

New insights into cancer immunoediting and its three component phases – elimination, equilibrium and escape

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The principles of cancer immunoediting have set the foundations for understanding the dual host-protective and tumour sculpting actions of immunity on cancer and establishing the basis for novel individualized cancer immunotherapies. During cancer immunoediting, the host immune system shapes tumour fate in three phases through the activation of innate and adaptive immune mechanisms. In the first phase, Elimination, transformed cells are destroyed by a competent immune system. Sporadic tumour cells that manage to survive immune destruction may then enter an Equilibrium phase where editing occurs. The Escape phase represents the third and final phase of the process, where immunologically sculpted tumours begin to grow progressively, become clinically apparent and establish an immunosuppressive tumour microenvironment. This review focuses on important recent developments that have enhanced our understanding of each phase of the cancer immunoediting process, summarizes the discovery of new predictive and prognostic biomarkers and discusses development of novel and objectively effective cancer immunotherapies.

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Introduction

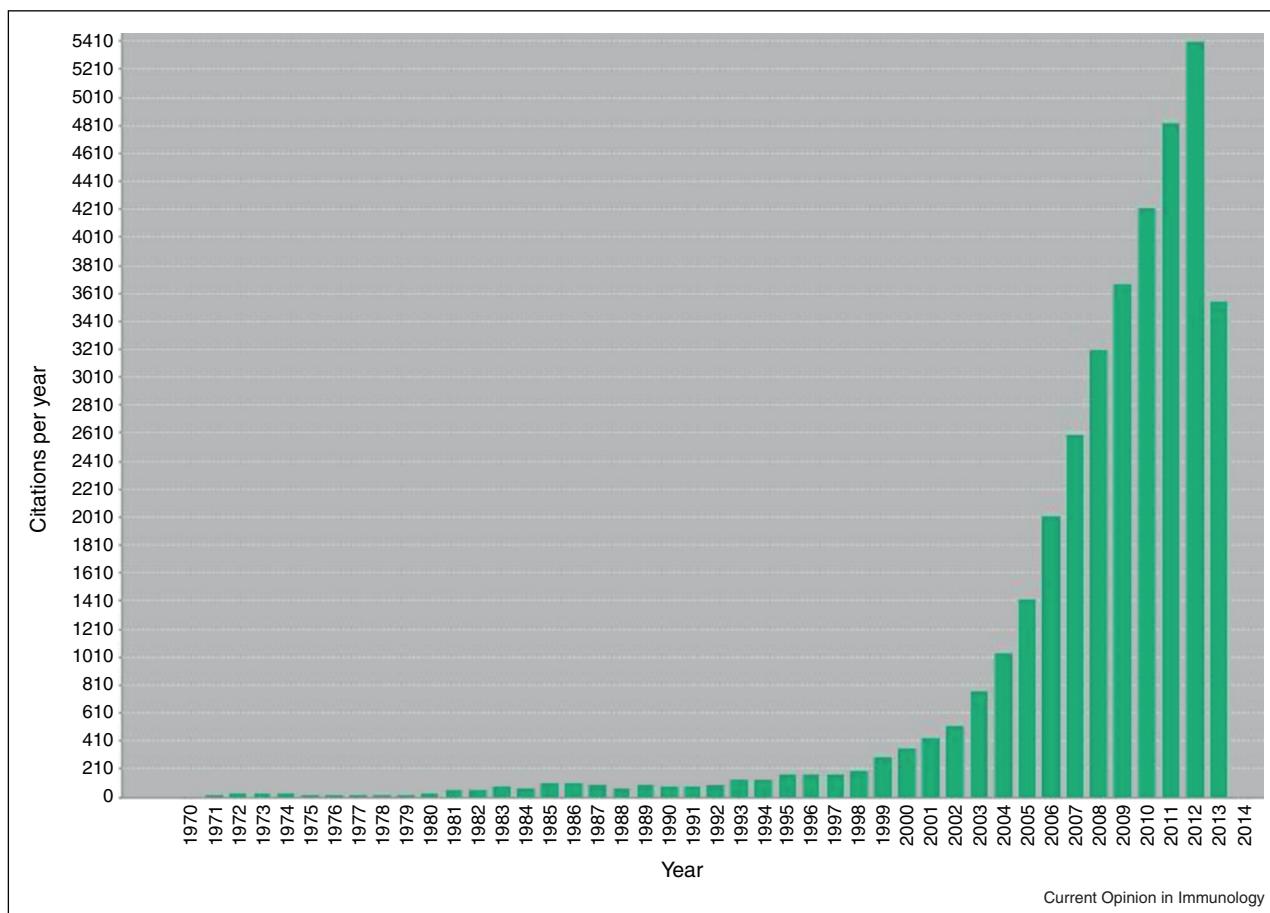
The last two decades have seen the end of the long-standing argument about whether the immune system has positive, negative or null effects on tumour development. Recent work from many labs have unequivocally documented that immunity can, in fact, facilitate cellular transformation, prevent or control tumour outgrowth and shape the immunogenicity of tumours. These three

apparently paradoxical functions of the immune system are separable based on their temporal occurrence during tumour formation, the nature of the transforming event, the particular components of immunity involved in each process, and in the nature of the tumour specific antigens expressed in the transformed cell. Whereas one body of work has clearly established the capacity of chronic inflammation to initiate and promote cancer [1], a second set of studies from other laboratories, including our own, has revealed that an intact immune system can prevent/control and shape/promote cancer by a process we call 'Cancer Immunoediting' [2,3]. The evolution of the cancer immunoediting concept from the older and perhaps more controversial 'cancer immunosurveillance' hypothesis has helped interpret the predictive and prognostic significance of immune infiltrates into tumours.

The immune surveillance theory originally proposed more than 50 years ago by Burnet and Thomas predicted that the immune system acted as a sentinel in recognizing and eliminating nascent transformed cells [4]. Extensive work over the past 15 years revealed that this surveillance function of immunity was only a part of the story and prompted us to refine and extend the concept into one we call 'cancer immunoediting' to more accurately describe the many facets of immune system–tumour interactions [2,3]. This dynamic process, whereby the immune system not only protects against cancer development but also shapes the character of emerging tumours, is composed of three phases — Elimination, Equilibrium and Escape — and has been extensively reviewed elsewhere [5,6]. Whereas the Elimination phase has largely been inferred from mouse tumour model studies, evidence for the Equilibrium and Escape phases have come from analyses of cancers in both mice and humans. Consequently, escape from immune control is now recognized to be one of the 'Hallmarks of Cancer' [7]. The emergence of cancer immunoediting as a framework to understand the extent of the immune system's interaction with cancer, has, in part, prompted a recent burgeoning of the scientific literature discussing this process as demonstrated by dramatically increased citation (Figure 1). Herein we review recent findings that have added to our understanding of cancer immunoediting and discuss the relevance of this process to cancer immunotherapy.

Immunoediting

The role of the immune system in shaping the immunogenicity of tumours has now been unequivocally

Figure 1

Number of 'cancer immunoediting' citations per year demonstrating the importance and increased interest in the field over recent years (derived from search of above terms in ISI Web of Science).

established. Tumours arising in RAG2-deficient mice (lacking T, B and NKT cells) are, as a group, more immunogenic than those derived from immunocompetent hosts [5]. A central principle of cancer immunoediting is that T-cell recognition of tumour antigens drives the immunological elimination or sculpting of a developing cancer. However, until recently, little was known about the antigens expressed in nascent tumour cells, whether they are sufficient to induce anti-tumour immune responses, or whether their expression can be altered following interaction with the immune system.

Recently, we used a genomics approach to determine the mutational landscape of a highly immunogenic, unedited sarcoma cell line derived from methylcholanthrene (MCA)-treated *Rag2*^{-/-} mice, which represent good models of nascent primary tumour cells. By combining exome sequence analysis with MHC class I prediction algorithms, a point mutation in Spectrin-β2 was identified and validated as the source of a neo-epitope in the unedited *Rag2*^{-/-} derived d42m1 MCA sarcoma that

functioned as a major immunodominant rejection antigen [8*]. This study showed that cancer immunoediting was the consequence of a T-cell-dependent immunoselection process leading to the outgrowth of tumour cell clones lacking immunodominant rejection antigens that displayed reduced immunogenicity.

A similar conclusion was reached by DuPage *et al.* using a genetic mouse model of sarcomagenesis. This model employed immunocompetent and immunodeficient mice engineered to express a constitutively activated, oncogenic form of Kras and a floxed *p53* tumour suppressor gene [9*,10]. Intramuscular injection of a lentivirus encoding the cre-recombinase plus strong class I model epitopes (SIINFEKL and SIYRYYGL) led to the outgrowth of SIINFEKL and SIYRYYGL expressing sarcomas in immunodeficient mice. By contrast, the appearance of sarcomas in lentivirus transduced immunocompetent mice was delayed and those that grew out lacked expression of the model antigens. In this model, editing of the SIINFEKL and SIYRYYGL expression

was the result of a T cell-dependent process that resulted in the epigenetic silencing of the exogenously introduced genes encoding the model antigens.

The clinical relevance of the above findings has been supported or confirmed in both preclinical and human studies. First our study, together with a distinct, independently published study by the Sahin group [11] established the principle that tumour exome analysis provides an opportunity to rapidly identify tumour specific mutational antigens thus forming the basis for development of individualized cancer immunotherapies. Subsequent work by others has indeed extended this finding to human cancer patients [12,13]. Second, a recent report, which followed the progression of a melanoma patient whose tumour was initially positive for the antigens NY-ESO-1, MAGE-C1 and Melan-A, showed that treatment with a vaccine targeting NY-ESO-1 lead to the outgrowth of tumours that lacked NY-ESO-1 but not MHC Class I, MAGE-C1 and Melan-A. This study, together with others like it, provides evidence that immunoediting also occurs as a consequence of immunotherapy in human cancer patients [14]. Additionally, analysis of patients in a phase I clinical trial receiving an NY-ESO vaccine showed that participants who relapsed had NY-ESO or MHC class I negative tumours [15]. Together these studies point out that cancer immunoediting is a process that occurs in both mice and humans. In addition, they reveal the crucial need to target multiple tumour antigens during cancer immunotherapy in order to prevent outgrowth of tumour cell variants that lack expression of individual tumour specific antigens.

Although these studies have established the importance of adaptive immunity, and particularly T cells, in the editing process, other recent studies have revealed that innate immunity can also edit tumour immunogenicity. To assess the role of the innate immune system in cancer immunoediting, the tumourigenicity and immunogenicity of MCA-induced sarcomas derived from WT, *Rag2*^{-/-}, and *Rag2*^{-/-} × *γc*^{-/-} (lacking all lymphocytes, including NK cells) were recently compared. MCA-induced sarcoma incidence was greatest in *Rag2*^{-/-} × *γc*^{-/-} mice, lower in *Rag2*^{-/-} mice and much lower in WT mice. Tumour cells generated in *Rag2*^{-/-} × *γc*^{-/-} mice showed an increased regressor frequency (rejected when transplanted into immunocompetent host) compared with cell lines derived from *Rag2*^{-/-} mice. Regressor cell lines were never isolated from MCA-treated WT mice [16*]. These results suggest that in the absence of adaptive immunity, innate cells present in *Rag2*^{-/-} mice but absent in *Rag2*^{-/-} × *γc*^{-/-} mice can shape, at least to some degree, the immunogenicity of tumours. In *Rag2*^{-/-} mice, NK cells (presumably activated by local elaboration of endogenous IL-12) can produce IFN-γ that in turn induces activation of CD45⁺CD11b⁺MHCII^{hi}CD206^{lo}Ly6C^{lo} M1 macrophages, which act as important effectors of cancer

immunoediting. These results show that the degree to which a tumour undergoes immunoediting is dependent on the degree of immunocompetence of the host [5,16*]. Further work is needed to define the structures that are recognized on tumour cells by activated M1 macrophages.

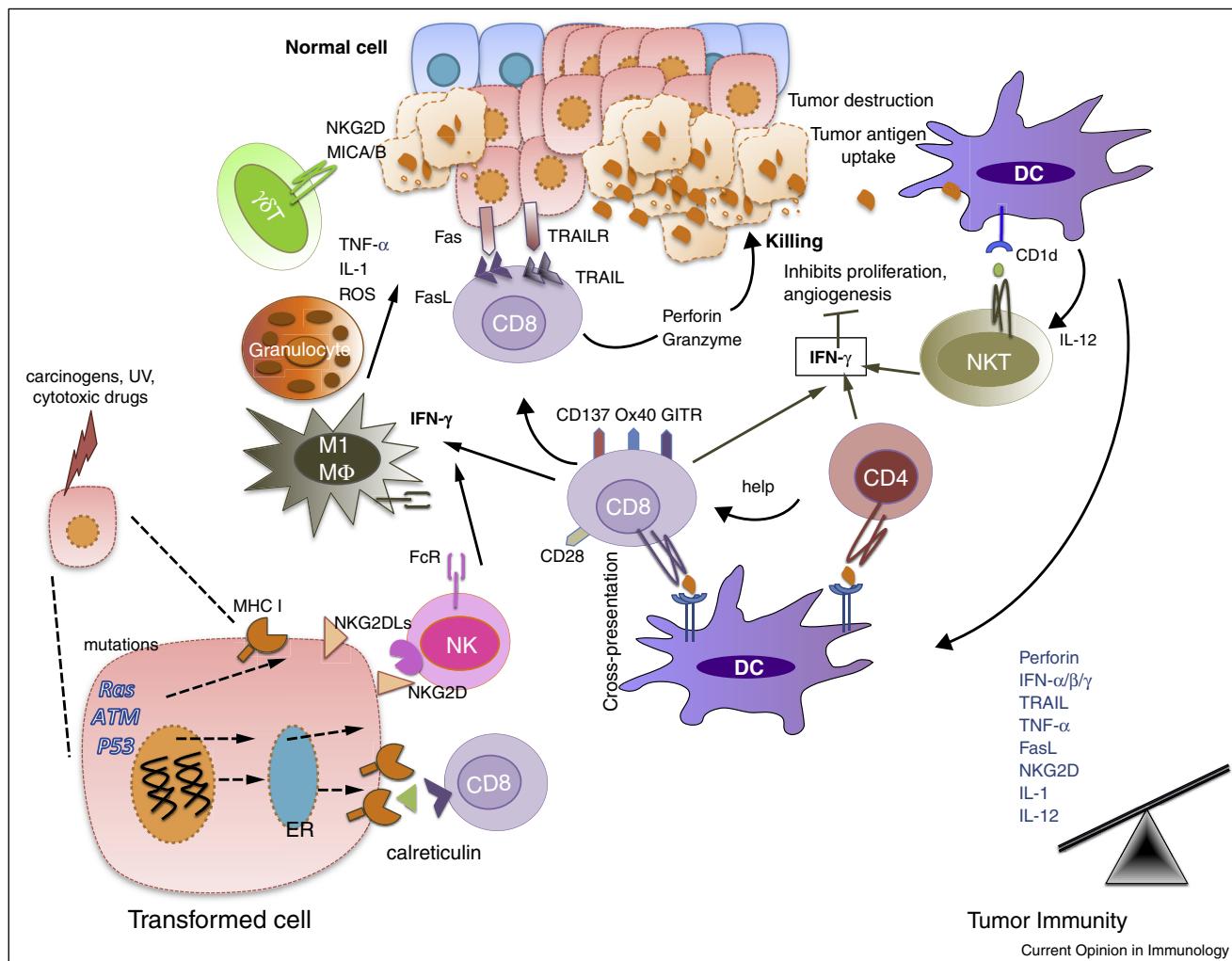
Elimination

Previous reviews have extensively described the studies that support the Elimination phase of cancer immunoediting and have summarized the mechanisms underlying the host protective phase of cancer immunoediting [5,6,17]. Immunodeficient mice develop more carcinogen-induced and spontaneous cancers than wild-type mice, and tumours derived from immunodeficient mice are more immunogenic than those from immunocompetent mice. Here the role of host effector molecules, such as IFN-γ, perforin, Fas/FasL, and TRAIL; recognition molecules such as NKG2D; and an intact lymphocyte compartment in protective anti-tumour immunity, are well recognized [5,6,17,18] (Figure 2). Both type I (IFN-α/β) and type II interferons (IFN-γ) are required for development of anti-tumour immune responses but play distinct roles in the cancer immunoediting process. Whereas IFN-γ targets both tumour cells and hematopoietic cells, IFN-α/β acts primarily on host cells. Two recent studies showed that type I IFNs are required for initiation of the early anti-tumour response and act on CD8α⁺/CD103⁺ DCs to enhance cross-presentation of tumour antigens to CD8⁺ T cells [18,19]. Type I IFN sensitivity in granulocytes, macrophages and NK cells, all of which express type I IFN receptors, was not required for tumour rejection [18].

Another recently published interesting study reported that aneuploidy, and particularly tetraploidization, while contributing to oncogenesis, increases tumour cell immunogenicity leading to immune elimination of these cells. Hyperploid cells have a constitutively active endoplasmic reticulum stress response, resulting in the aberrant cell surface exposure of calreticulin and increased cellular immunogenicity [20]. The immunosurveillance/elimination mechanism for control of ploidy was dependent on CD4⁺ and CD8⁺ T cells, IFN-γ and the IFN-γ receptor which is consistent with other experimental models of immunosurveillance/elimination (reviewed in [17]). These provocative results suggest that hyperploidy in carcinogen-induced and oncogene-induced cancer provides a recognition function for this mechanism of immune elimination of tumours.

Clearly one of the confounding issues in the study of Elimination is that the read-out is the absence of tumours (i.e. a negative result). Thus, there is always a question of whether tumours had originally formed and were eliminated or were never there to begin with. The absences of models that provide positive evidence for elimination have not been forthcoming until recently. Croxford

Figure 2



Elimination is a phase of cancer immunoediting where both the innate and adaptive immune system together detect and destroy early tumours before they become clinically visible. Normal cells (blue) are transformed into tumour cells by carcinogens and other genotoxic insults along with the failure of intrinsic tumour suppressor mechanisms (e.g. p53, ATM). These tumour cells express stress-induced molecules such as surface calreticulin, tumour antigens in context of MHC class I molecules, and/or NKG2D ligands recognized by CD8 $^{+}$ effector cells and NK cells, respectively. DCs can also take up and cross-present tumour antigens to T cells including NKT cells (glycolipid antigens presenting via CD1d). These activated effector cells release IFN- γ that can mediate anti-tumour effects by inhibiting tumour cell proliferation and angiogenesis. CD8 $^{+}$ T cells can induce tumour cell apoptosis by interacting with Fas and TRAIL receptors on tumour cells, or by secreting perforin and granzymes. Effector T cells express co-stimulatory molecules such as CD28, CD137, GITR, OX-40 that enhance their proliferation and survival. $\gamma\delta$ T cells can also recognize and kill tumours expressing NKG2D ligands (MICA/B in humans). Innate immune cells such as macrophages (M1) and granulocytes also contribute to anti-tumour immunity by secreting TNF- α , IL-1, IL-12 and ROS. In the Elimination phase, the balance is towards anti-tumour immunity due to an increase in expression of tumour antigens, MHC class I, Fas and TRAIL receptor on tumour cells and perforin, granzymes, IFN- $\alpha/\beta/\gamma$, IL-1, IL-12, TNF- α in the tumour microenvironment.

et al. have now shown that early-stage B cells in E μ -myc mice are tumourigenic and sharply regress in the periphery between 6 and 9 weeks of age [21 *]. In this model, sustained myc expression induces DNA damage (via the serine/threonine protein kinase ATM) and the DNA damage response can induce ligands that enhance immune recognition. Regression of early-stage transformed B cells was impaired by blockade of DNAM-1, a lymphocyte receptor for one such ligand (CD155), or

deletion of T cells and NK1.1 $^{+}$ cells. Studies of tumour incidence in E μ -myc mice under these conditions remain to be reported.

In addition to conventional elimination mechanisms involving adaptive immunity, new pathways of elimination have been recently reported that rely predominantly on cells of the innate immune system. In a mouse model of liver carcinoma, NK cells were shown to eliminate

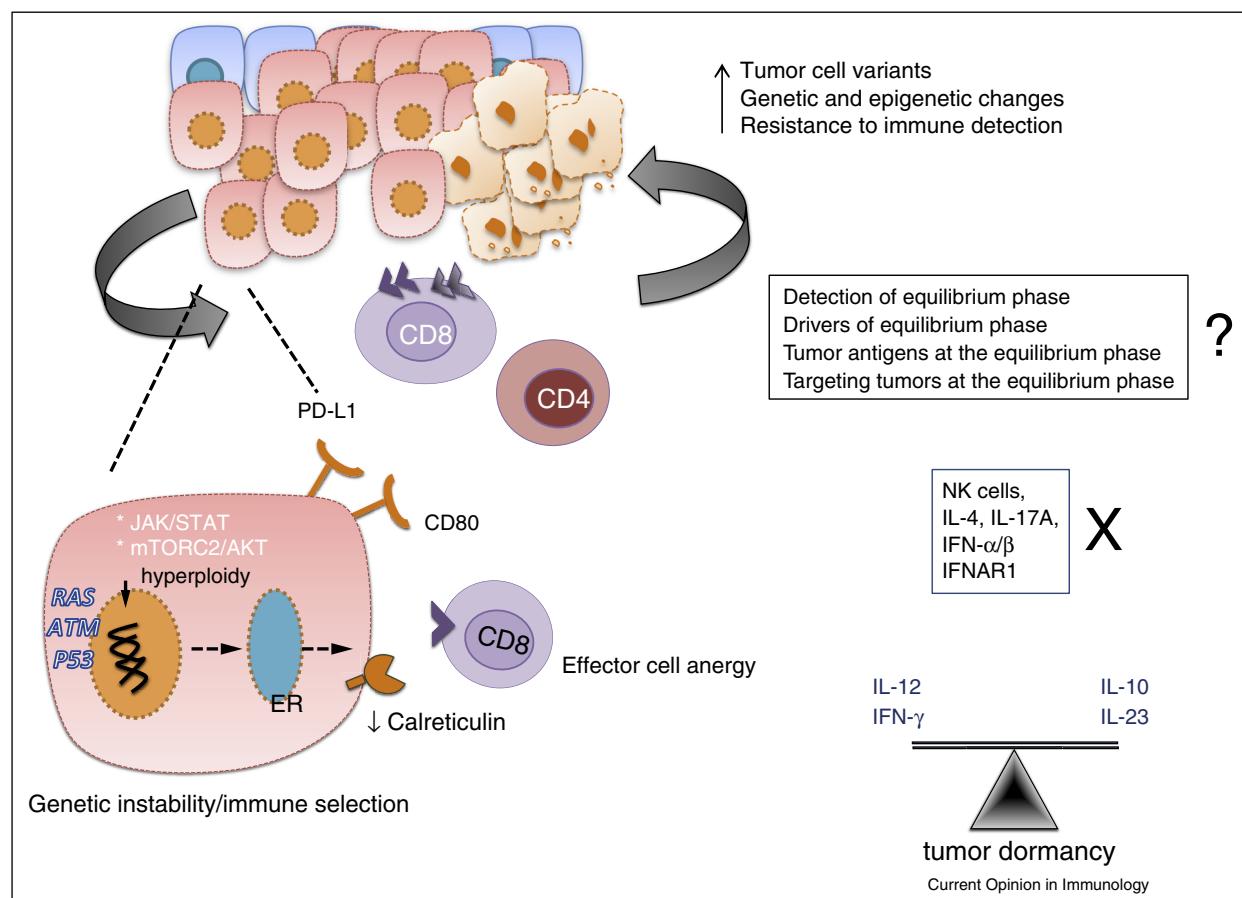
senescent tumour cells in a manner that was dependent on tumour cell intrinsic expression of p53 [22]. Upon expression of p53, tumour cells underwent senescence and secreted various interleukins (IL-6, IL-12 and IL-15) and chemokines such as CCL2 that recruited NK cells to senescent tumours. NK cells eliminated these lesions via mechanisms involving tumour cell expression of NKG2D ligands. NKG2D ligands can be induced on tumour cells by a variety of other stimuli, including Ras signalling and the DNA damage response [23,24]. Similarly, in a model of K-ras induced hepatocellular carcinoma, it was shown that premalignant senescent hepatocytes were cleared by liver-infiltrating K-ras-specific CD4⁺ T cells with the help of macrophages [25]. Impaired surveillance of senescent tissue resulted in liver cancer development [25].

Indirect evidence for an important innate role of NK cells in Elimination has also been uncovered. Dysfunctional telomeres suppress tumour progression by activating cell-intrinsic programs that lead to growth arrest. Increased

levels of TRF2, a key factor in telomere protection, are observed in various human malignancies and contribute to oncogenesis. Recently, it was shown a high level of TRF2 in tumour cells decreased their ability to recruit and activate NK cells [26]. By screening for TRF2-bound genes, it was found that HS3ST4, a gene encoding for the heparan sulphate (glucosamine) 3-O-sulphotransferase 4, was regulated by TRF2 and inhibited the recruitment of NK cells in an epistatic relationship with TRF2. Overall, these results revealed a TRF2-dependent pathway that is tumour-cell intrinsic and regulates the host protective functions of NK cells against cancer development.

Another recent series of studies point to a possible, druggable role for macrophages in innate tumour elimination. Tumour cells have been shown to constitutively express CD47 that functions as a 'don't eat me' signal upon interacting with the SIRP α inhibitory receptor on macrophages. Monoclonal antibody blockade of CD47 led to phagocyte dependent elimination of tumour cells

Figure 3



In the Equilibrium phase of cancer immunoediting, the immune system holds the tumour in a state of functional dormancy. Some tumour cells undergo genetic and epigenetic changes and due to constant immune pressure, tumour cell variants evolve that resist immune recognition (antigen loss or defects in antigen-presentation) and induce immunosuppression (PD-L1). The Equilibrium phase is a balance between anti-tumour (IL-12, IFN- γ) and tumour promoting cytokines (IL-10, IL-23). The adaptive immune system is required to maintain tumour in a functionally dormant state while NK cells and cytokines such as IL-4, IL-17A and IFN- α/β are dispensable.

[27]. However, since red blood cells also express CD47, such a treatment could potentially lead to catastrophic anemia. Therefore, as an alternative approach, high affinity soluble human SIRP α variants were designed and were shown in various mouse cancer models to inhibit the interaction of macrophage SIRP α with tumour cell expressed CD47 [28*]. These variants induced no toxicity and acted synergistically with tumour-specific therapeutic antibodies (acting via antibody-dependent cellular cytotoxicity) in mouse models of lymphoma and HER2 $^+$ breast carcinoma.

Equilibrium

The molecular mechanisms that trigger immune-mediated tumour dormancy (Equilibrium phase) are poorly understood because this phase has been difficult to model in mice and have been described only anecdotally in humans (reviewed in [5]). A landmark study in 2007 demonstrated the role of adaptive Th1-like immunity in immune-mediated dormancy of fibrosarcoma [29]. A follow up study using the same mouse model of MCA-induced fibrosarcoma and p53 mutant tumours has shown that immune-mediated tumour dormancy may be a very prolonged process [30]. Importantly, the balance of IL-12 promoting elimination, and IL-23 (sharing the common subunit IL-12p40) promoting persistence, maintains tumours in equilibrium (Figure 3) [30]. Although a minor tumour-promoting role for IL-10 was also uncovered, many other pathways (e.g. IL-4, IL-17A, TNF, IFN- α/β) were shown to be dispensable for this phase.

Another recent report compared the cellular environment of tumours in equilibrium versus those that escape and found high proportions of CD8 $^+$ T cells, NK cells, $\gamma\delta$ T cells and low proportions of NKT cells, Foxp3 $^+$ Treg cells, and MDSCs were associated with maintaining occult cancer in an immune-mediated equilibrium state [31]. This study further supports the concept that the relative balance of immunosuppressive cells and immune cells capable of manifesting antitumour effector functions in the tumour microenvironment is associated with maintaining tumour cells in a state of immune-mediated dormancy. It remains to be determined what shifts the balance in favor of immune escape or elimination following equilibrium but since editing occurs in this phase, it is likely that evolution of tumour cell variants that either lack key tumour specific antigens or the capacity to present them is an important factor that contributes to the decision process.

Additional support for the occurrence of the Equilibrium phase comes from the finding that tumour antigen-specific T cells can arrest the growth of experimentally induced pancreatic tumours (Tag-induced multistage carcinogenesis) in mice by a coordinated interaction between IFN- γ and TNF [32]. In the absence of either TNFR or IFN- γ , the same T cells promoted angiogenesis

and multistage carcinogenesis. It was further shown that, the combination of IFN- γ and TNF drive Tag-expressing cancers into senescence by inducing permanent growth arrest in G1/G0, activation of p16INK4a, and downstream Rb hypophosphorylation at Ser795 [33]. This cytokine-induced senescence requires STAT1 and TNFR1 signaling in addition to p16INK4a. Since IFN- γ and TNF induce senescence in numerous murine and human cancers, this may be a general mechanism for arresting cancer progression.

Identification of occult cancer in equilibrium in mice and humans remains a significant technical challenge, but advances in imaging technology with suitable antigen markers may allow circulating tumour cells and niches to be explored further.

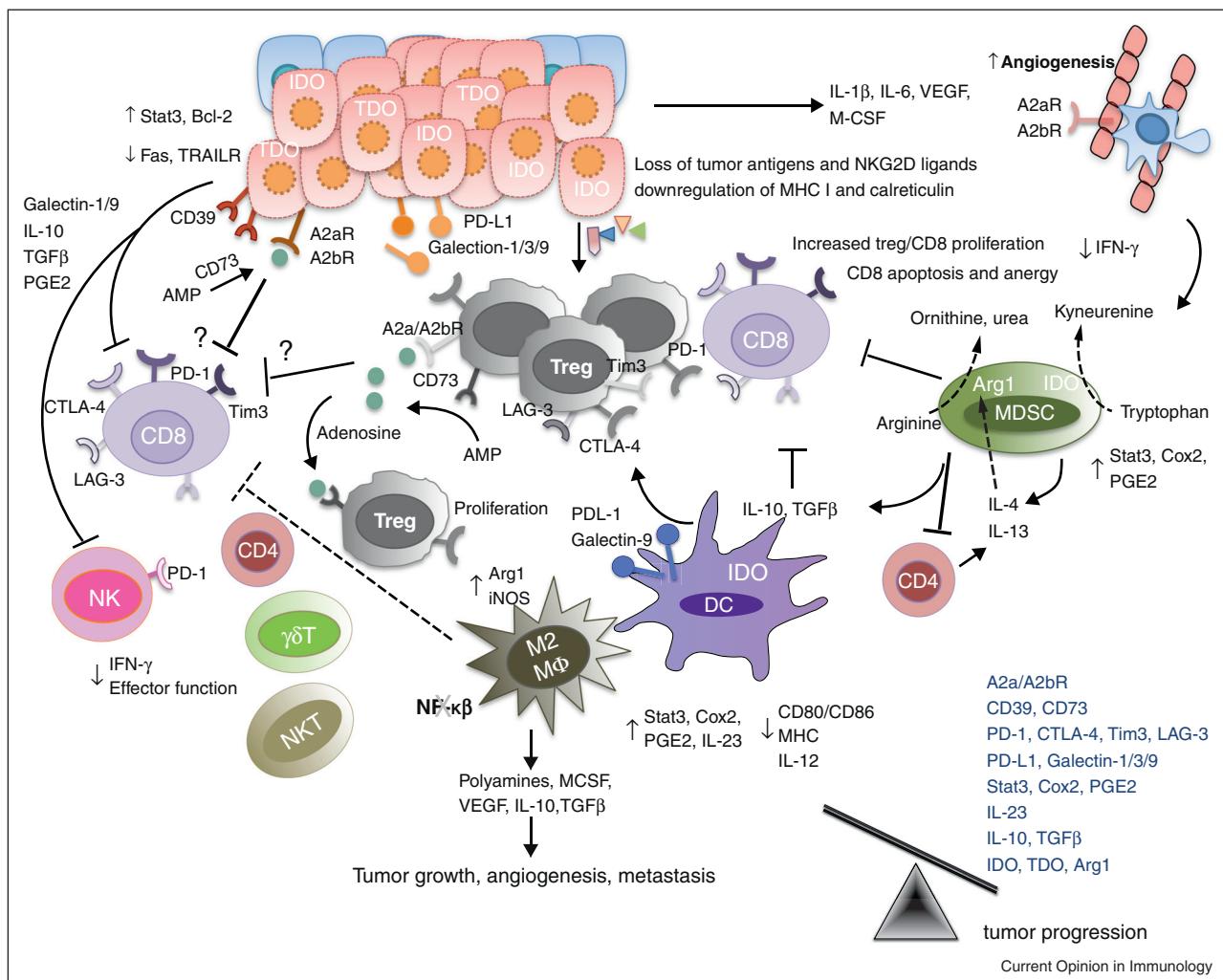
Escape

Tumour cell escape can occur through many different mechanisms including: reduced immune recognition (such as the absence of strong tumour antigens, or loss of MHC class I, class I-like, or co-stimulatory molecules), increased resistance or survival (such as increased expression of STAT-3 or anti-apoptotic molecule Bcl2), or development of an immunosuppressive tumour microenvironment (cytokines such as VEGF, TGF- β ; immunoregulatory molecules such as IDO, PD-1/PD-L1, Tim-3/galectin-9, LAG-3). These mechanisms have been extensively discussed elsewhere [5,6,17,34–36] and are summarized in Figure 4. In the past few years, we have witnessed a growing list of new moieties that contribute to tumour-induced immunosuppression, such as CD73 [37], adenosine receptors [38], and new B7 family checkpoint molecules including VISTA [39,40] and BTLA [41,42].

Targeted immunotherapies based on immune escape mechanisms

Immunotherapy has recently emerged as a viable and potentially transformative approach to cancer treatment. However, therapeutic intervention often fails because of the plasticity of cells within the tumour microenvironment. Thus, approaches that involve combinations that target multiple pathways may prove synergistic and often are capable of generating a systemically effective memory response [43*]. Many preclinical approaches have been extensively reviewed and a number of promising approaches are soon to enter the clinic [34,44,45]. In particular, the recent clinical combination of Ipilimumab (anti-CTLA-4) [46] and Nivolumab (anti-PD-1) [47,48] may have revolutionized thinking about the treatment strategy for melanoma patients [49**]. This combination has shown responses in almost half of the metastatic melanoma patients, for which conventional therapies have failed [49**]. The use of combination therapies provides increased opportunities for more effective beneficial clinical effects but also carry an increased risk

Figure 4



During the Escape phase of cancer immunoediting, the immune system fails to restrict tumour outgrowth and tumour cells emerge causing clinically apparent disease. In this phase, tumour cells evade immune recognition (loss of tumour antigens, MHC class I or co-stimulatory molecules), express molecules of increased resistance (STAT-3), survival (anti-apoptotic molecule bcl2) and immunosuppression (IDO, TDO, PD-L1, galectin-1/3/9, CD39, CD73, adenosine receptors) and secrete cytokines VEGF, TGF- β , IL-6, M-CSF that enhance angiogenesis. Furthermore, MDSCs, M2 macrophages and DCs may also express immunoregulatory molecules such as arginase, iNOS and IDO and secrete immunosuppressive cytokines IL-10 and TGF- β that can inhibit CD8 $^+$ proliferation or induce apoptosis. MDSCs and IDO expressing DCs also induces the generation of regulatory T cells. IDO, arginase, CD39 and CD73 are immunoregulatory enzymes whereas IDO catabolize tryptophan to kynureneine, arginase catabolize L-arginine to ornithine and urea, CD39 metabolise ATP to AMP which can further be metabolised to adenosine by CD73. Adenosine can bind to adenosine receptors – A2aR and A2bR expressed on tumour cells, endothelial cells and immune cells. T cells including Tregs may express inhibitory receptors such as PD-1, CTLA-4, Tim-3 and LAG3 that suppresses anti-tumour immune response and favor tumour outgrowth. In the Escape phase, the balance is skewed towards tumour progression due to the presence of immunosuppressive cytokines and molecules such as IL-10, TGF- β , VEGF, IDO, PD-L1.

of immunopathology. Therefore, it will be important in the future to seek a balance between tumour immunity and inflammatory pathology [50]. Tumour immunity and inflammatory pathology are closely related, but nonetheless separable and pre-clinical models that can tease out the mechanisms underlying this double edged sword of combination therapies will be invaluable for determining which combinations provide an increased therapeutic index.

Immune cells in tumours: predictive and prognostic significance

One significant advantage of measurable anti-tumour effects of immunotherapies in patients is our emerging capability to undertake genomic studies of tumours and the host to define key parameters that differentiate between responders from non-responders, and develop new approaches to stratify patients and their tumours. We have witnessed a tremendous explosion in the identifi-

cation of immune signatures for various types of human cancer [51–54]. Similar signatures are observed within cancers with a better prognostic outcome, and cancers with an increased likelihood to respond to therapy or undergo complete regression. The parameters establishing the immune contexture are comprised of the density of CD3⁺, CD8⁺, and CD45RO⁺ T cells and their location at the tumour center and invasive margin combined with the quality of the tertiary lymphoid islets in the affected organ (the 'Immunoscore'). These features are associated with an activated Th1 signature, including IFN- γ , STAT1, IL-12, IRF1, T-bet, perforin, granzymes, CXCR3 and CCR5 ligand chemokines, CXCL9, CXCL10 and CCL5, and adhesion molecules (MADCAM1, ICAM1 and VCAM1) [52]. There are many examples where recent work in the field has suggested that tumour infiltrates can be successfully used as a prognostic biomarker to predict the outcome of treatment [55,56]. Furthermore, in some cancers, these features have been found to be a more powerful prognostic indicator for tumour staging than previous pathological criteria. Other signatures have emerged from patients receiving IL-2 [57], MAGE-A3 vaccination [58], and Ipilimumab [59], and many targeted and conventional therapies also display similar lymphocyte signatures [60–62].

Role of microbiota in tumour growth and response to therapy

Another recent, clinically relevant finding concerning the interaction of immunity with cancer is the emerging role of the host microbiota during tumour formation and progression. Infiltrating Th1 cells and CD8⁺ cytotoxic T cells constitute a positive prognostic sign in colorectal cancer. By contrast, myeloid cells and Th17 cells promote tumourigenesis, and a Th17 expression signature in stage I/II colorectal cancer is associated with a drastic decrease in disease-free survival [63]. Many epithelial cancers develop proximally to microbial communities, which are only physically separated from immune cells by an epithelial barrier. Barrier deterioration induced by colorectal-cancer-initiating genetic lesions results in adenoma invasion by microbial products that trigger tumour-elicited inflammation, which in turn drives tumour growth. IL-23 mainly produced by tumour-associated myeloid cells that are likely to be activated by microbial products, is key in this process [64[•]]. More recently, additional studies have demonstrated the importance of the host microbiome to carcinogenesis and tumour response to therapy [65].

Conclusion – the influences of cancer immunoediting on cancer immunotherapy

Recognizing cancer immunoediting and understanding the mechanisms that underpin it have provided the justification for many new immune-based cancer treatments. Some of these treatments are demonstrating remarkable responses in cancer patients, alone or in combination. In cancer, the immune system is not ignorant of the presence

disease, but rather is actively suppressed by it. The challenge now lies in determining which patients are most suitable to receive these immunotherapies and how we can use information about their tumours and tumour microenvironments to inform us about the most effective treatments on a personalized basis.

In spite of the recent success of immunotherapy in various human cancers, central questions remain unanswered. Even though we now have a better understanding of the mechanisms of tumour escape and equilibrium, questions still remain as to why some tumours escape immune control while others do not. Is the strength of the TCR response to antigen central in determining whether nascent cells are eliminated and why do some tumour clones further progress into equilibrium and escape? What has occurred in a large proportion of patients who have no obvious or apparent immune reaction with their cancer? Did they ever develop one? If not, can one be engineered? In those patients with an immune reaction, what is the simplest combination of therapies to achieve disease free survival? Will we eventually be able to develop personalized cancer immunotherapies designed specifically for an individual cancer patient and their individual tumours? Surely, with the recent explosion in our understanding of cancer immunogenomics and our rapidly expanding recognition of and ability to manipulate immune components that positively and negatively affect tumour immunity, we can expect significant improvements in the next few years in cancer treatment in general and cancer immunotherapy in particular.

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