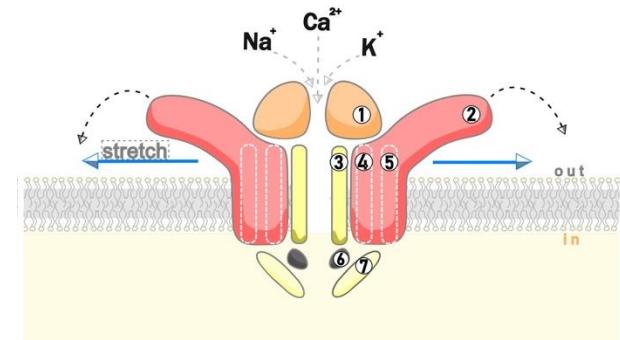
Other mechanosensing paradigms – mechanosensitive channels

Responsive to membrane tension – e.g. Piezo1

A

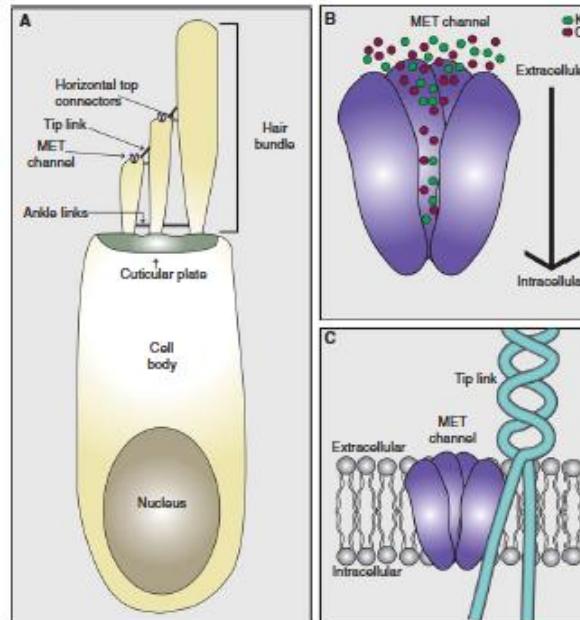


B



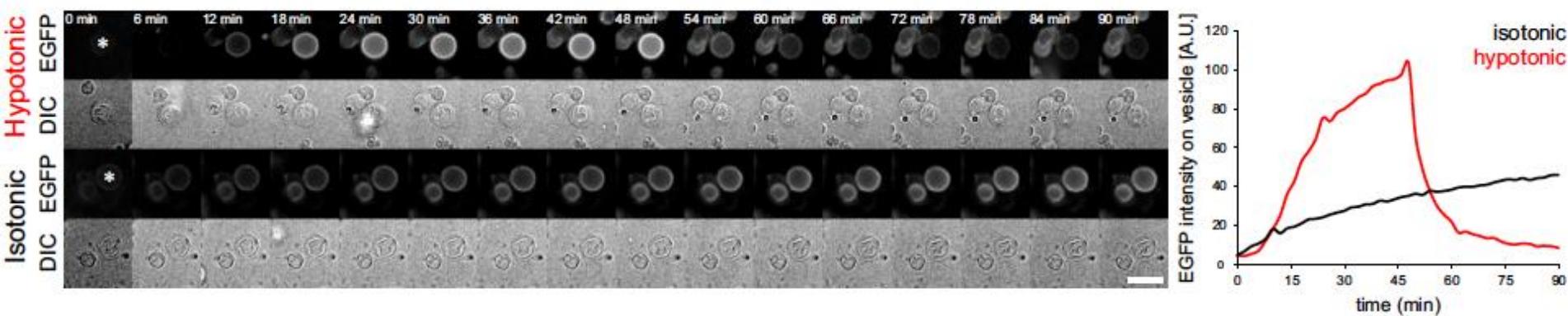
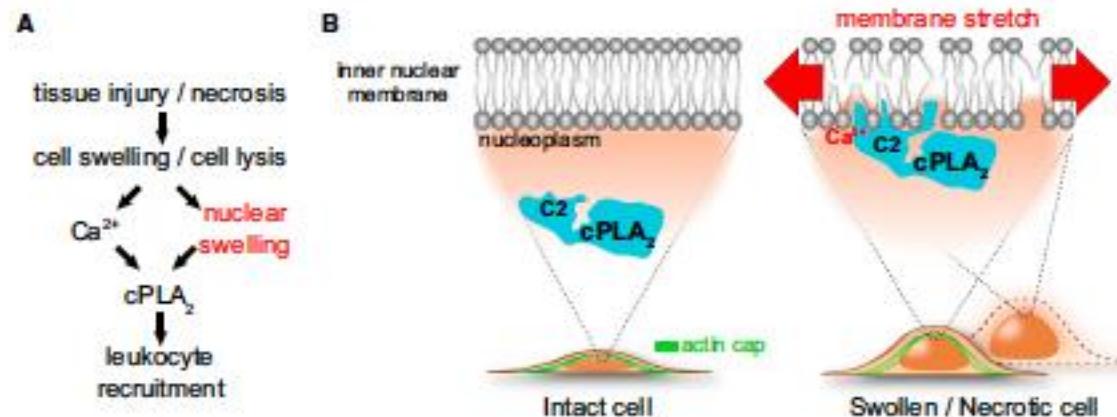
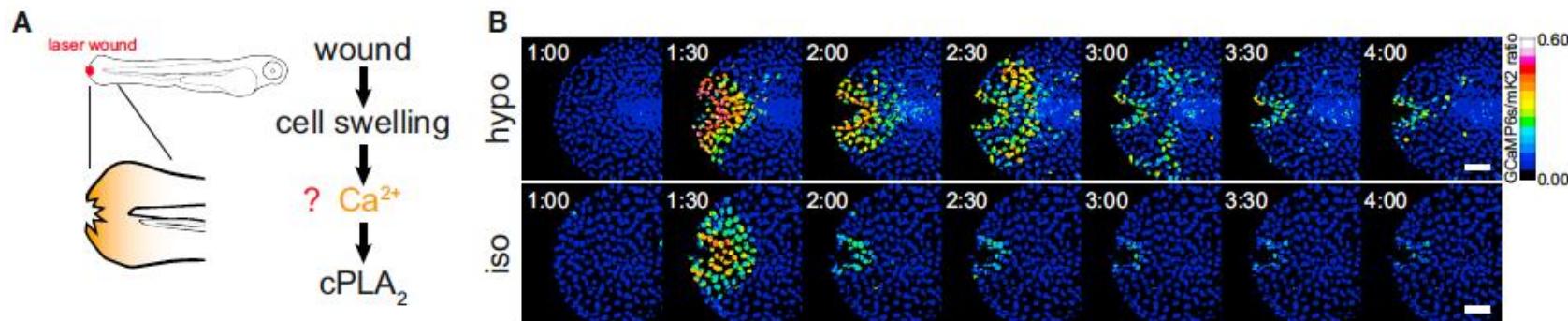
Simon Yel, wikipedia

Mechanoelectrical transduction in the ear



Cunningham and Müller, 2019

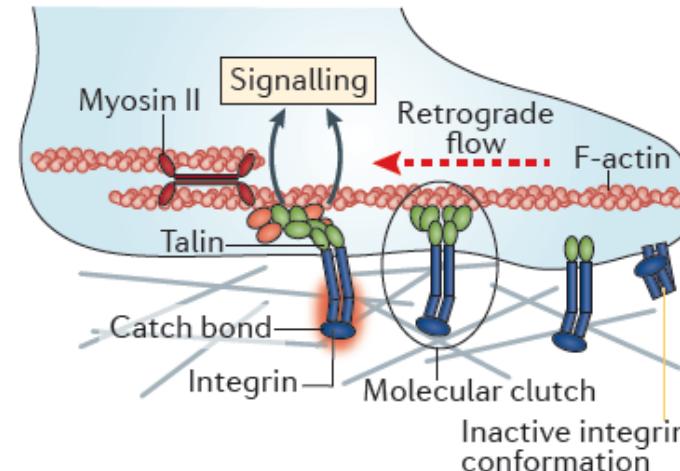
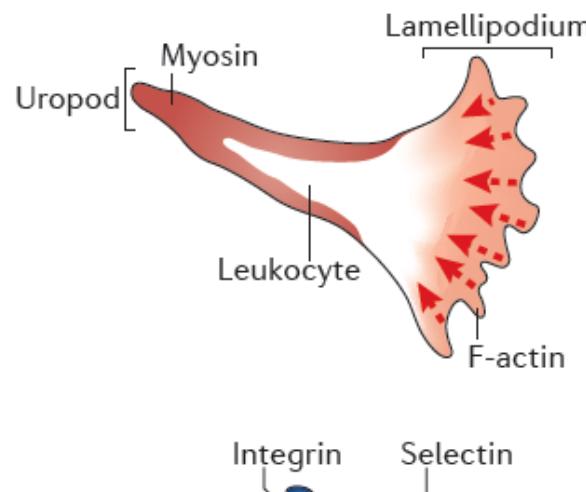
Other mechanosensing paradigms – mechanosensitive enzymes



Lecture outline

- 1) The mechanical properties of cells.
- 2) Mechanical force exertion by cells.
- 3) Mechanotransduction: pathways and functions.
- 4) Cell motility.**
- 5) Cell extrinsic forces.
- 6) Cell-cell interactions.

Cell motility – the mechanical clutch model

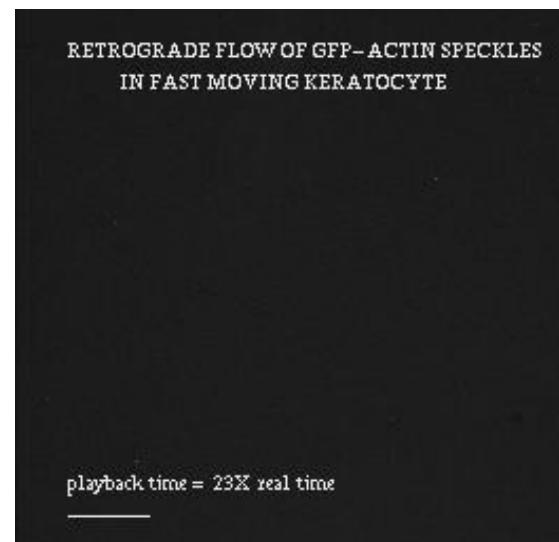


Huse, 2017

Fish keratocyte

Actin speckle microscopy

Notice that load on clutch is not 100%.
In other words, the clutch slips!



Jurado et al., 2004

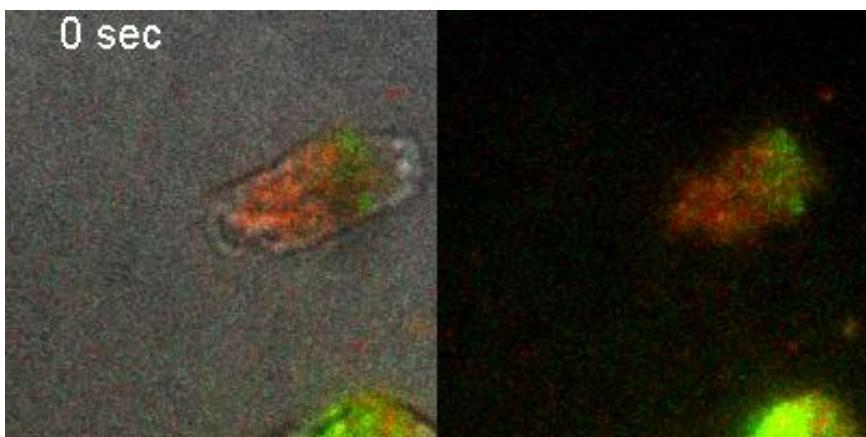
Different classes of motility

Canonical molecular clutch model developed from studies of mesenchymal cell migration in 2D environments. What about more dynamic cells (i.e. leukocytes) in 3D environments?

T cells exhibit both “walking” and “sliding” motility in 2D

Sliding

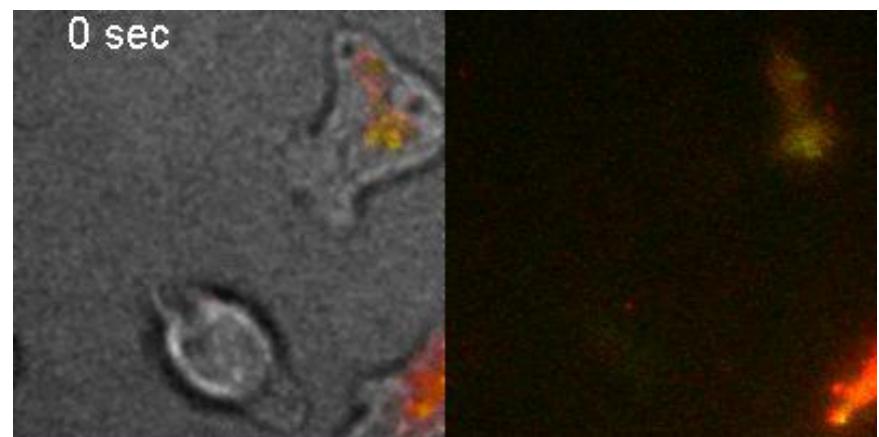
0 sec



Myosin

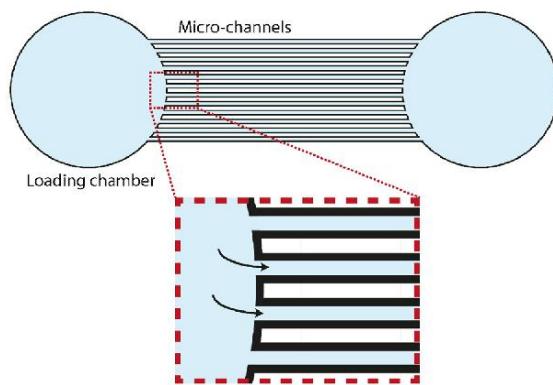
Walking

0 sec

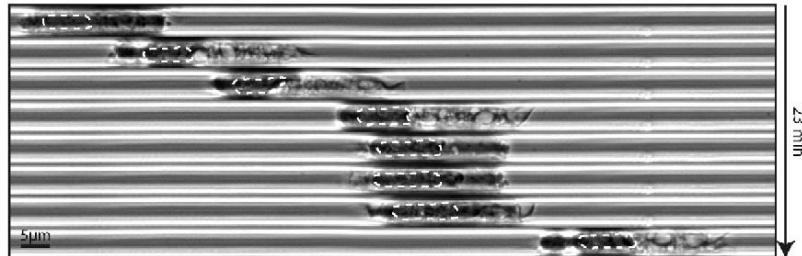


Actin

Different classes of motility

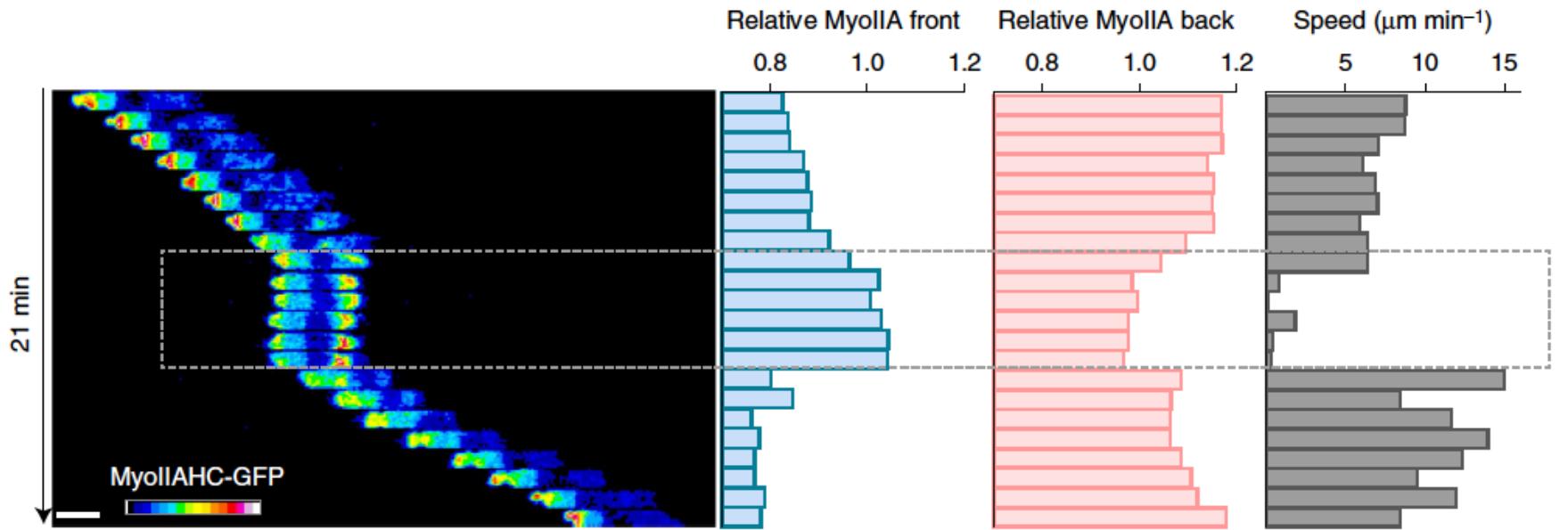


Dendritic cells in microchannels.



Vargas et al., 2015

Myosin accumulation at the back associated with forward movement.



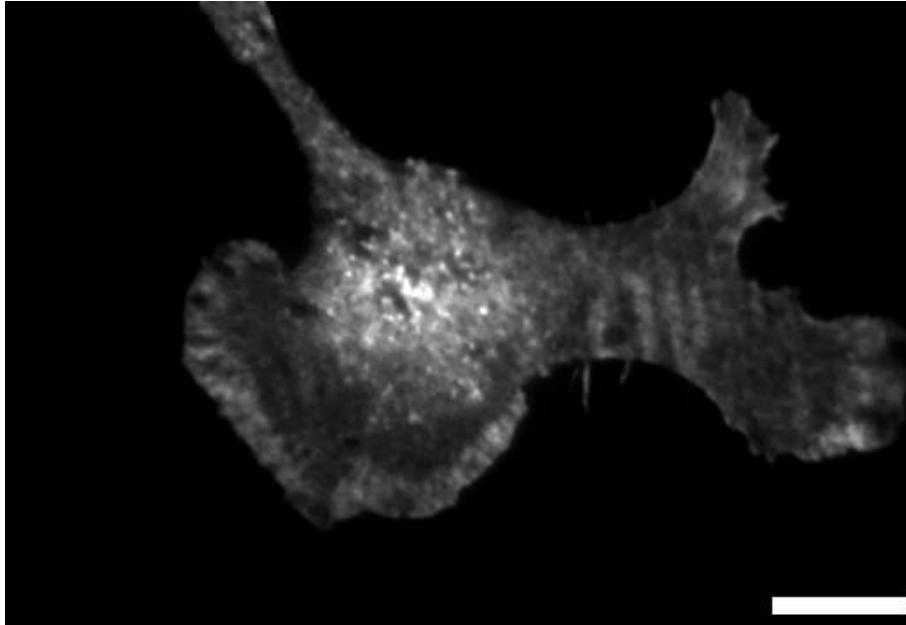
Chabaud et al., 2015

Different classes of motility

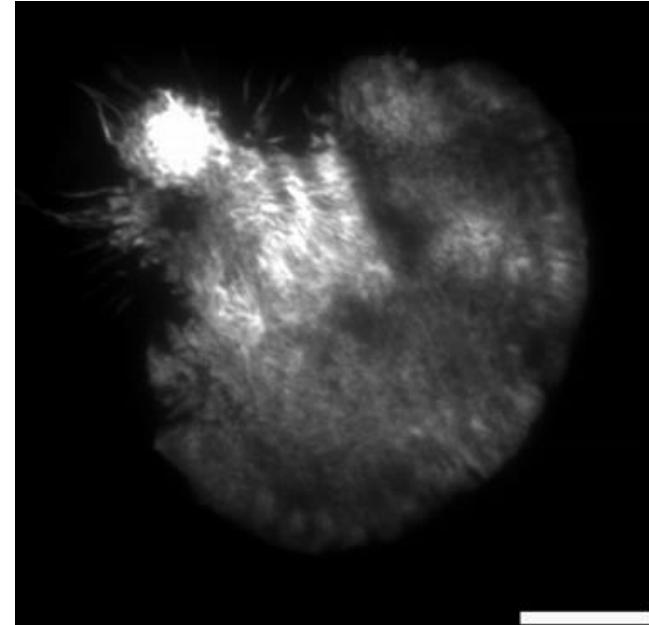
Migration is mechanically adaptive.

DCs migrating in different adhesion regimes, between coverslip and agarose pad.

Wild type



$\text{Itg}^{-/-}$

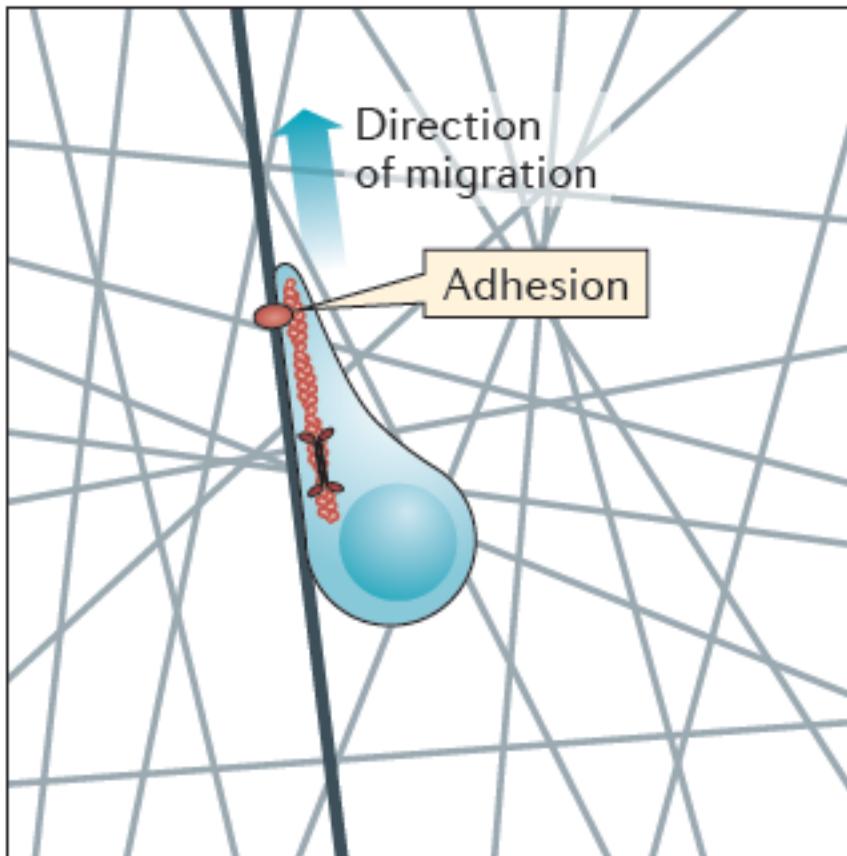


Renkawitz et al., 2009

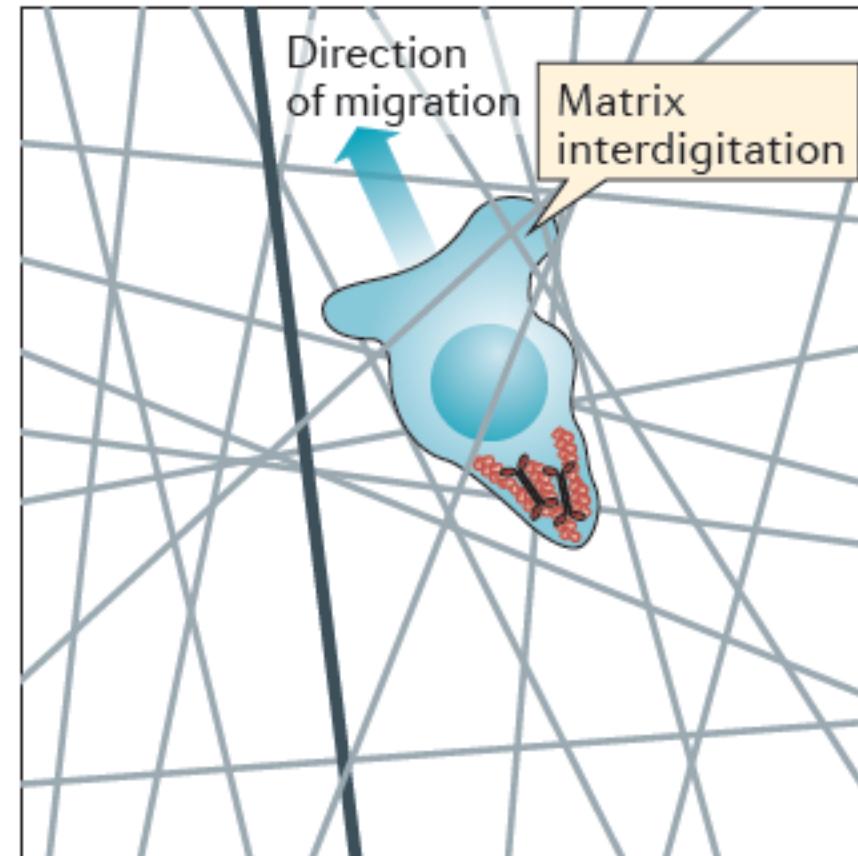
Forward progress is the same. Enhanced retrograde flow compensates for slippage.

Different classes of motility

3D environments give different options for motion.

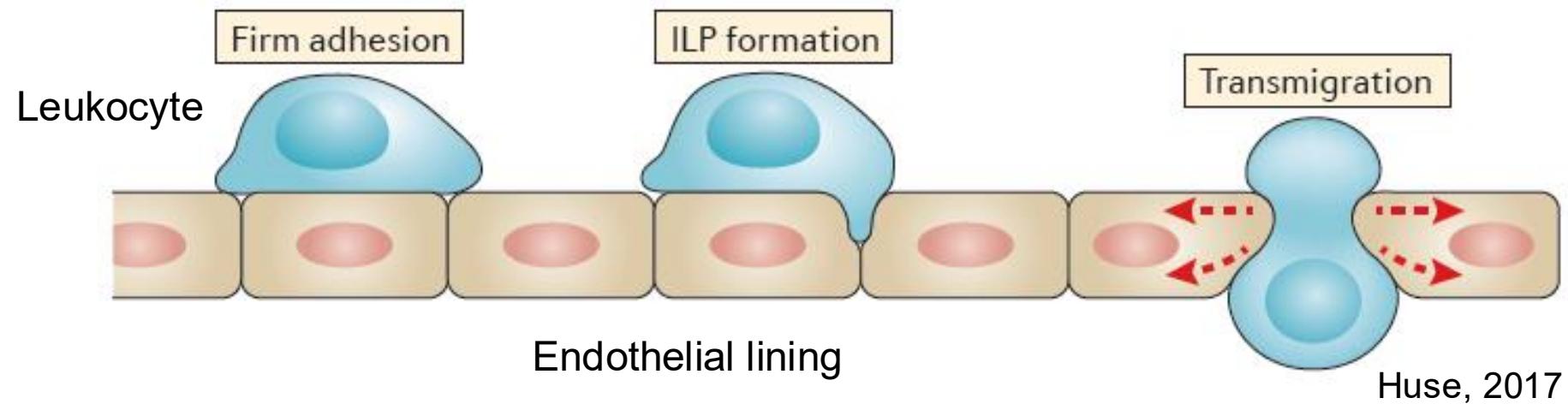


Integrin-dependent motility

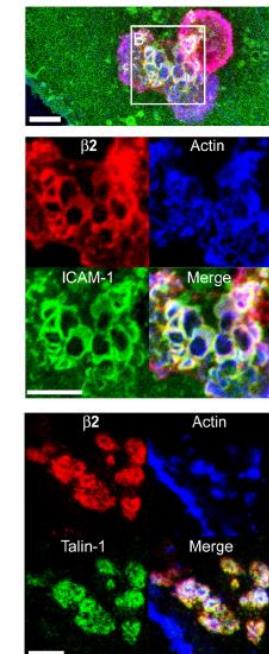
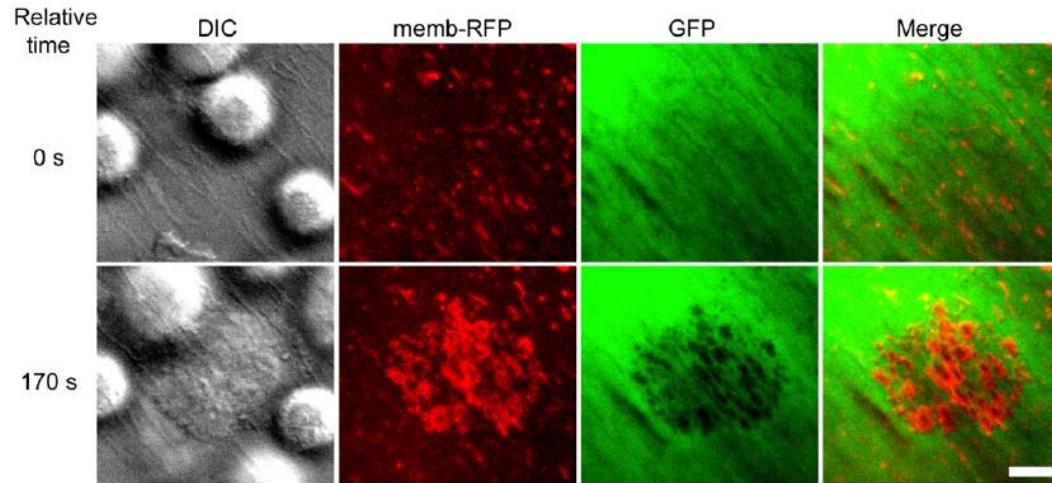


Integrin-independent blebbing motility

Motility - extravasation

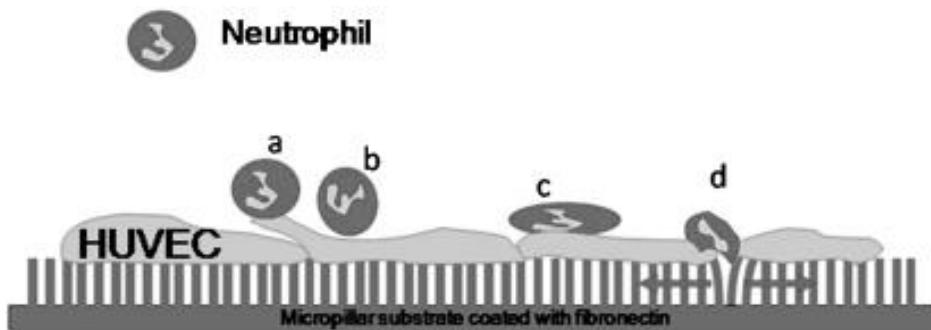


Carman. Invadopodial type pushing to “palpate” the monolayer.



Carman et al., 2007

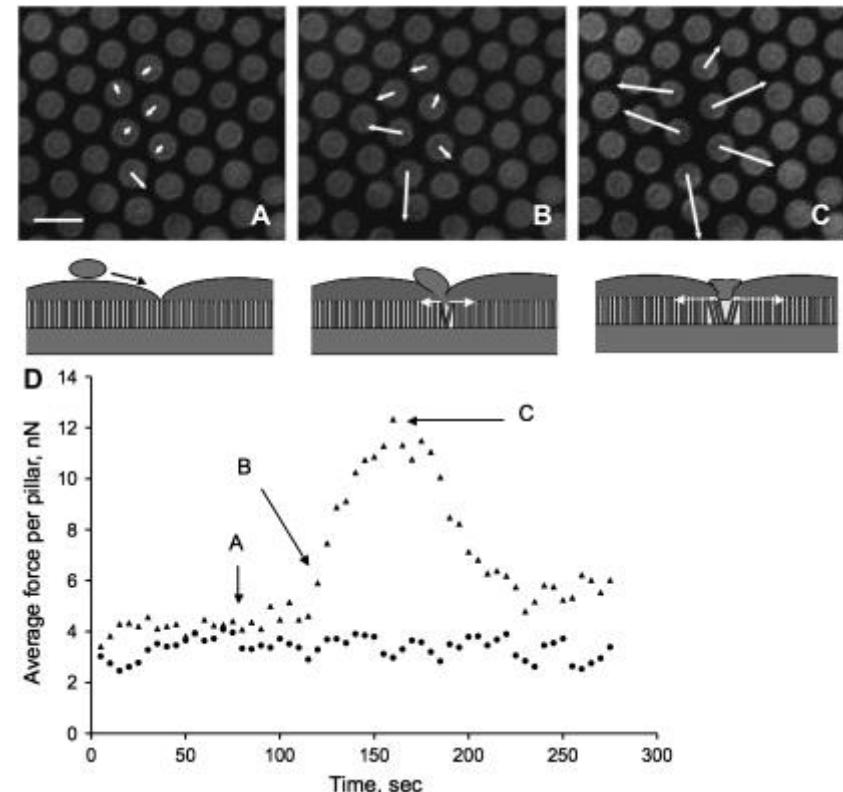
Extravasation requires nuclear squeezing



HUVEC endothelial layer grown on top of flexible micropillars.

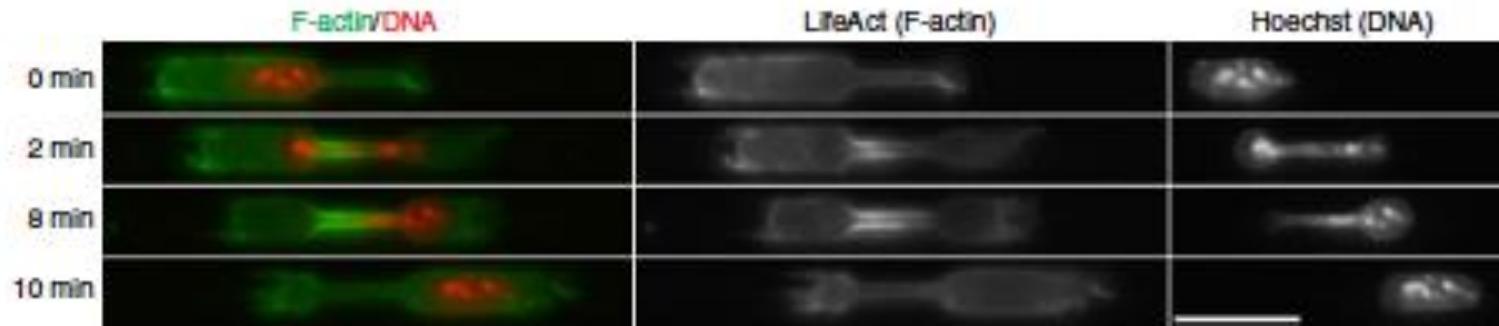
Peak force exertion during neutrophil extravasation is associated with passage of cell body.

Nucleus is particularly rigid and not deformable.



Nuclear squeezing leads to DNA damage

Actin and Arp2/3 dependent mechanism for nuclear squeezing.



Thiam et al., 2016

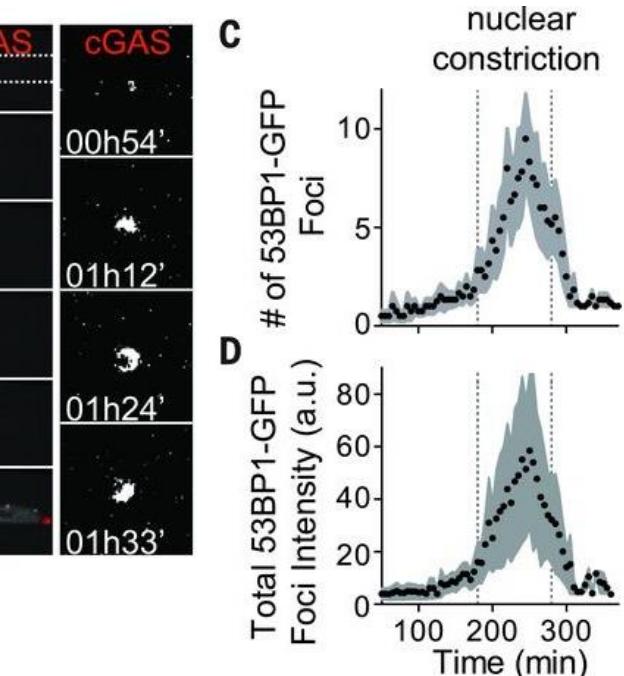
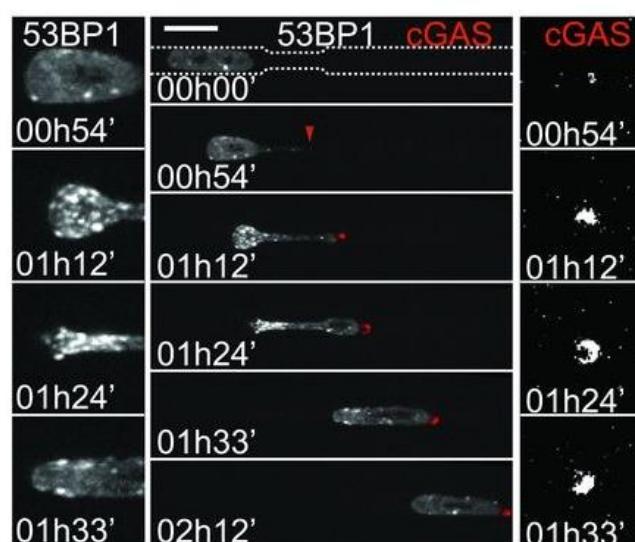
Nuclear squeezing results in DNA damage.

53BP1 – binds to dsDNA breaks.

cGAS – cytoplasmic DNA sensor.

Mechanisms for repairing damage.

Implications for cancer progression.



Raab et al., 2016

Lecture outline

- 1) The mechanical properties of cells.
- 2) Mechanical force exertion by cells.
- 3) Mechanotransduction: pathways and functions.
- 4) Cell motility.
- 5) **Cell extrinsic forces.**
- 6) Cell-cell interactions.

Environmental mechanics – extracellular matrix

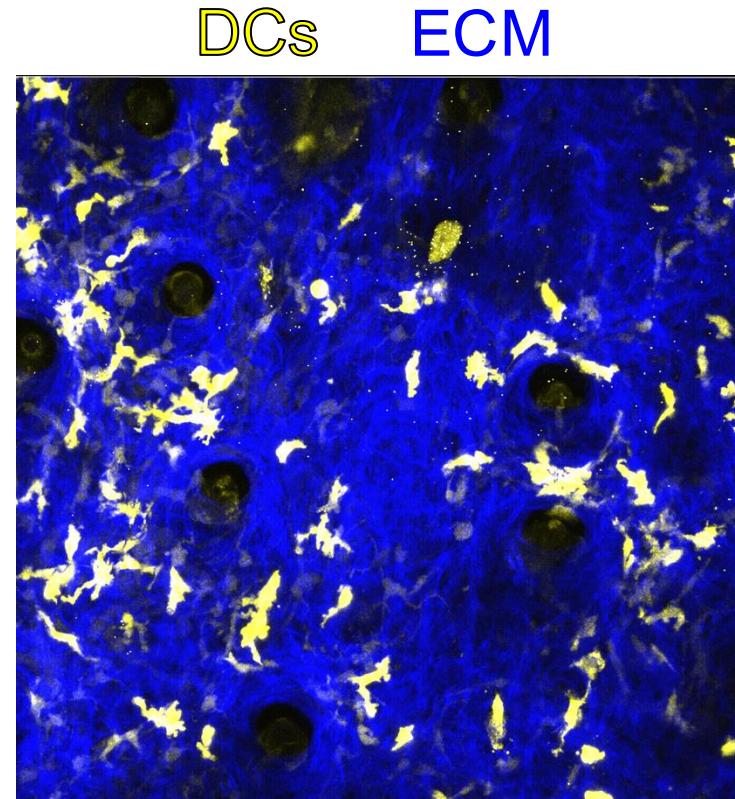
Complex and diverse composition, which dictates its mechanical properties.

Collagen – most abundant protein in mammals. Forms long triple helix. Contains modified amino acids, in particular hydroxyproline. Physical properties depend on composition and degree of crosslinking.

Glycoproteins – glycosylated polypeptides including fibronectin and syndecans.

Hyaluronic acid – Glycosaminoglycan, extruded from cells to form extracellular network analogous to cortical F-actin.

Laminins – Trimeric glycoproteins. Are a critical component of basal lamina.

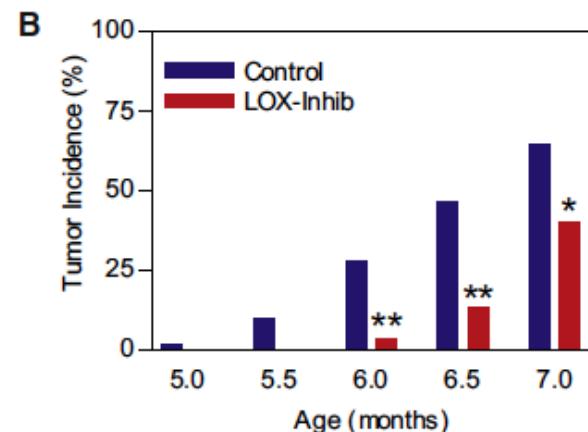
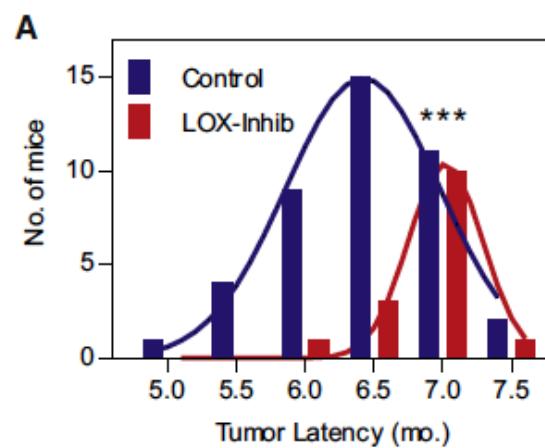
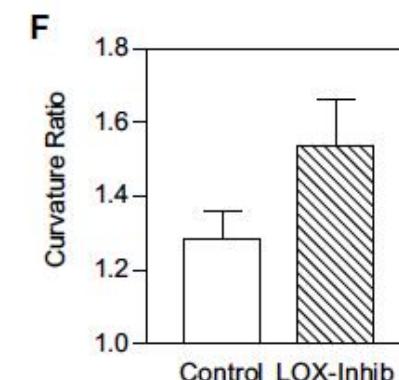
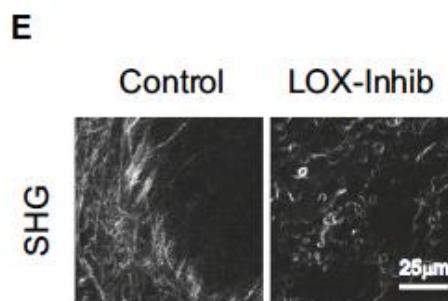
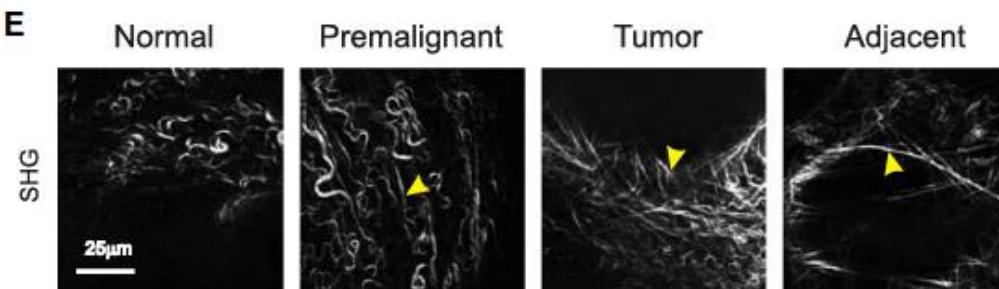


Environmental mechanics – extracellular matrix

MMTV-Neu autochthonous model of breast cancer.

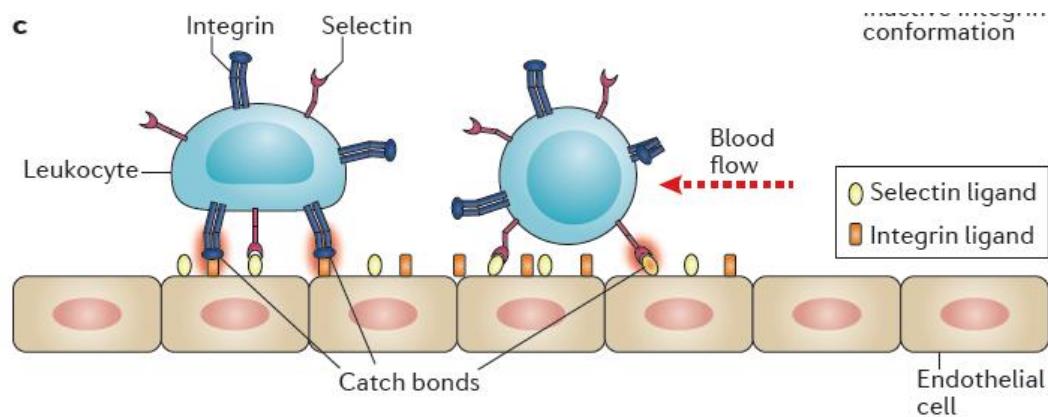
LOX = lysyl oxidase, required for collagen crosslinking.

LOX inhibition normalizes collagen and suppresses tumor development.



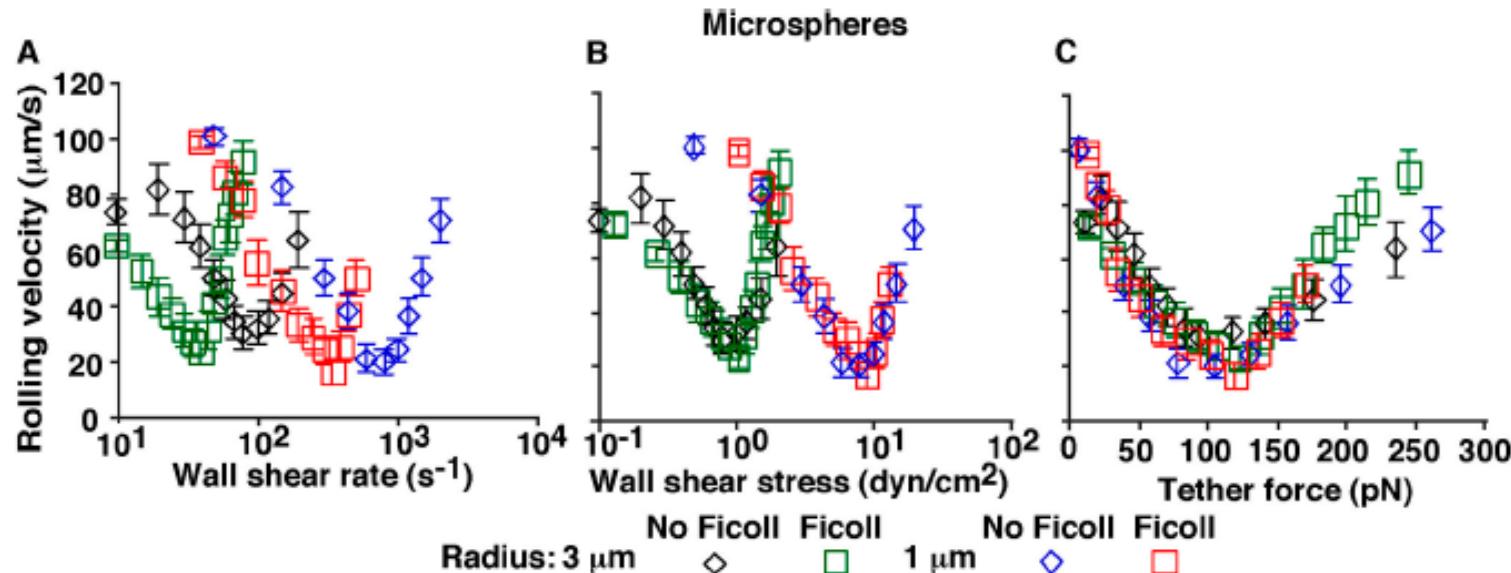
Environmental mechanics – flow

Certain adhesive proteins (selectins and integrins) have evolved to function under flow.



Huse, 2017

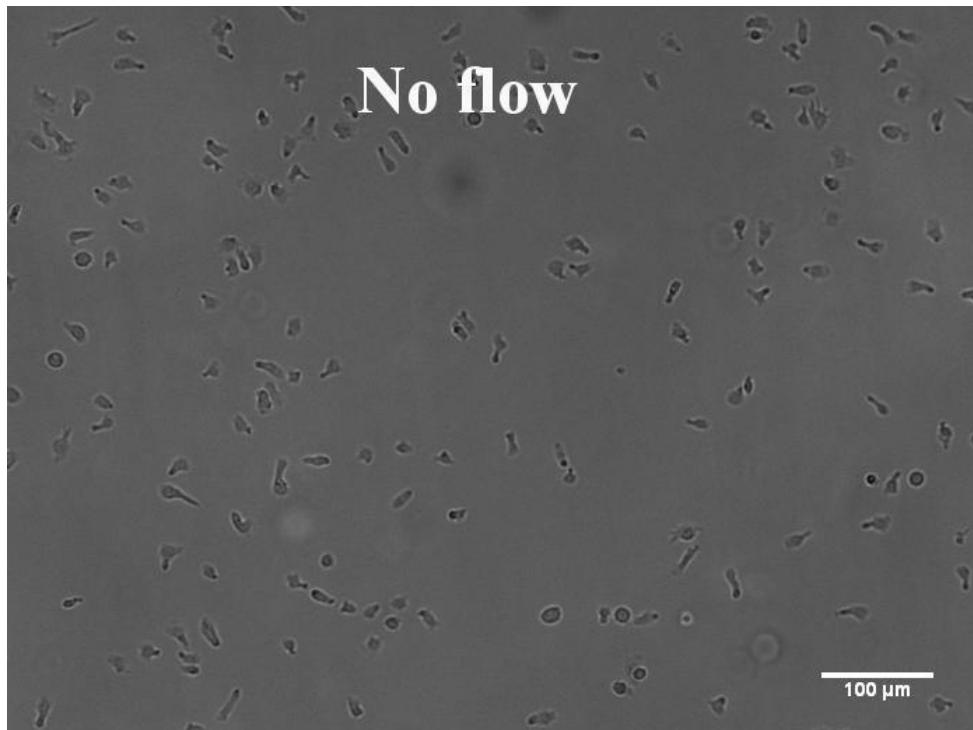
Selectin coated beads in ligand coated flow chamber. Rolling exhibits catch bond behavior.



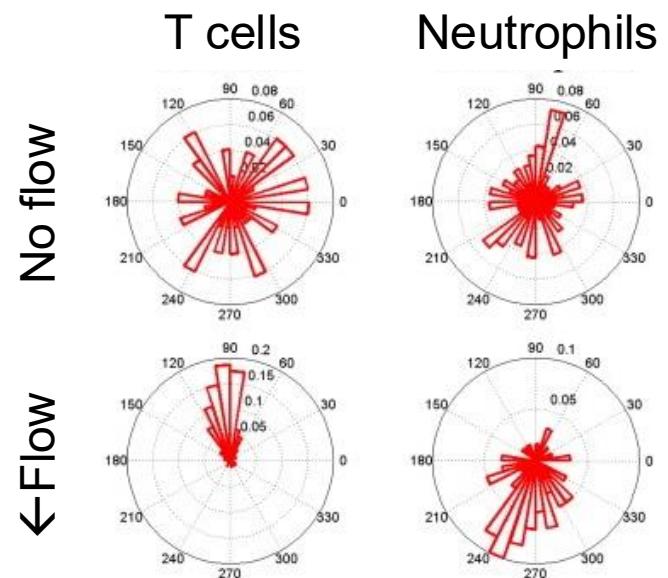
Yago et al., 2004

Environmental mechanics – flow

Leukocyte migration on integrin ligands is sensitive to the direction of flow.



T cells



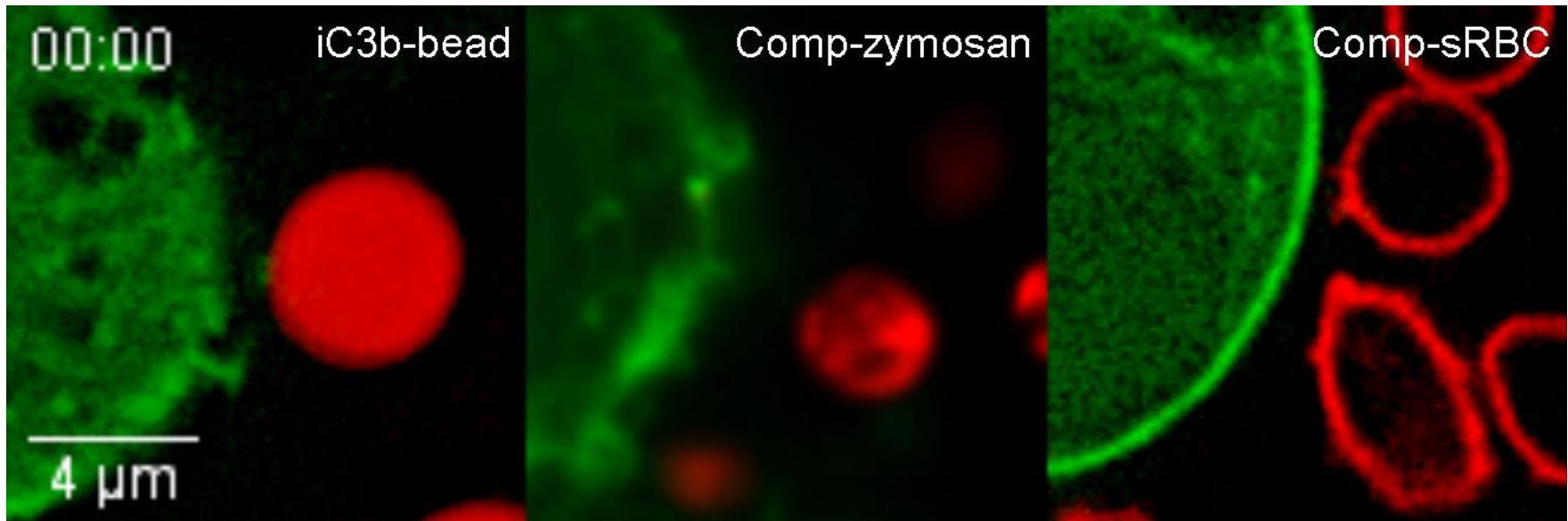
Valignet et al., 2013

Lecture outline

- 1) The mechanical properties of cells.
- 2) Mechanical force exertion by cells.
- 3) Mechanotransduction: pathways and functions.
- 4) Cell motility.
- 5) Cell extrinsic forces.
- 6) Cell-cell interactions.

Phagocytosis

Critical for pathogen clearance but also homeostatic uptake of debris and dead cells. Must be both chemically and biophysically adaptable.

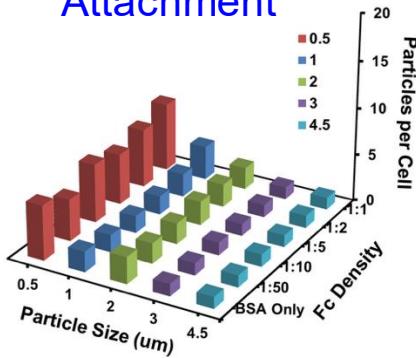


Jaumouillé et al., 2019

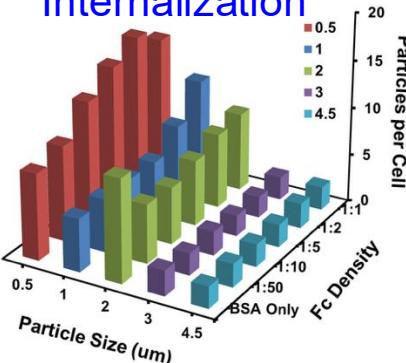
Phagocytosis

Size dependence

Attachment

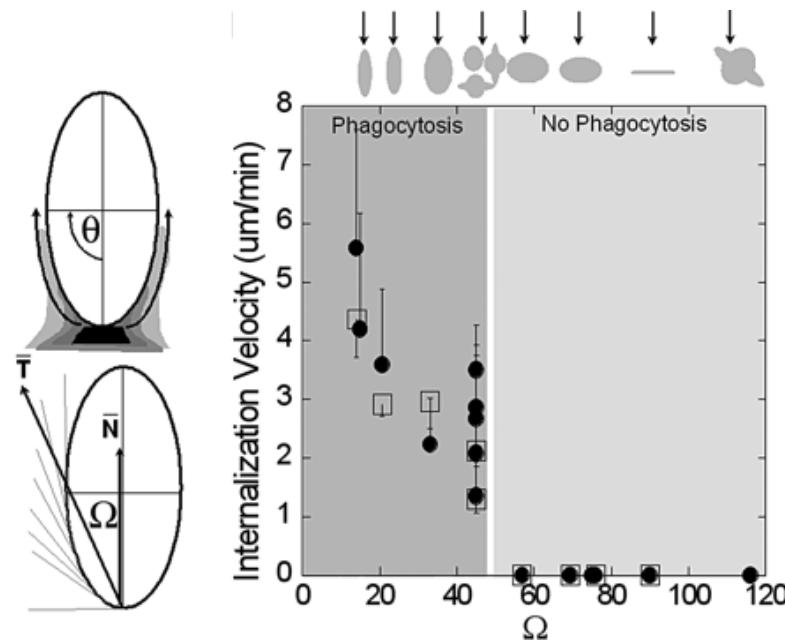


Internalization



Pacheco et al., 2013

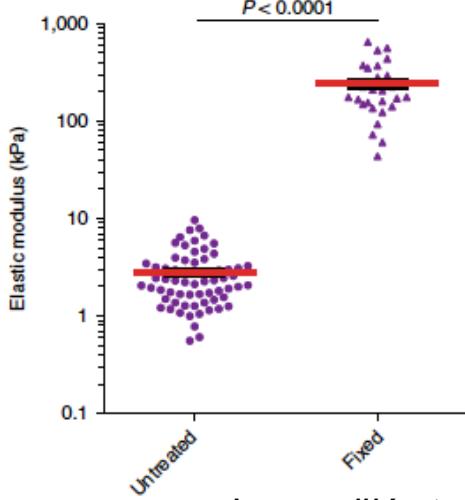
Shape dependence



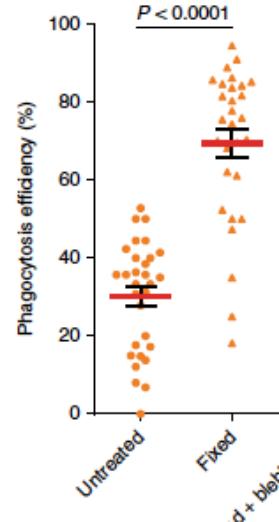
Champion and Mitragotri, 2006

Stiffness dependence

Stiffness



Uptake

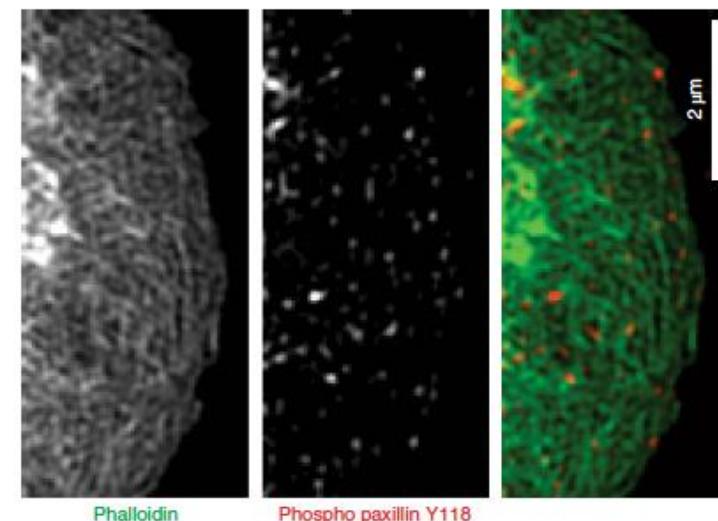
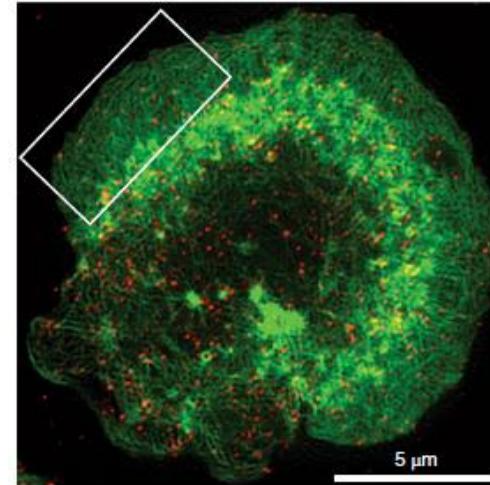
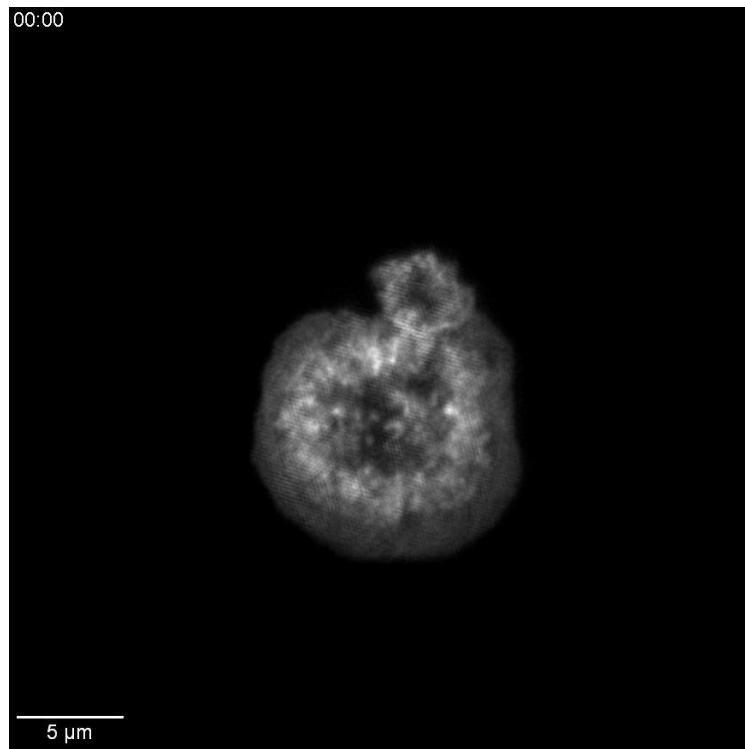


Jaumouillé et al., 2019

Phagocytosis

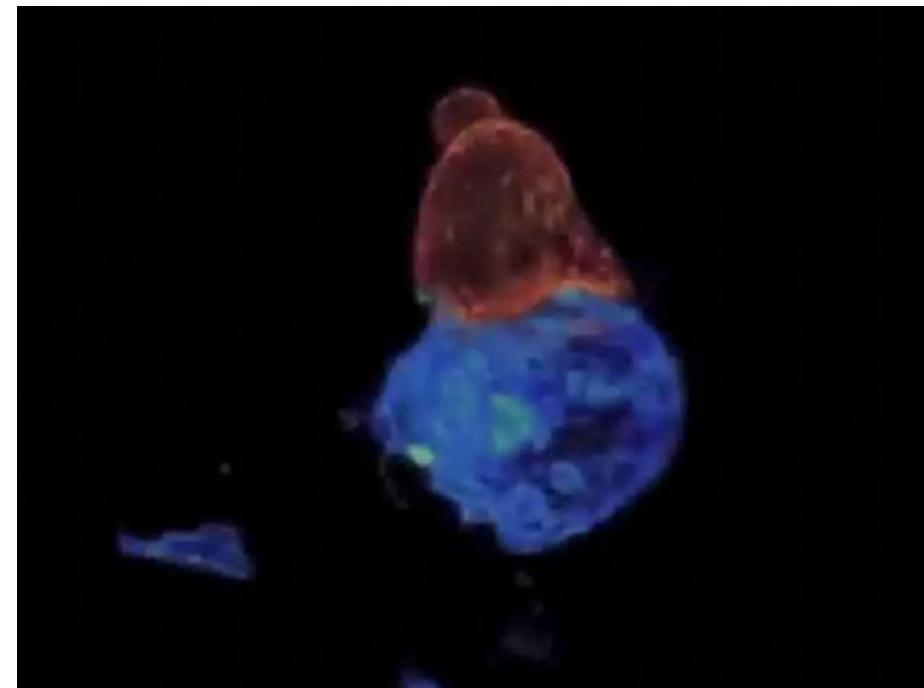
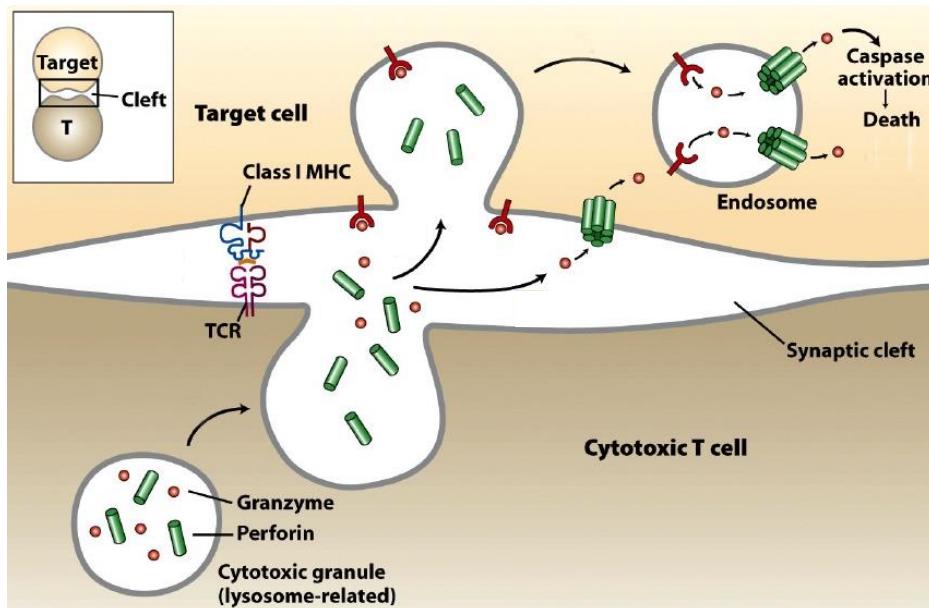
Clutch like adhesions are employed during phagocytosis.

Frustrated phagocytosis model.



Immune synapse

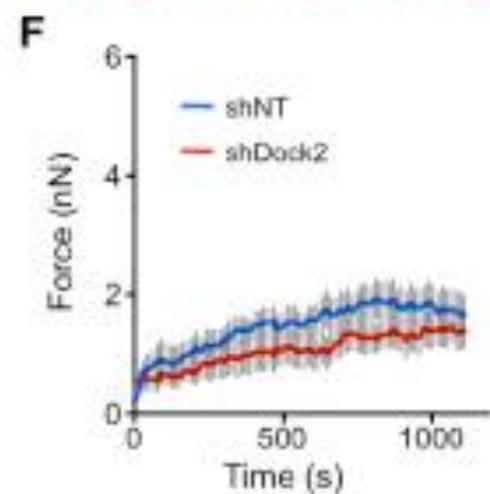
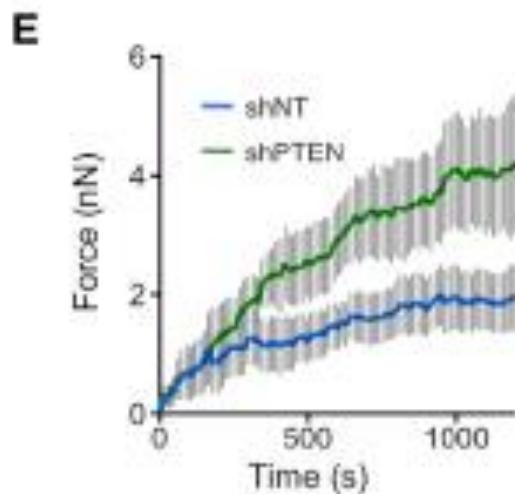
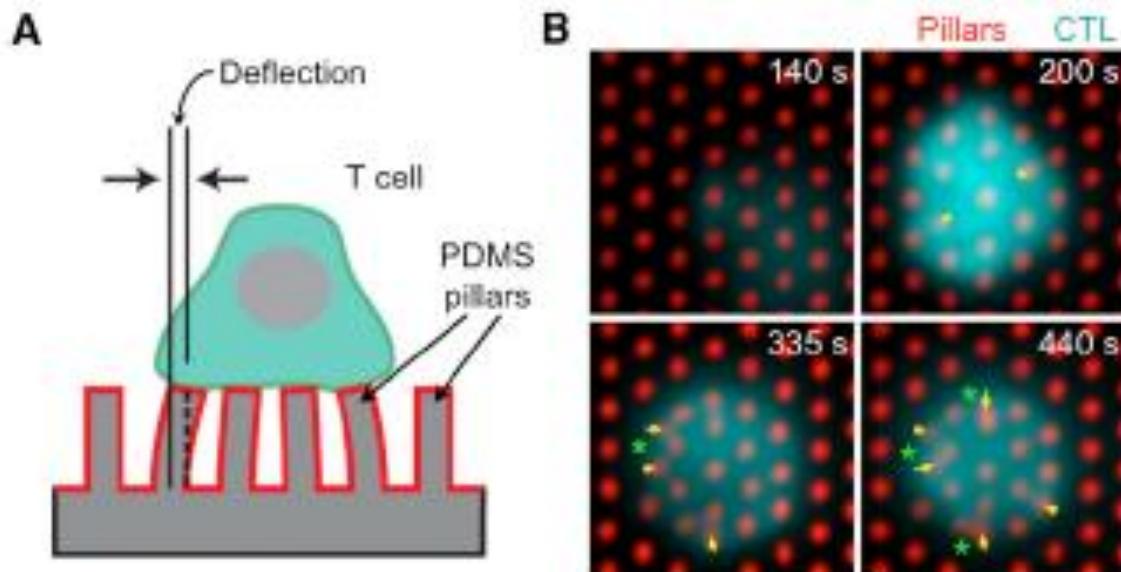
Critical for immune responses against intracellular pathogens and tumors.
Mediated by a cellular venom (perforin and granzyme), but also highly dynamic.



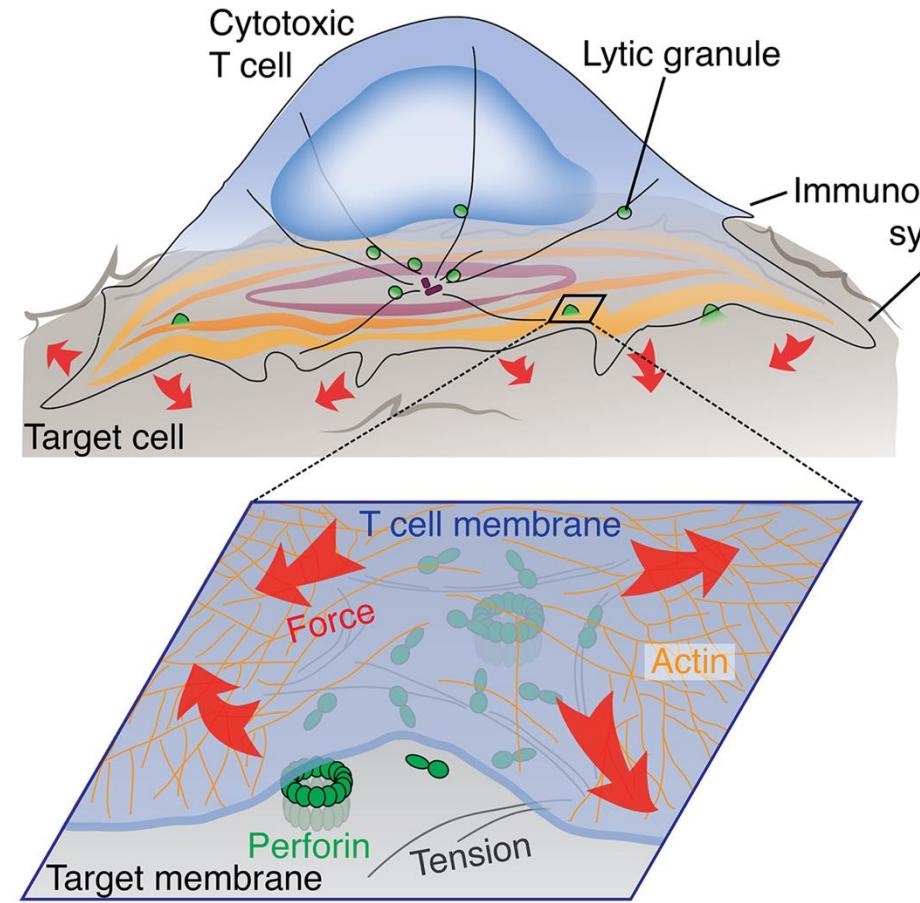
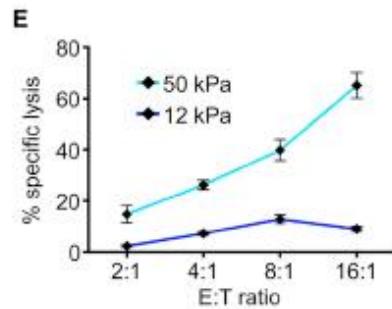
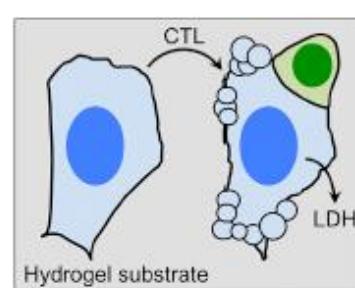
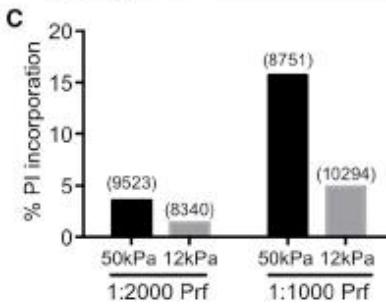
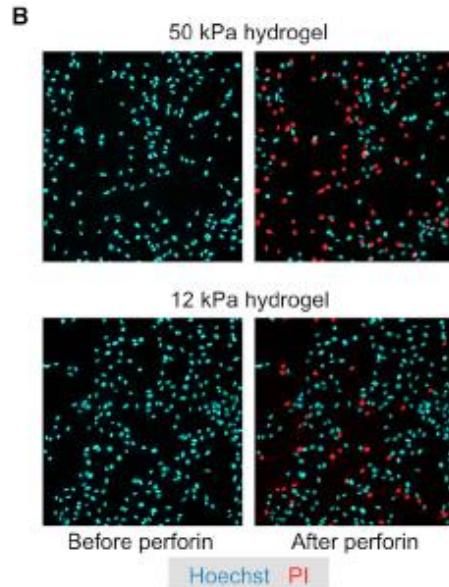
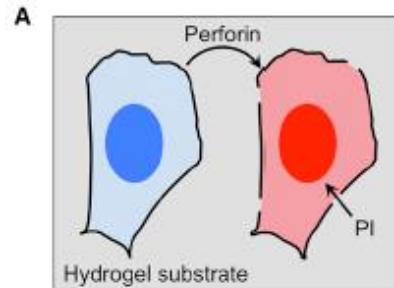
Adapted from StudyBlue, Iowa State University

Ritter et al., 2015

Synaptic force correlated with cytotoxicity

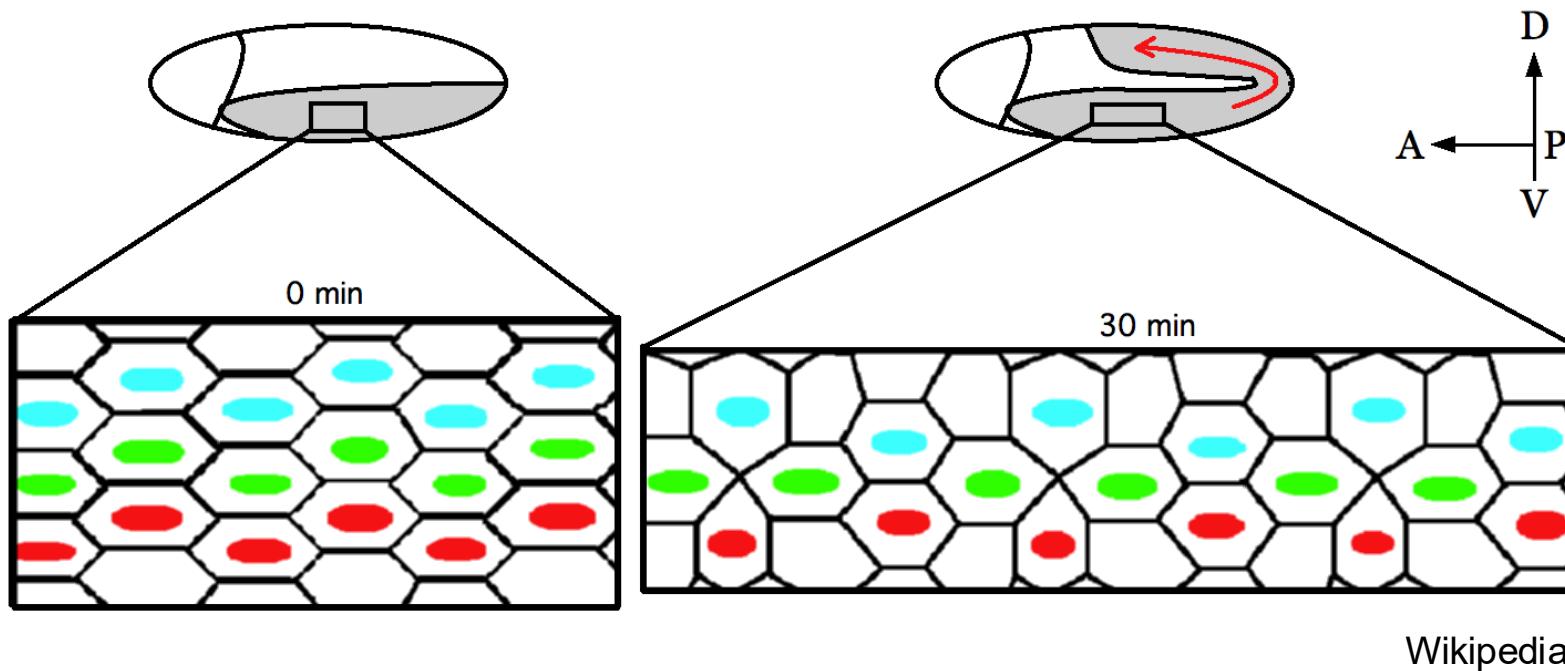


Tension potentiates perforin-mediated lysis



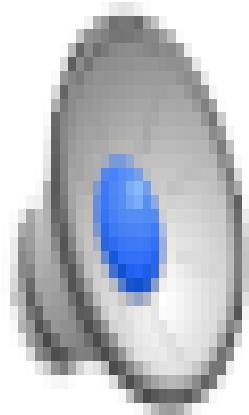
Embryonic development – convergent extension

Enables development of a body axis from a homogeneous layer of cells.

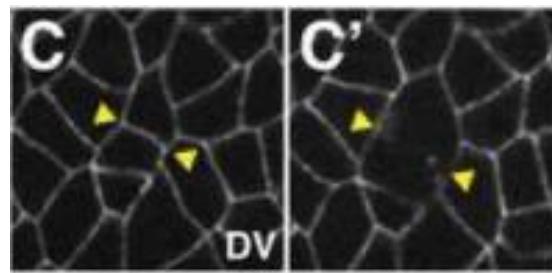
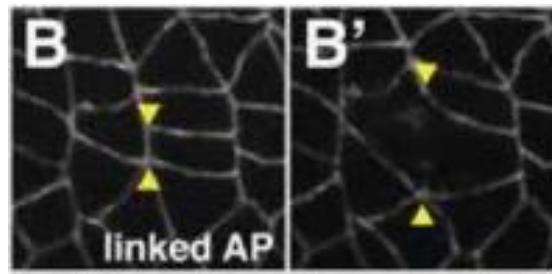
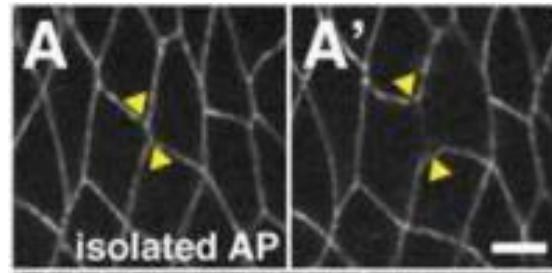


Myosin contractility is crucial for convergent extension

Myosin label



Laser ablation of cell-cell junctions.



Tension stimulates mechanosensitive junction formation

