

MHC and Antigen Presentation

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GSK Course, February 19, 2025

T cells recognize antigens in the form of short peptides bound to molecules encoded within the major histocompatibility gene complex (MHC)

Peptide binding to MHC class I and MHC class II molecules

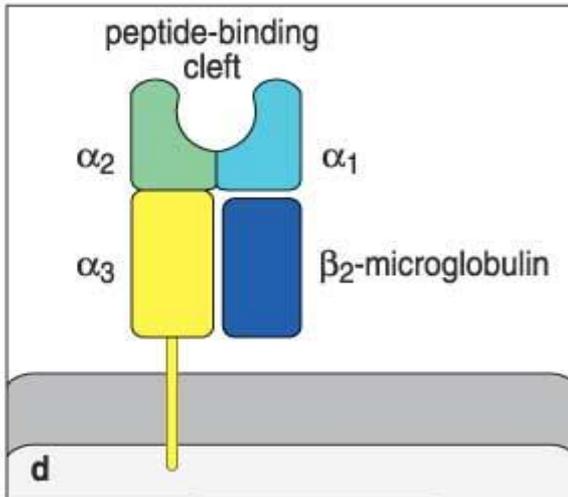
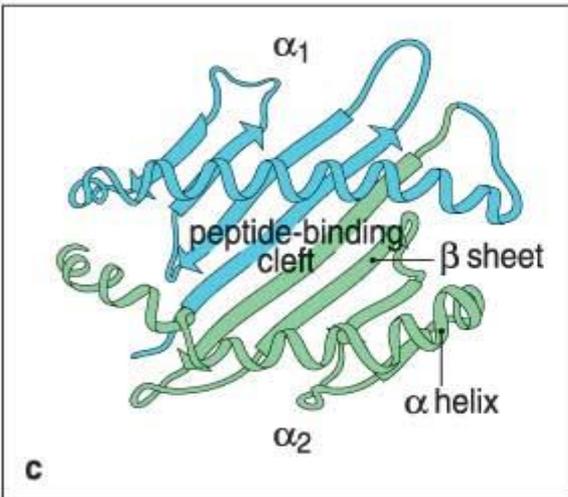
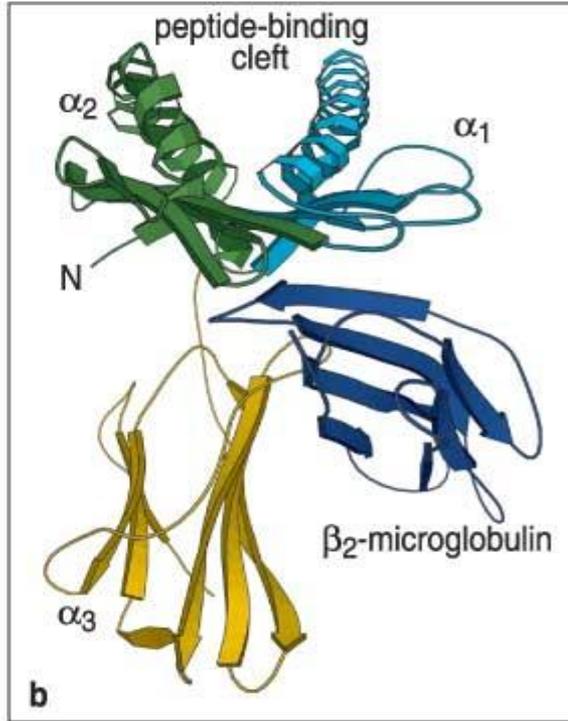
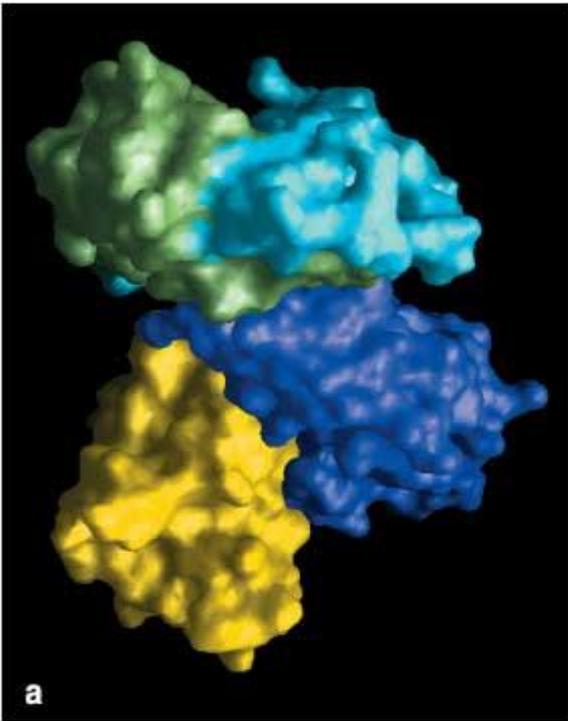
Two major pathways of antigen presentation sample two main sites of protein turnover: cytosol and endo-lysosomes

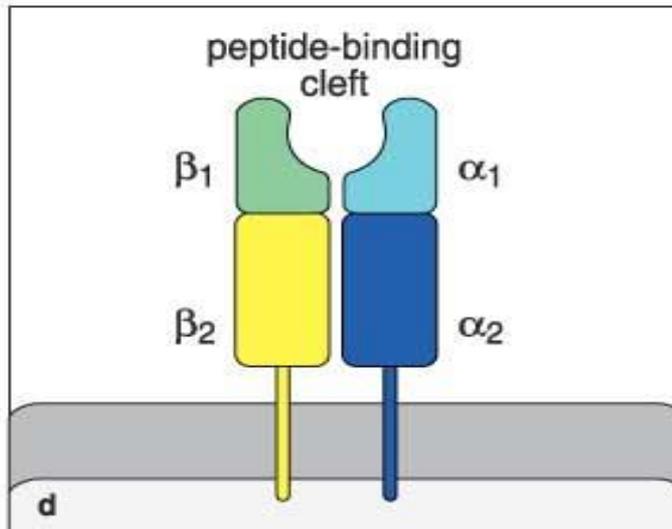
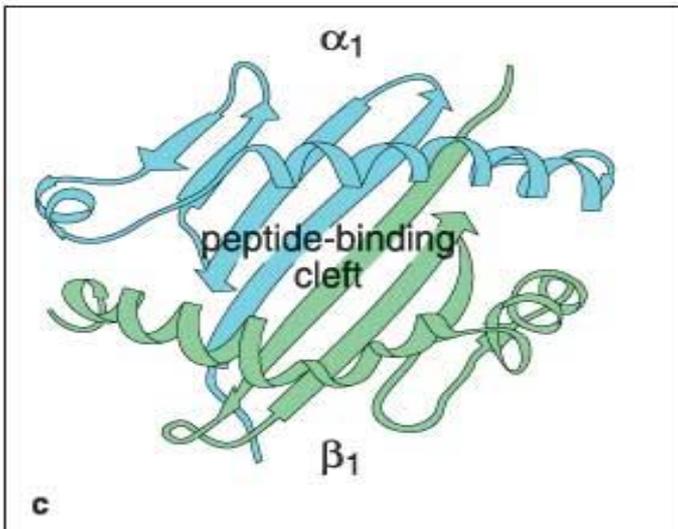
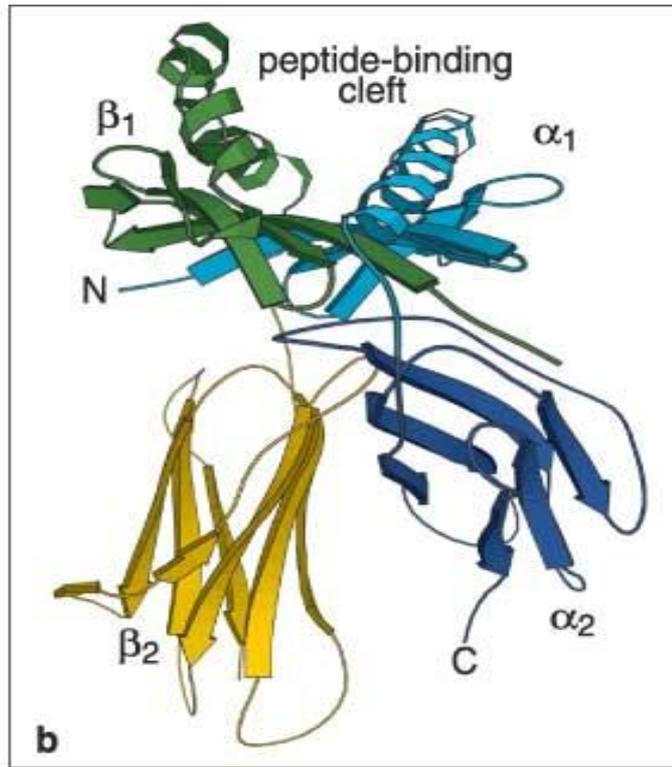
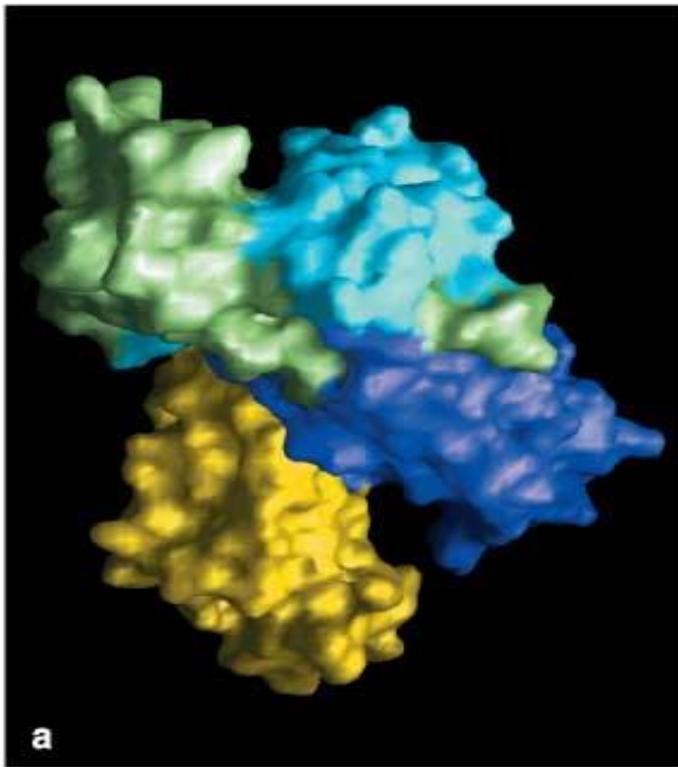
Endosomal antigen presentation by MHC class II molecules

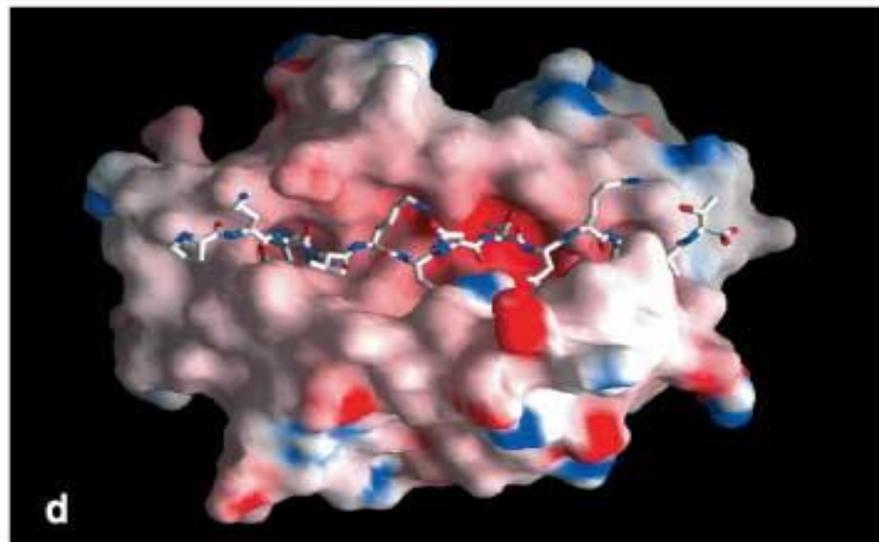
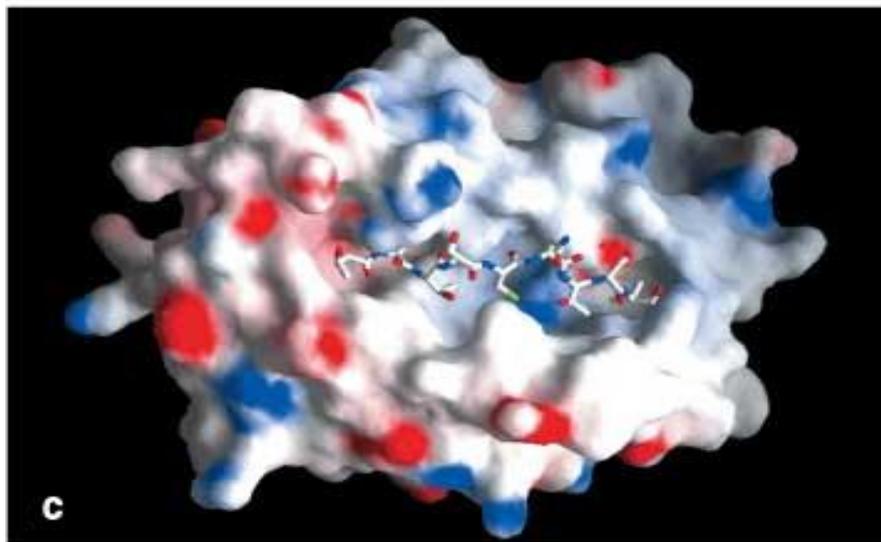
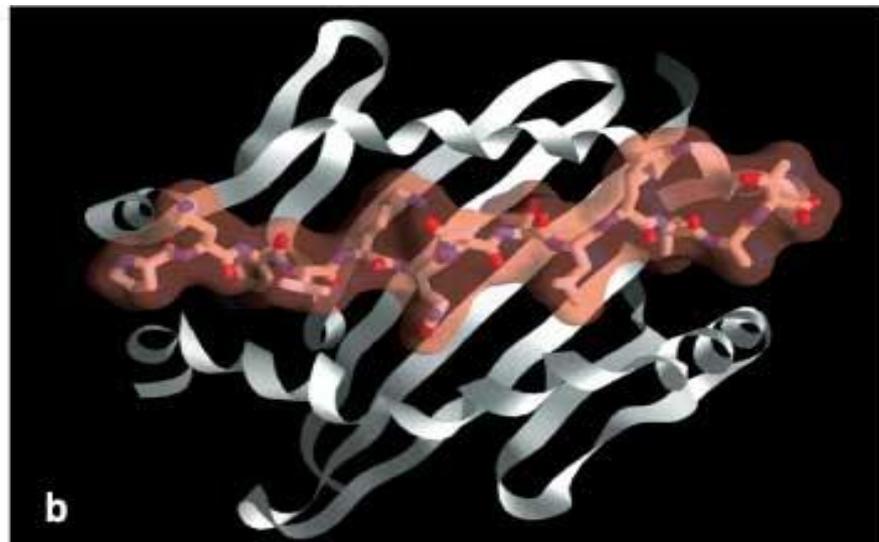
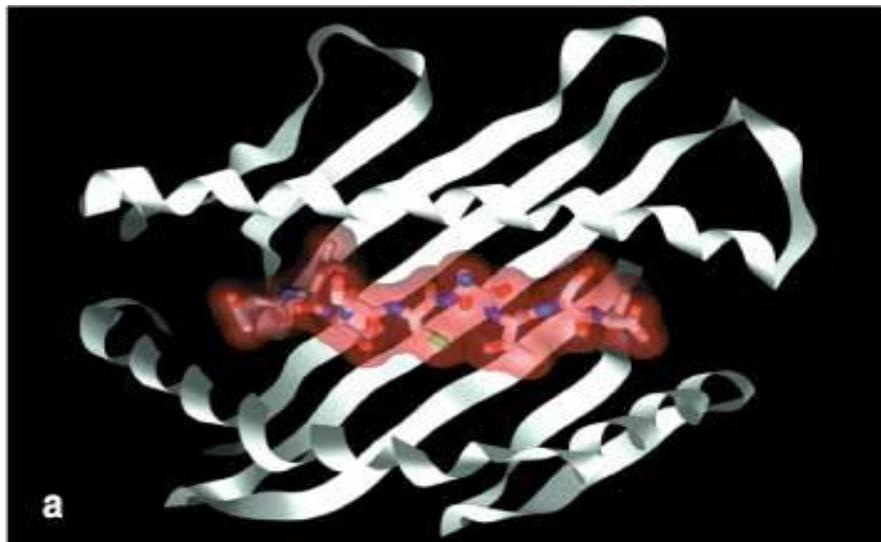
Cytosolic antigen presentation by classical MHC class I molecules

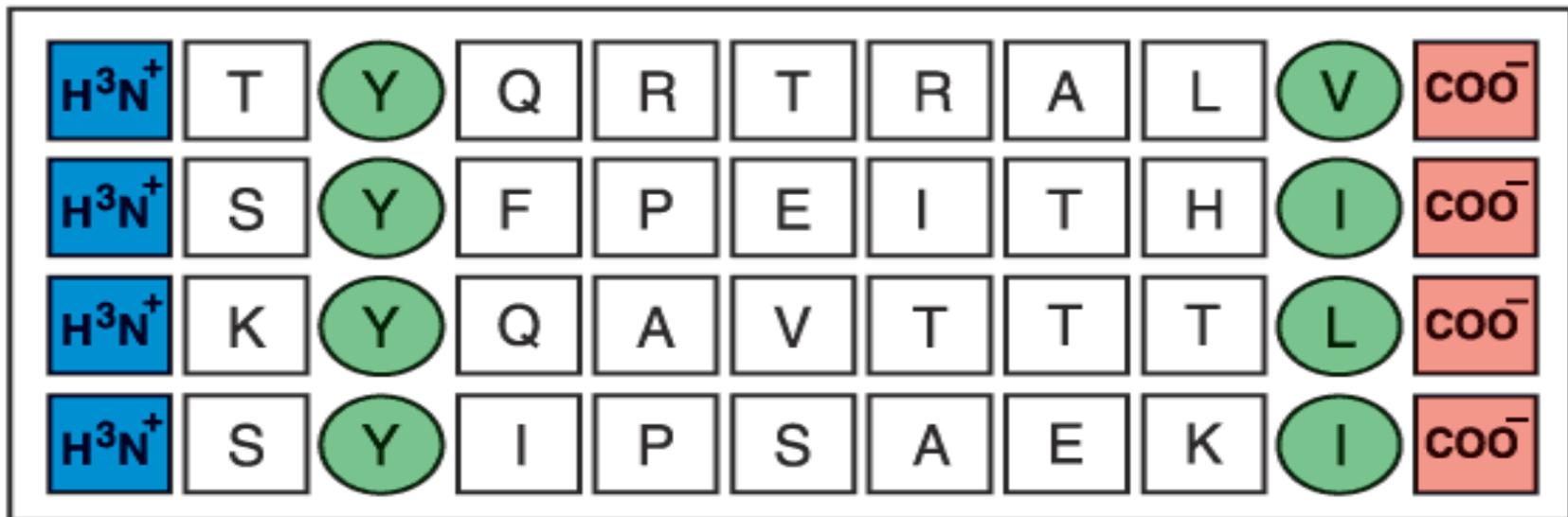
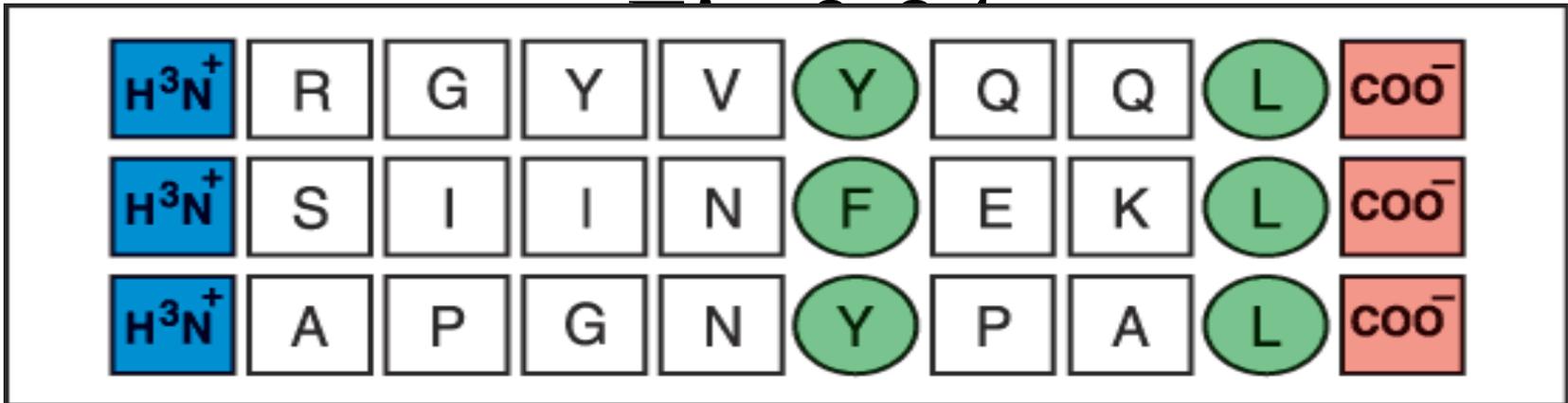
Viral mechanisms to evade antigen presentation by MHC class I molecules

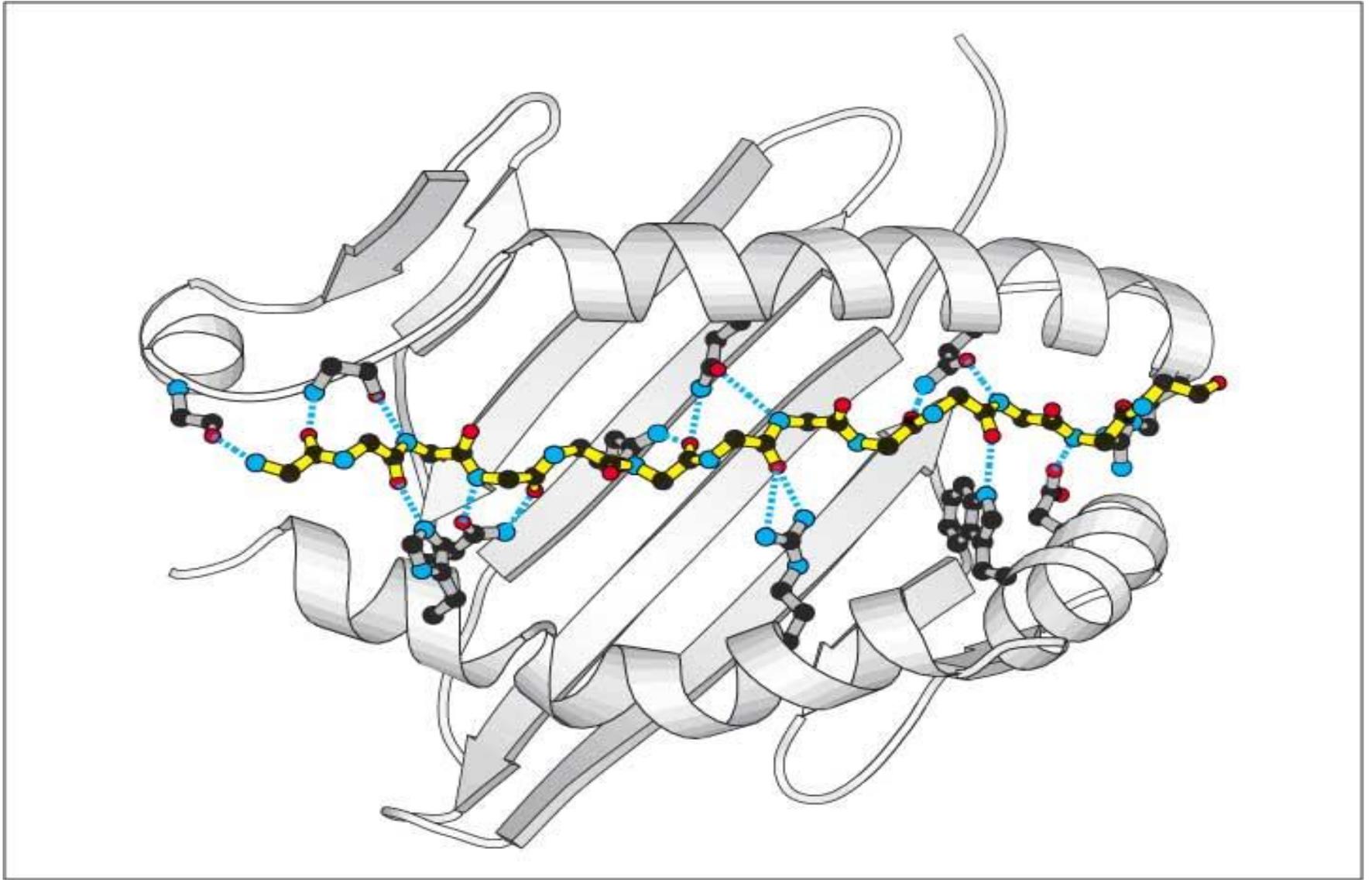
Organization and genetics of MHC locus











MHC class I

**Transmembrane
MHC-encoded α
chain and soluble β 2-m**

Short cytoplasmic tail

“Closed” groove

8-10 aa peptides

Ligands for CD8 T cells

Short-lived

MHC class II

**Transmembrane
MHC-encoded α
and MHC-encoded β chain**

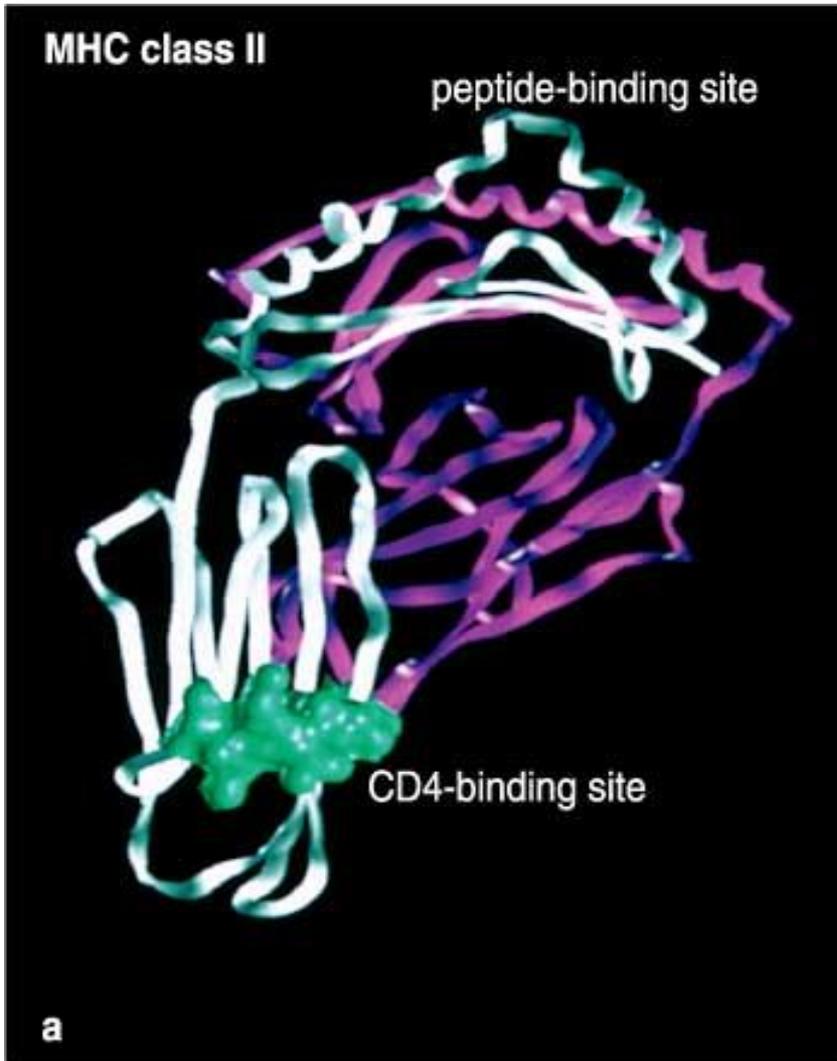
Short cytoplasmic tail

“Open” groove

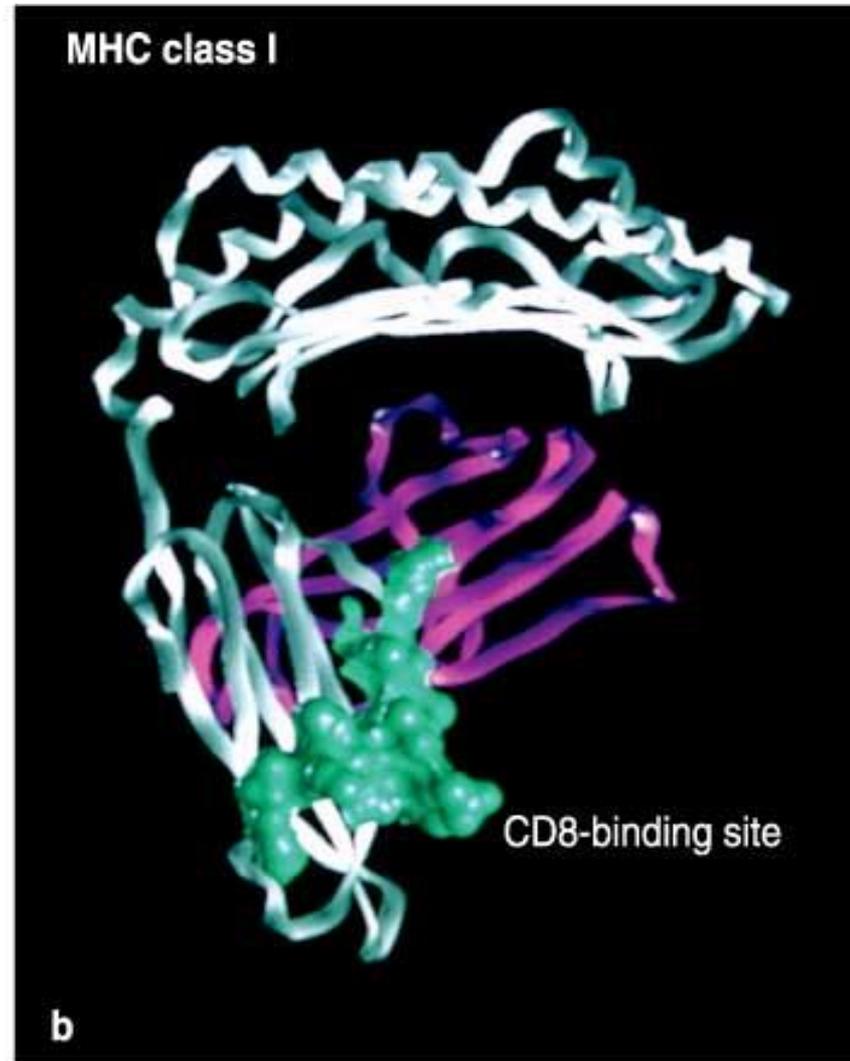
12-17 aa peptides (or longer)

Ligands for CD4 T cells

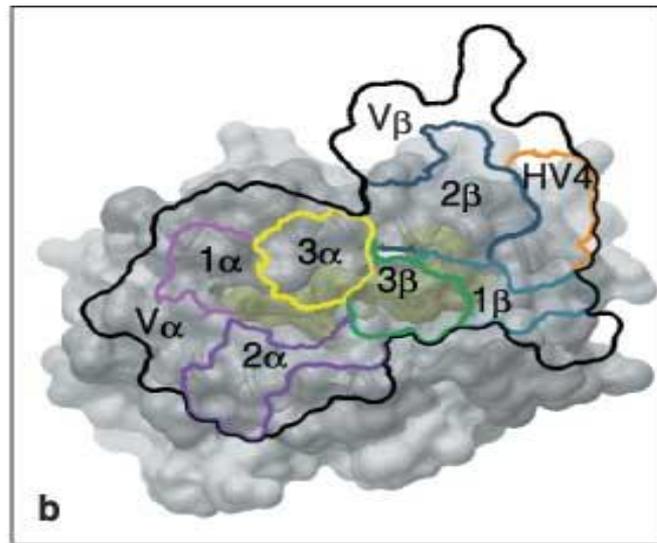
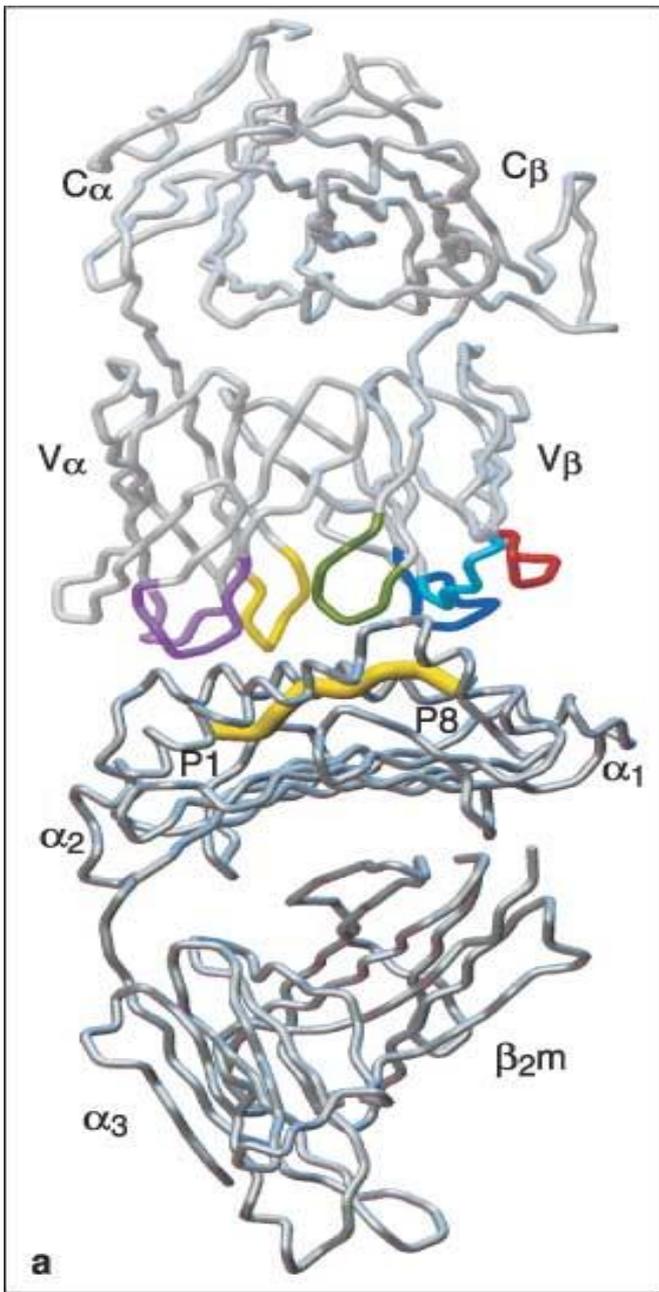
Long-lived



β 2 domain



α 3 domain



MHC class II presentation of viral and bacterial superantigens

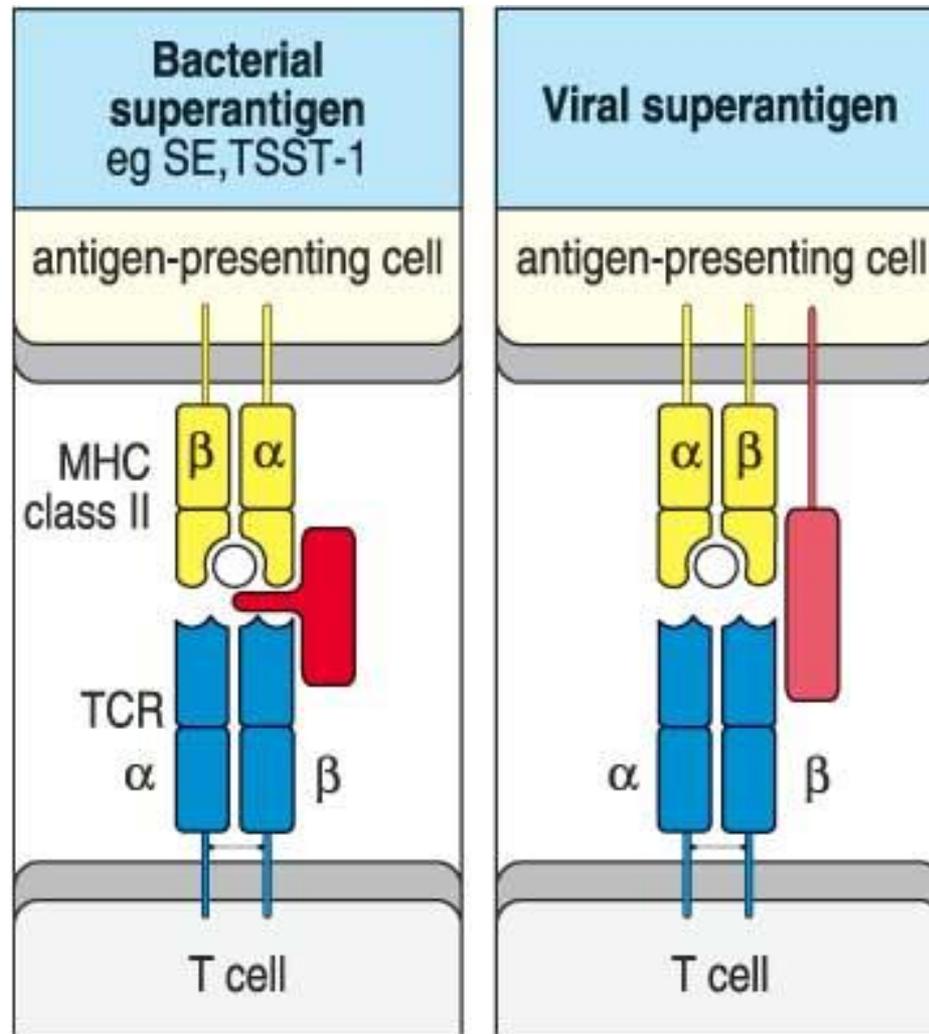
Viral superantigens (vSAG) first discovered as *minor lymphocyte stimulating antigens* or MIs; endogenous mammary tumor viruses

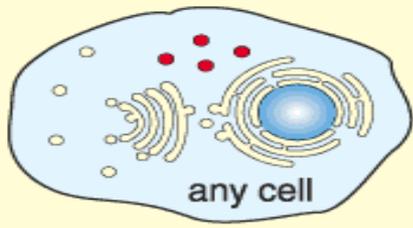
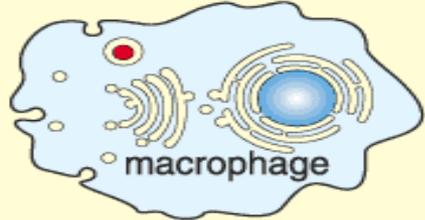
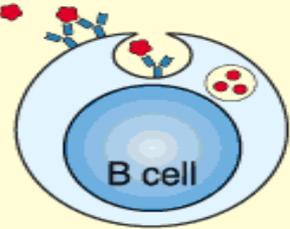
Bacterial superantigens discovered as potent T cell mitogens inducing toxic shock [staphylococcal enterotoxin B and A (SEB and SEA), toxic shock syndrome toxin-1 (TSST-1)]

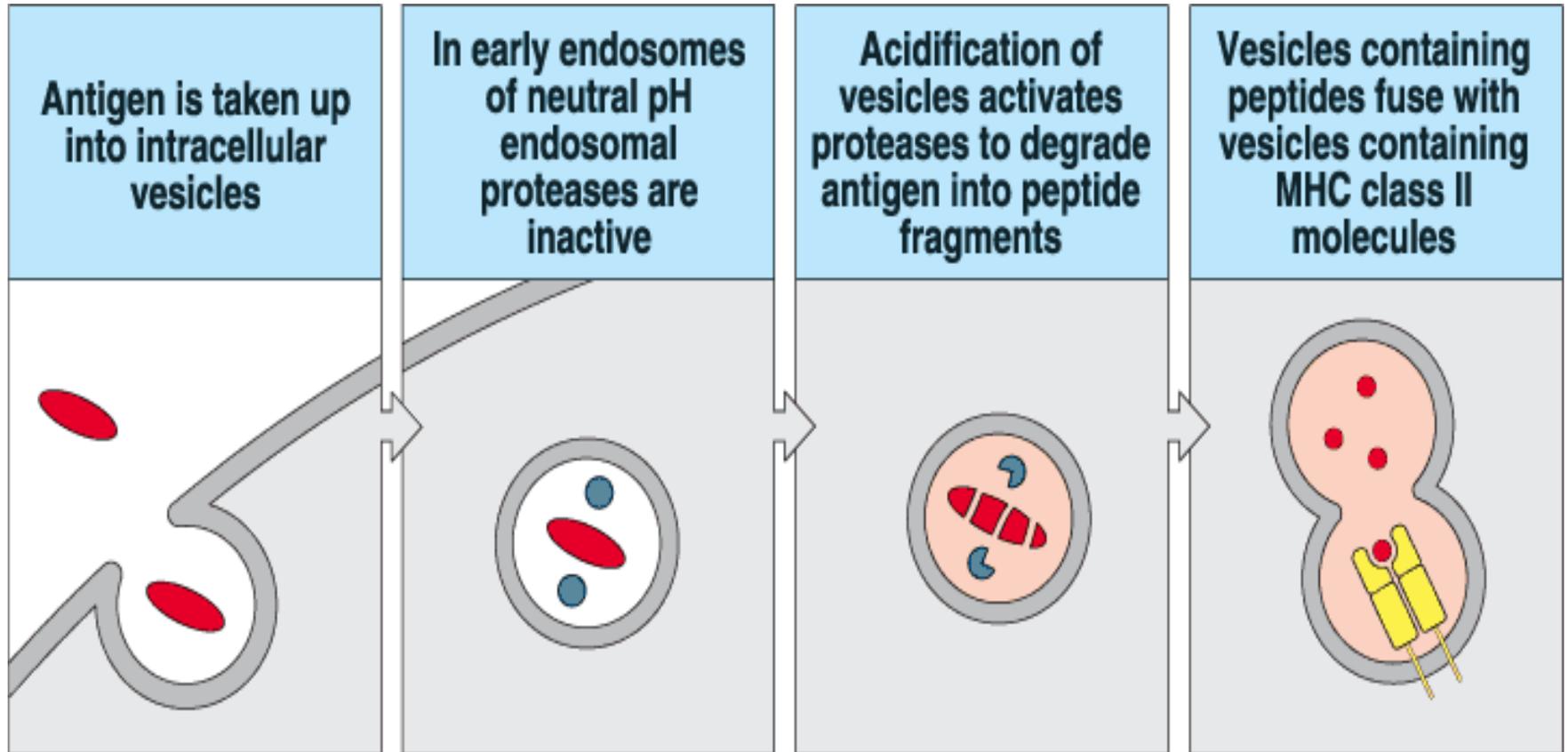
Do not require proteolytic processing

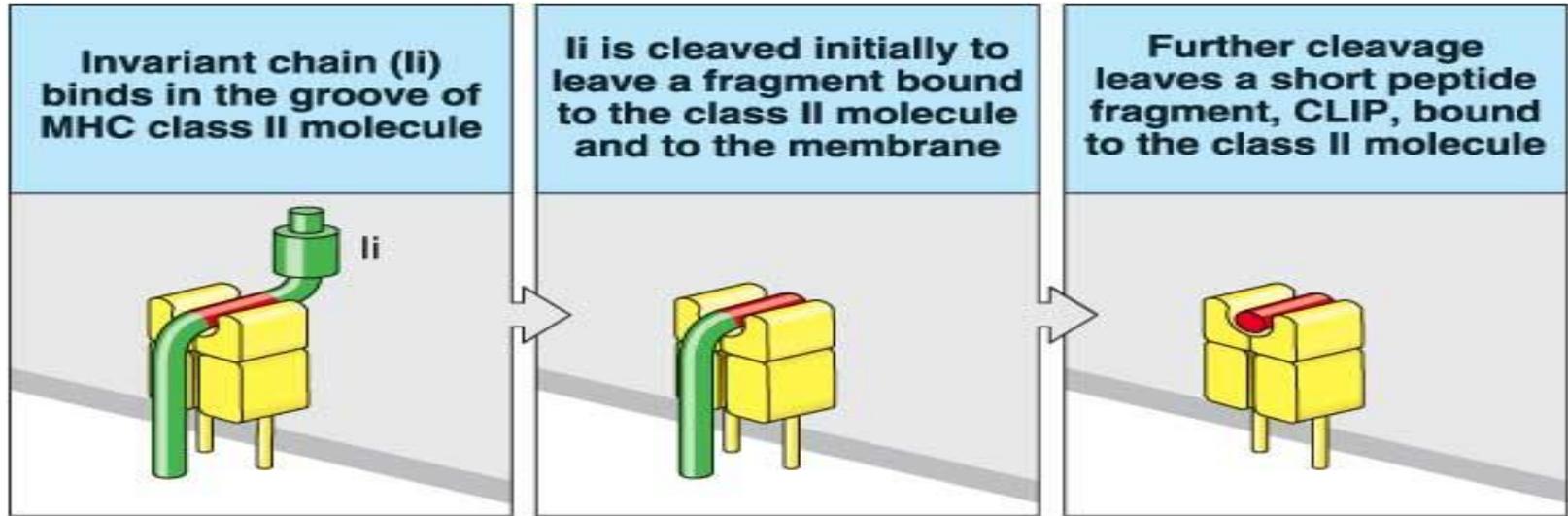
Interact with relatively conserved regions of MHC class II molecules and a particular V β element (e.g. V β 8 or V β 3)

Activate large number of T cells (all T cells expressing corresponding V β).



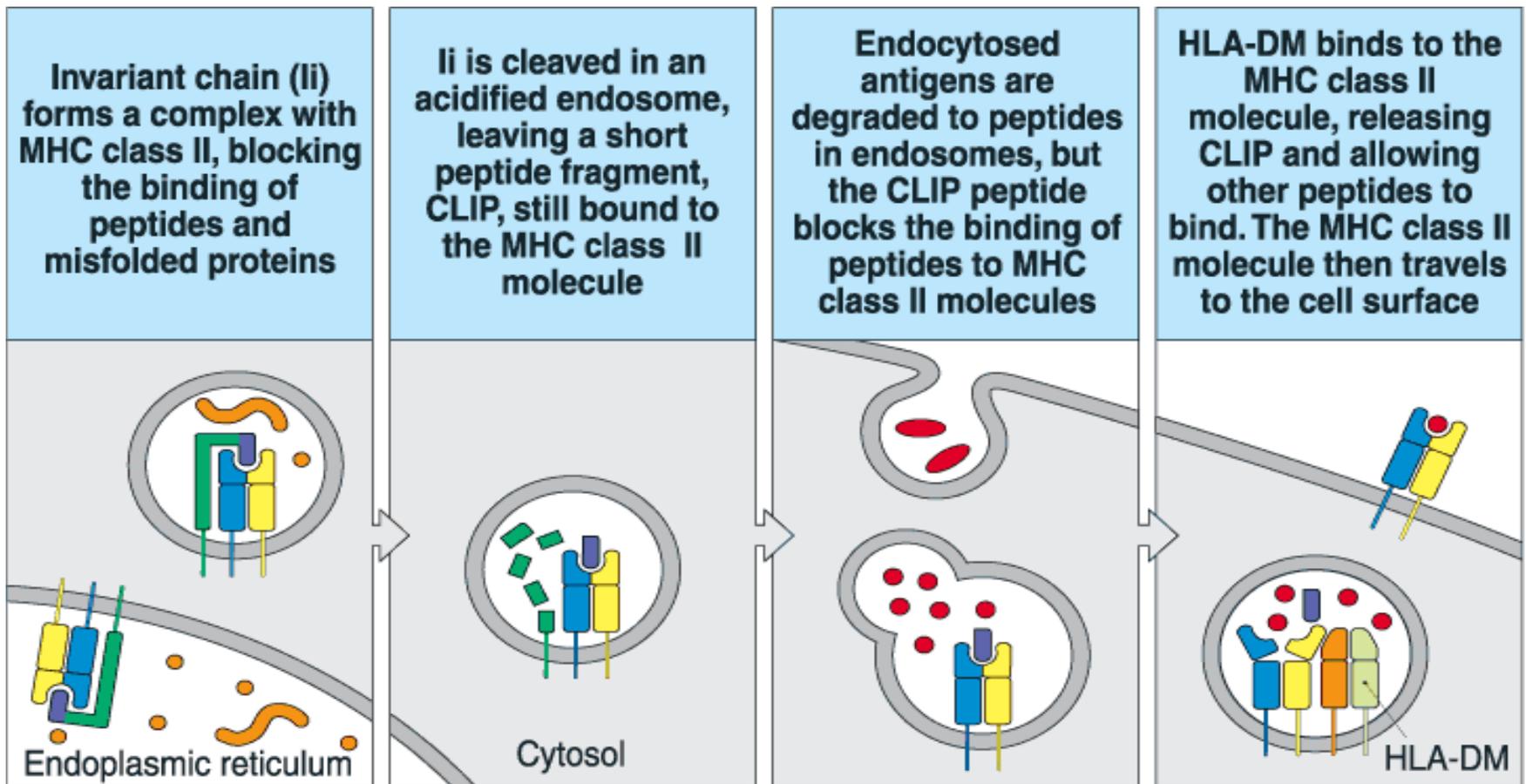
	Cytosolic pathogens	Intravesicular pathogens	Extracellular pathogens and toxins
	 any cell	 macrophage	 B cell
Degraded in	Cytosol	Endocytic vesicles (low pH)	Endocytic vesicles (low pH)
Peptides bind to	MHC class I	MHC class II	MHC class II
Presented to	CD8 T cells	CD4 T cells	CD4 T cells
Effect on presenting cell	Cell death	Activation to kill intravesicular bacteria and parasites	Activation of B cells to secrete Ig to eliminate extracellular bacteria/toxins





p33
 (p35)
p41
 (p43)





HLA-DO/H-2O

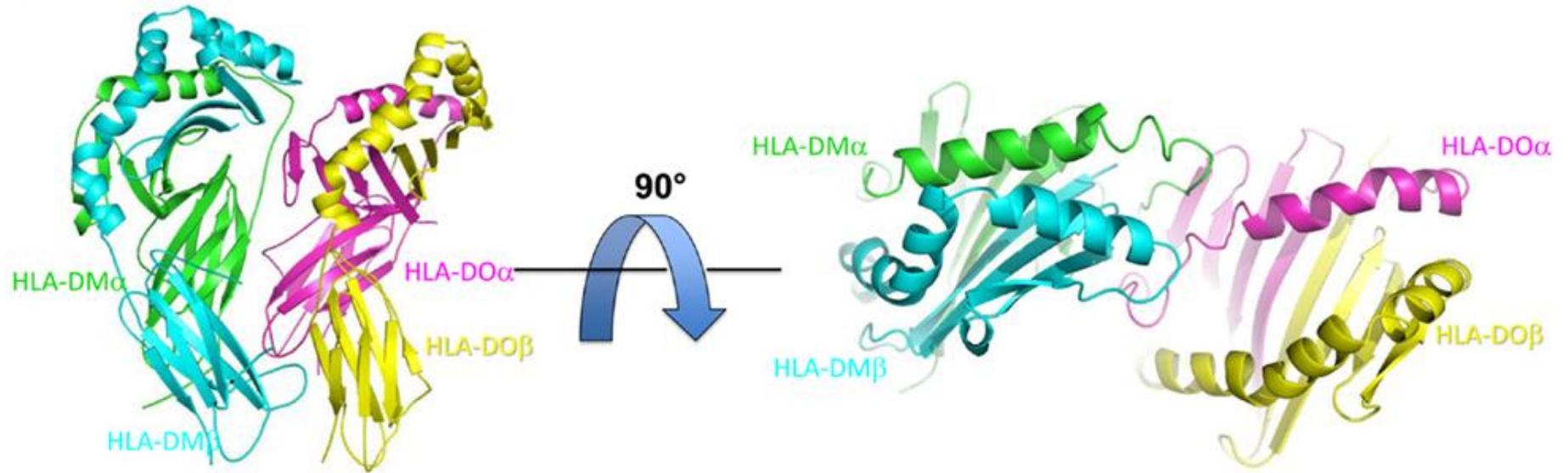
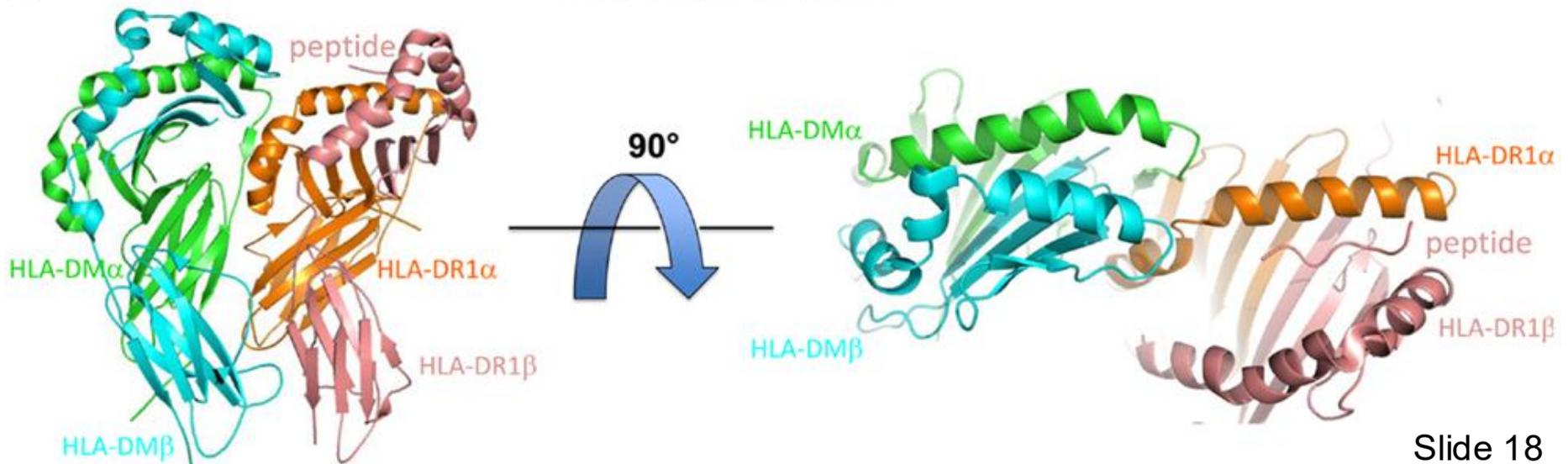
Expressed in B cells and mTEC

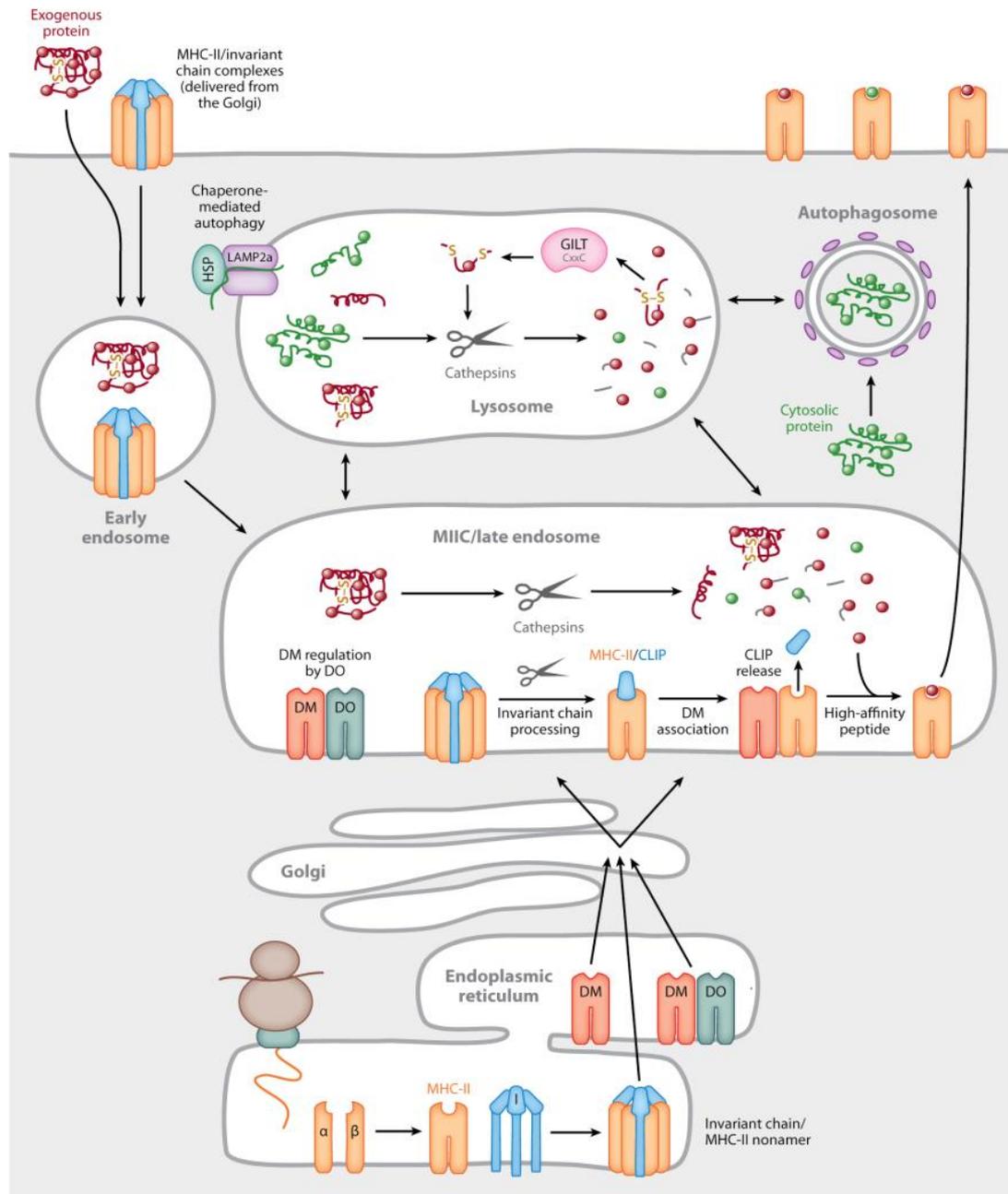
Binds DM

Inhibits DM function in a pH-dependent manner

Free DM is localized to internal vesicles of multi-vesicular bodies

DM-DO are localized to the outside membrane

A**HLA-DM-HLA-DO****B****HLA-DM-HLA-DR1**



Lysosomal proteases:

AEP/legumain/?
(Manoury et al., 2003)

Cathepsin S and L
(Nakagawa et al., 1998;1999)

SPPL2a
(Schneppenheim et al., 2013;
Beisner et al., 2013)

Lysosomal thioreductases:
GILT
(Maric et al., 2001)

MHC class I-like CD1 molecules present glycolipids to NKT cells (and CD8 T cells in humans)

CD1d (conserved), CD1b,c,d (human)

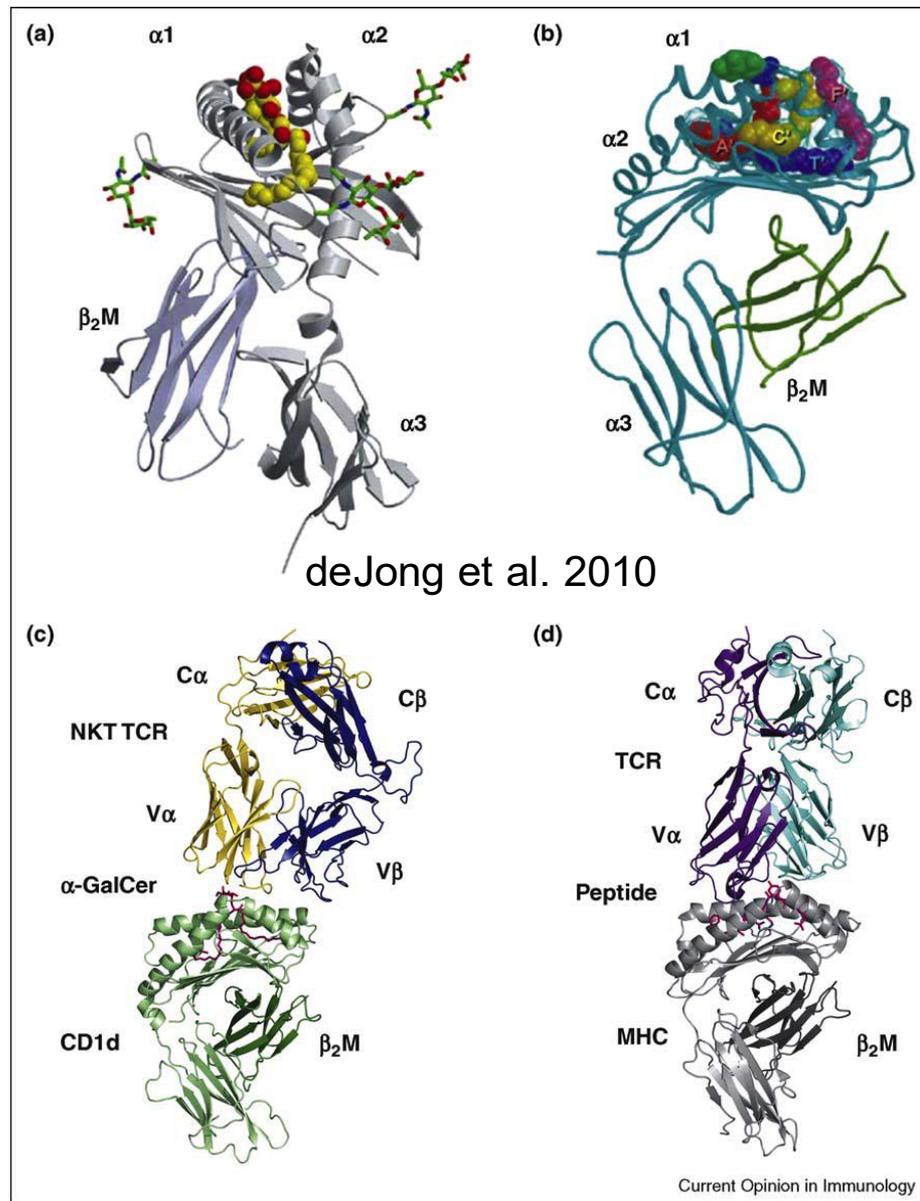
Formed by non-MHC encoded α -chain and soluble $\beta 2m$

Cytoplasmic tail contains endosomal localization sequence

Expressed by B cells, T cells, dendritic cells, epithelial cells

Present glycolipids in the endosomal/lysosomal compartment (MIIC)

Mycobacterial lipids and lipopeptides, e.g.
lipoarabinomannan, mycobactin

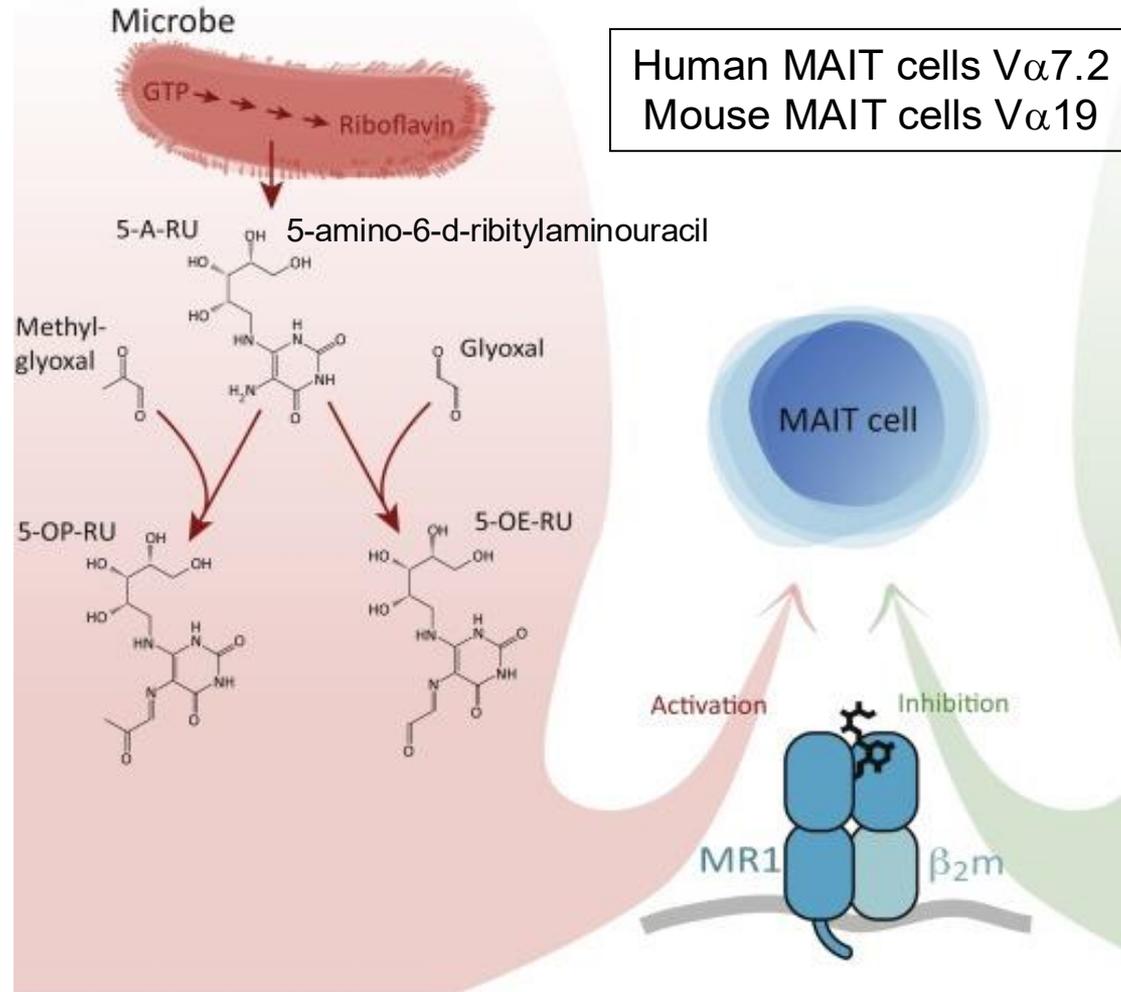


Human NKT cells $V\alpha 24J\alpha 18$
 Mouse NKT cells $V\alpha 14J\alpha 18$

(a) The crystal structure of CD1a in the presence of the self-antigen sulfatide [48]. In green are three N-linked sugar moieties while yellow spheres are ligand (3-O-sulfogalactosylceramide) carbon atoms. (b) CD1b was crystallized in complex with both phosphatidylinositol (PI) and a ganglioside GM2 [66]. (c) The first co-crystal of a CD1 family member with a TCR was generated for human CD1d- α -GalCer in complex with the iNKT TCR revealing a novel binding mode that had not previously been seen with classical MHC class I-restricted and II-restricted TCR [62]. (d) HLA-B8-peptide-TCR (LC13) complex

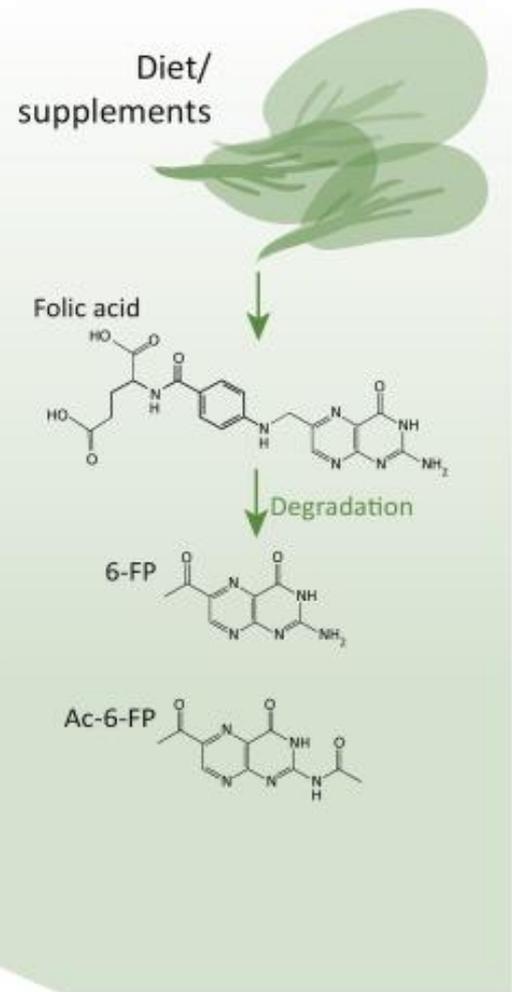
MR-1 presents vitamin B2 and B9 biosynthesis derivatives to MAIT cells

Vitamin B2 derivatives



Human MAIT cells V α 7.2
 Mouse MAIT cells V α 19

Vitamin B9 derivatives

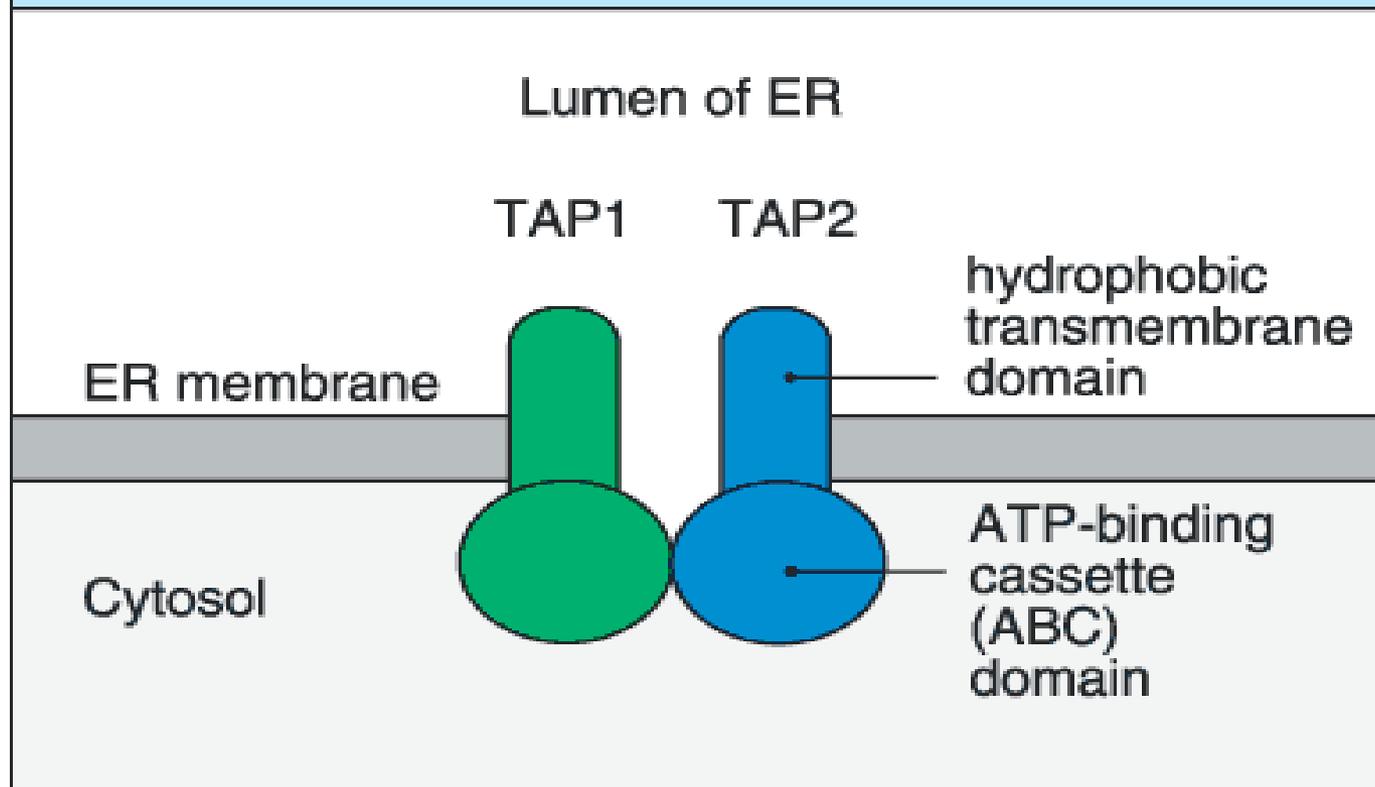


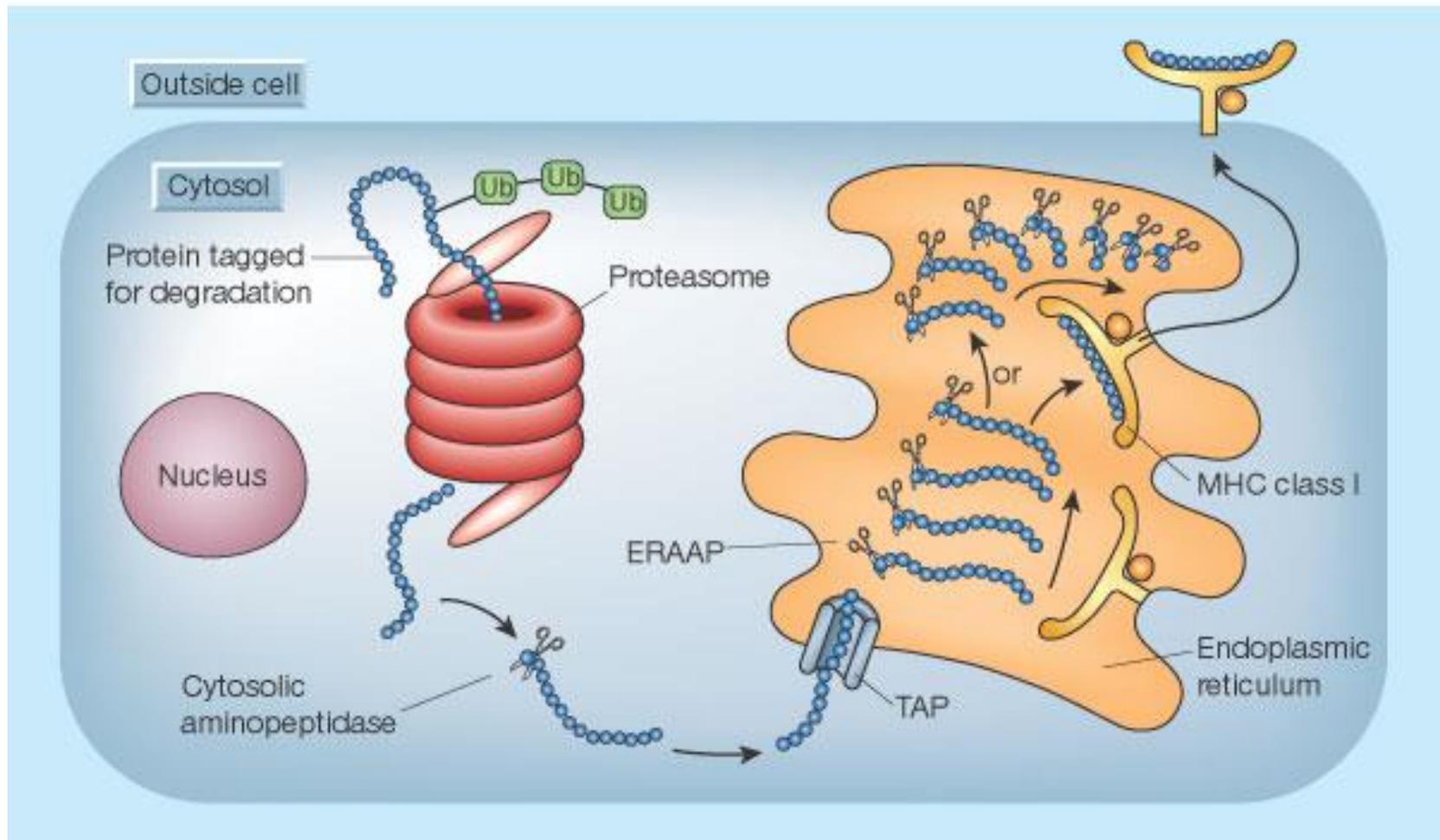
5-(2-oxopropylideneamino)-6-d-ribitylaminouracil (5-OP-RU)
 5-(2-oxoethylideneamino)-6-d-ribitylaminouracil (5-EP-RU)

6-formyl-pterin
 acetyl-6-formyl-pterin

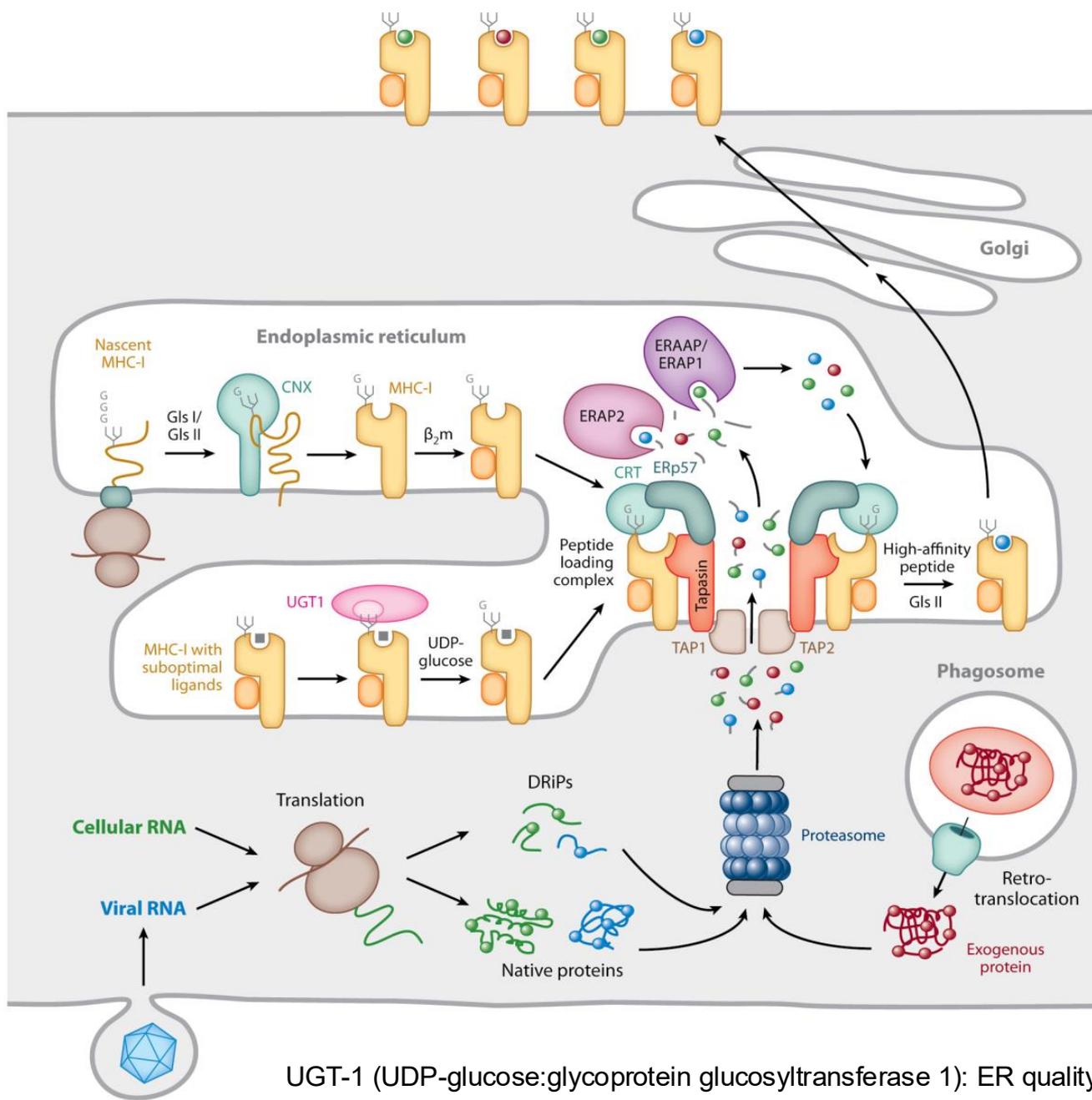
Intracellular assembly of MHC class I peptide complexes

Schematic diagram of TAP





26S proteasome (20S proteolytic core + 19S regulatory complexes)
 constitutive and inducible subunits of immunoproteasome (LMP2, LMP7 and MECL-1, PA28)
 ERAAP ER aminopeptidase associated with antigen processing



UGT-1 (UDP-glucose:glycoprotein glucosyltransferase 1): ER quality gatekeeper

Gls I/II – alpha-glucosidase I/II



Blum JS, et al. 2013.

Annu. Rev. Immunol. 31:443–73

MHC class I Antigen Presentation of Cytosolic Peptides

Protein degradation in the cytosol

Proteasomes - multicatalytic protease complex involved in degradation of ubiquitin-marked proteins

Two types of proteasomes with constitutive and inducible subunits (LMP2, LMP7 and MECL-1, regulator PA28)

Peptide Transport

TAP-1/TAP-2 (transporters associated with antigen presentation) ABC (ATP-Binding Cassette) family members

Molecules involved in MHC class I presentation

Chaperones: calnexin, calreticulin, tapasin

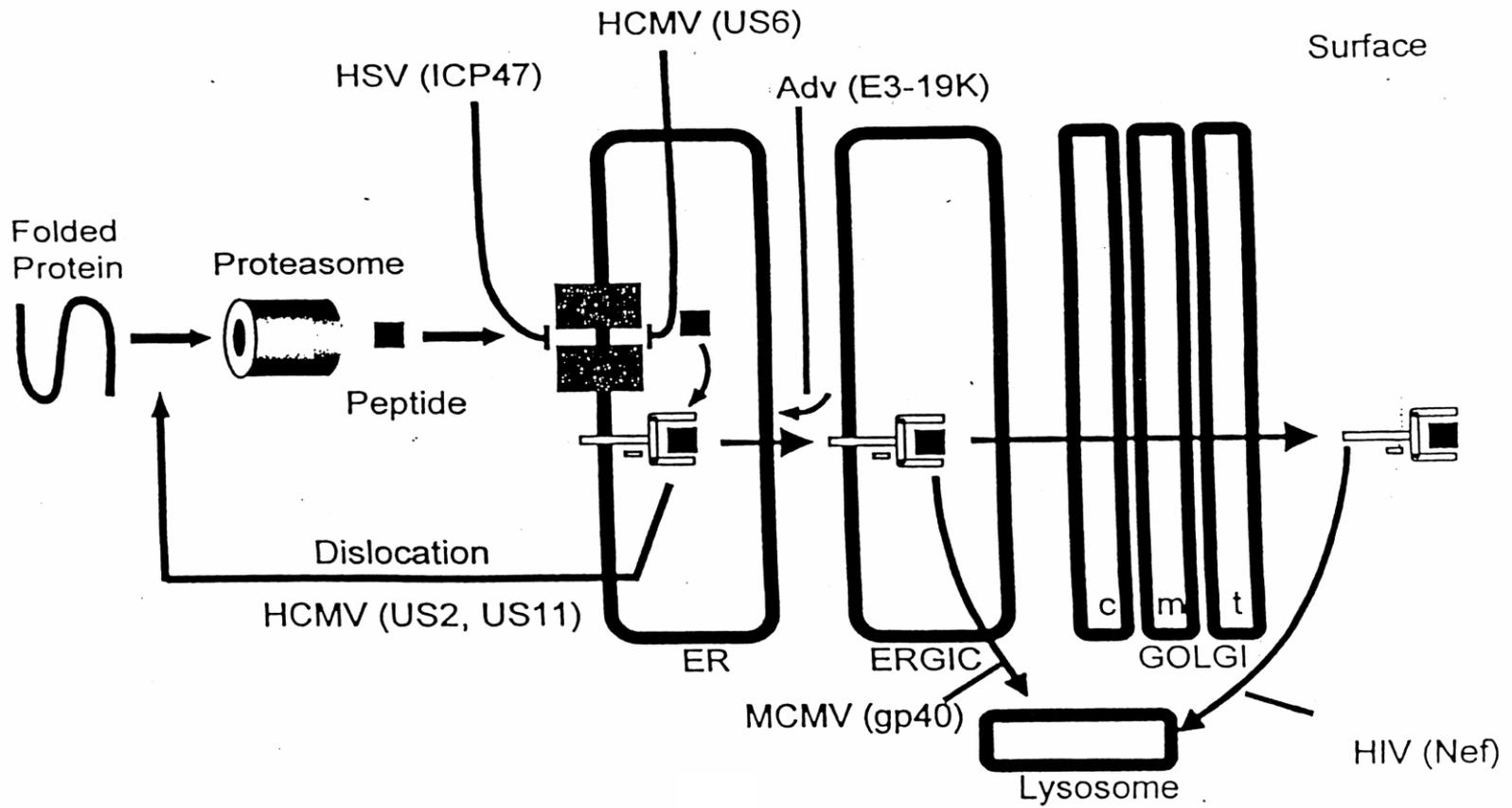
Erp57 thioreductase

TAP-1/TAP-2

Proteolytic enzymes: proteasomes, aminopeptidases (ERAAP/ERAP-1; ERAP-2)

Expression of MHC class I, TAP, LMP-2, -7, MECL-1 and PA28 is induced by IFN- γ

Figure 2B. Viral proteins that block class I assembly



Antigen presenting cells

cTEC

mTEC

Dendritic cells (cDC1; cDC2; PDC, Langerhans cells, etc.)

Macrophages

B cells

Fibroblast-like stromal (FLS) cells

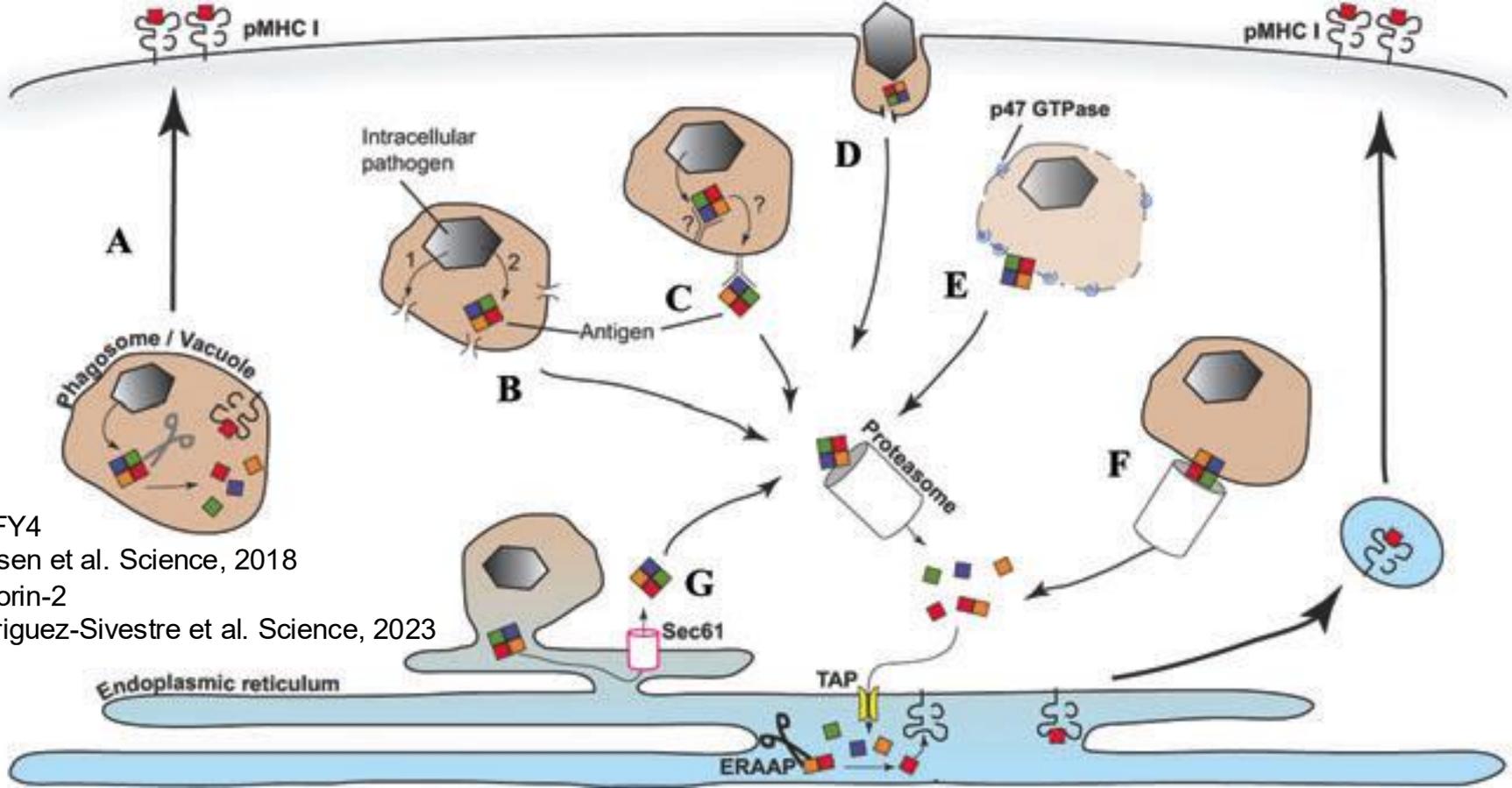
T cells (humans, rats)

Endothelial cells

Epithelial cells

MHC class II regulation by CIITA (RFX5, NF-Y)

CROSS-PRIMING/CROSS-PRESENTATION



WDFY4
 Theisen et al. Science, 2018
 Perforin-2
 Rodriguez-Sivestre et al. Science, 2023

Schematic model for generation of peptide major histocompatibility complex class I (pMHC I) from vacuolar and cytosolic processing pathways. (A) In the transporter associated with antigen processing (TAP)-independent vacuolar pathway, antigens are degraded and loaded on MHC I molecules within the phagosome/vacuole and displayed on the cell surface. (B–G) In the TAP-dependent cytosolic pathway, the antigen gains access to the cytosol where it is processed primarily by the proteasome. Several models have been proposed that differ in the manner the antigenic material enters the cytoplasm. (B) A secreted factor forms pores in the vacuole membrane, allowing for subsequent diffusion of antigen. (C) The antigen is shuttled to the cytoplasm through an active transport mechanism that may involve recognition of specific motifs and/or a specific translocation machinery. (D) An early break in the plasma membrane during invasion permits passage of the antigen to the cytoplasm (“kiss and spit”). (E) Immunity related GTPases (IRGs) are recruited to the vacuole membrane. Their action mediates membrane disruption and release of potentially antigenic content into the cytosol. (F) The antigen is inserted into the vacuole membrane with a topology consistent with action of the proteasome on the cytosolic side. (G) Direct fusion of the vacuole with the host endoplasmic reticulum (ER) delivers the antigen to the Sec61-based retro-translocation machinery, which shuttles the antigen to the cytoplasm. In all cases, antigen processing occurs in a proteasome-dependent fashion. Antigenic peptide precursors are then imported into the ER for possible trimming by ER aminopeptidase associated with antigen processing (ERAAP) and final MHC I loading.

MHC

(*Major Histocompatibility gene Complex*)

POLYGENIC and highly POLYMORPHIC gene complex

MHC class II

Classical

Non-classical

MHC class I (Ia)

MHC class I (Ib)

MHC class II	Classical MHC class I (Ia)	Non-classical MHC class I (Ib)
Polymorphic MHC-encoded α and β chains	Polymorphic MHC-encoded α chain+β2m	Non-Polymorphic MHC-encoded α chain+β2m

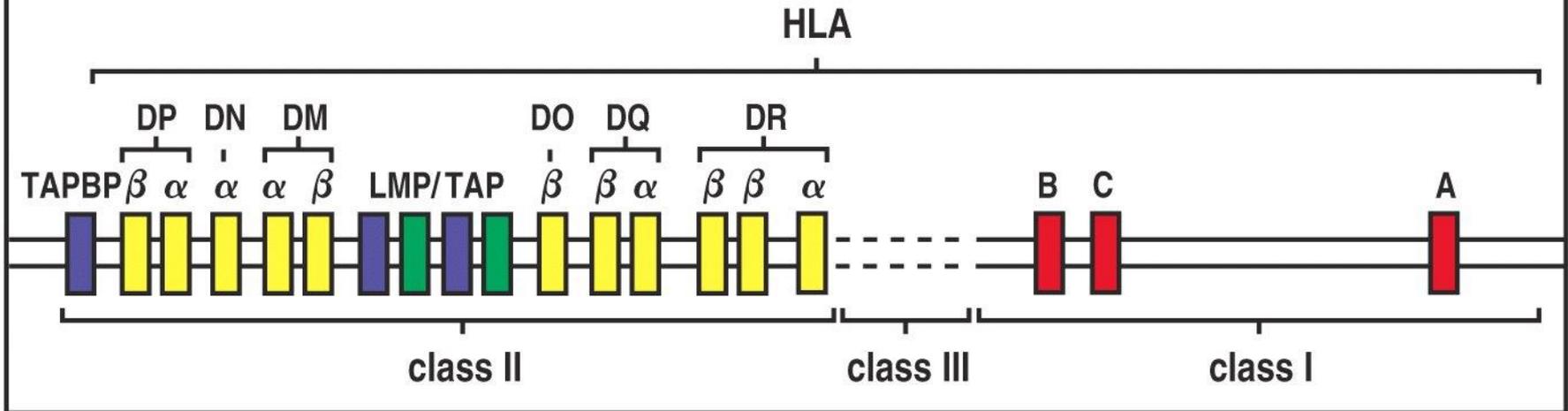
In humans HLA (*Human Leukocyte Antigen*)

HLA-DR	HLA-A	HLA-E, HLA-G
HLA-DQ	HLA-B	MICA/MICB
HLA-DP	HLA-C	

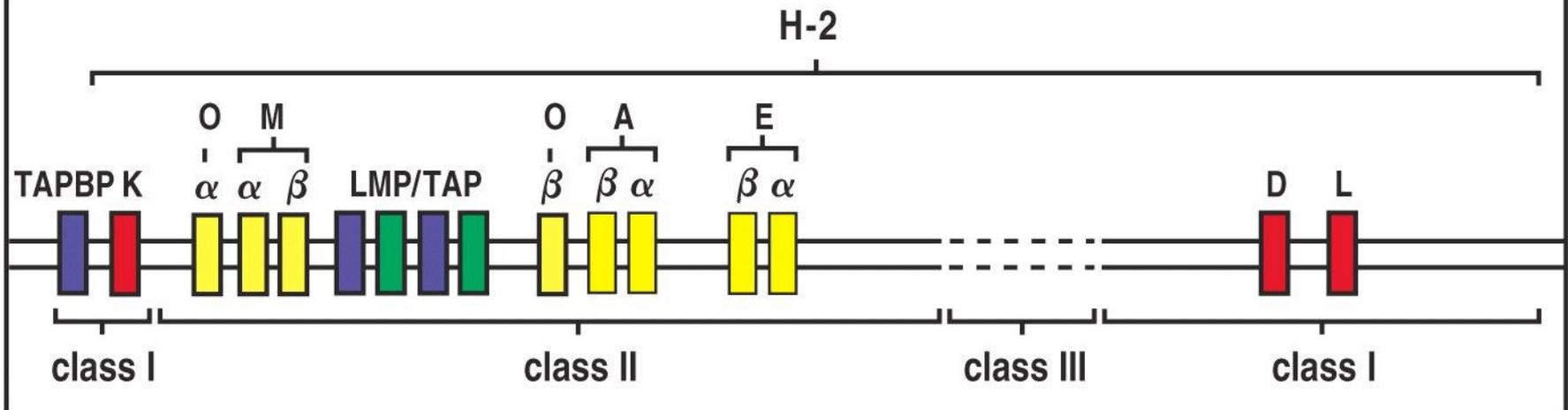
In mice H-2 (*Histocompatibility locus-2*)

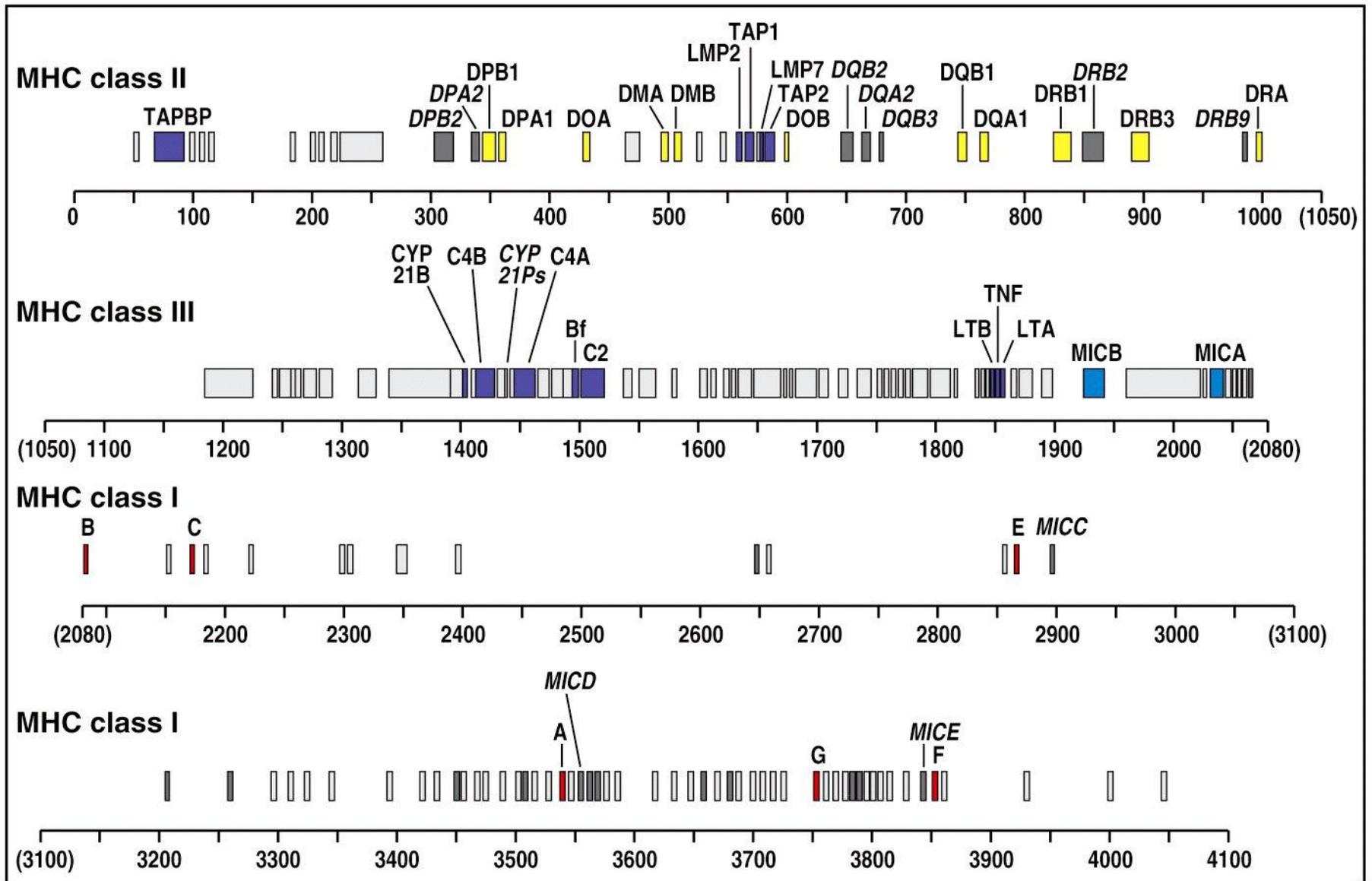
I-A	H-2K	H-2M3
I-E	H-2D	Qa-1
	H-2L	

Gene structure of the human MHC



Gene structure of the mouse MHC





DISCOVERY OF MHC

**Tumor and skin transplantation
between INBRED strains of mice**

**INBRED strains of mice are derived by
sequential brother-sister mating.**

All animals are genetically identical.

All alleles are homozygous.

MHC^a -> MHC^a acceptance

MHC^a -> MHC^b rejection

MHC POLYMORPHISM

A particular combination of MHC alleles found on a single chromosome is called MHC haplotype

Inbred mice

H-2^d, b, k, s etc.

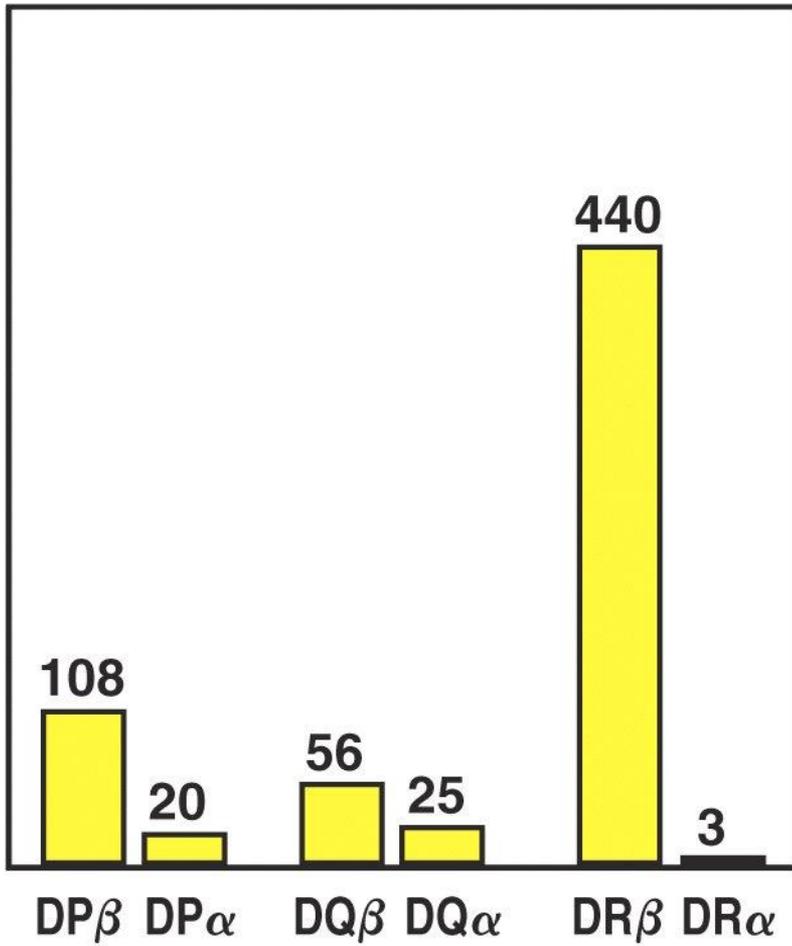
H-2^d

H-2K^d H-2D^d H-2L^d I-A^d I-E^d

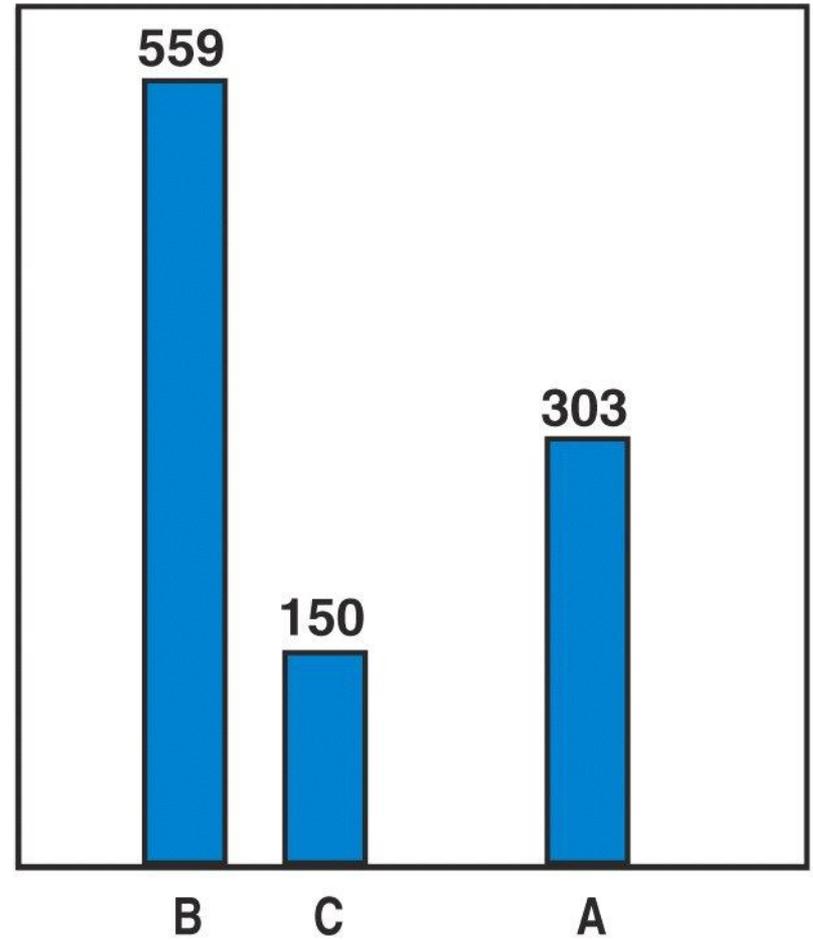
Outbred humans

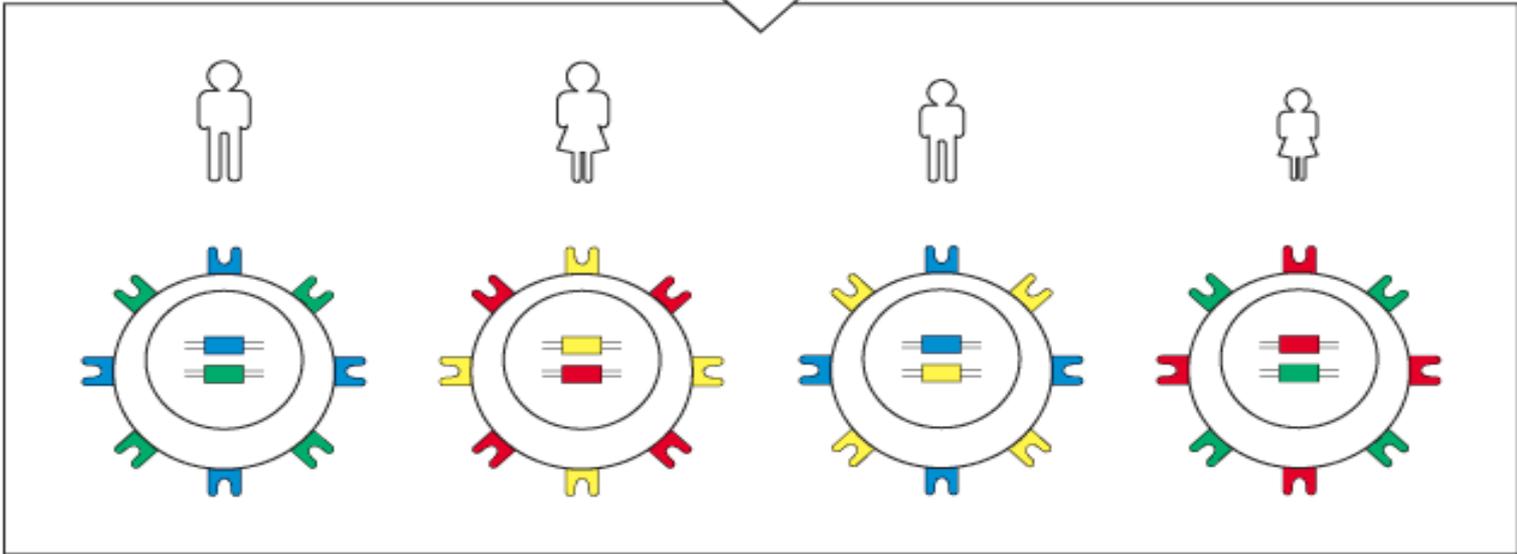
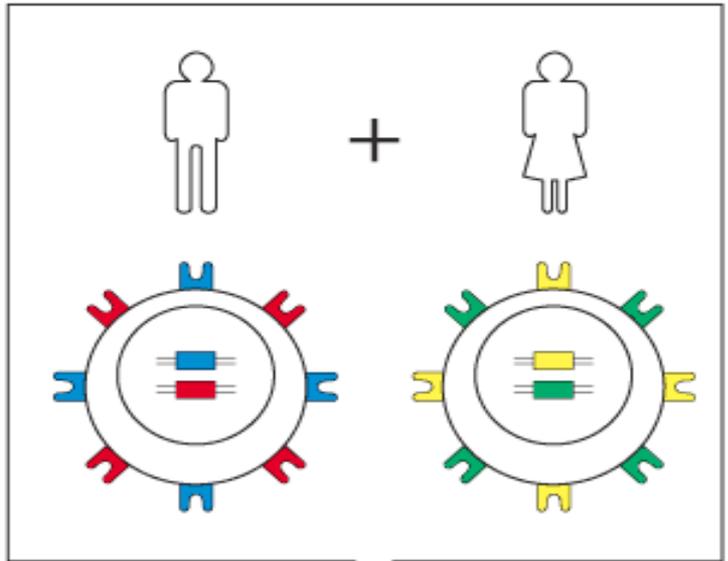
HLA-A2, HLA-B27, HLA-Cw3, HLA-DR4, HLA-DQ52, HLA-DP3

MHC class II

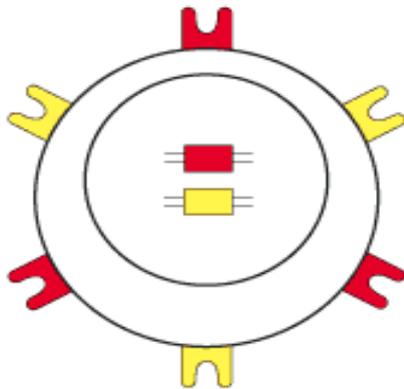


MHC class I

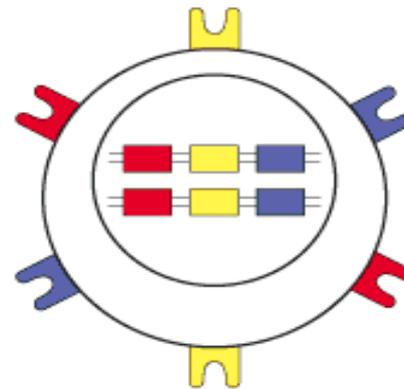




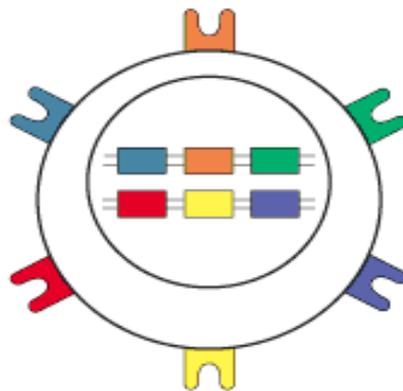
Polymorphism

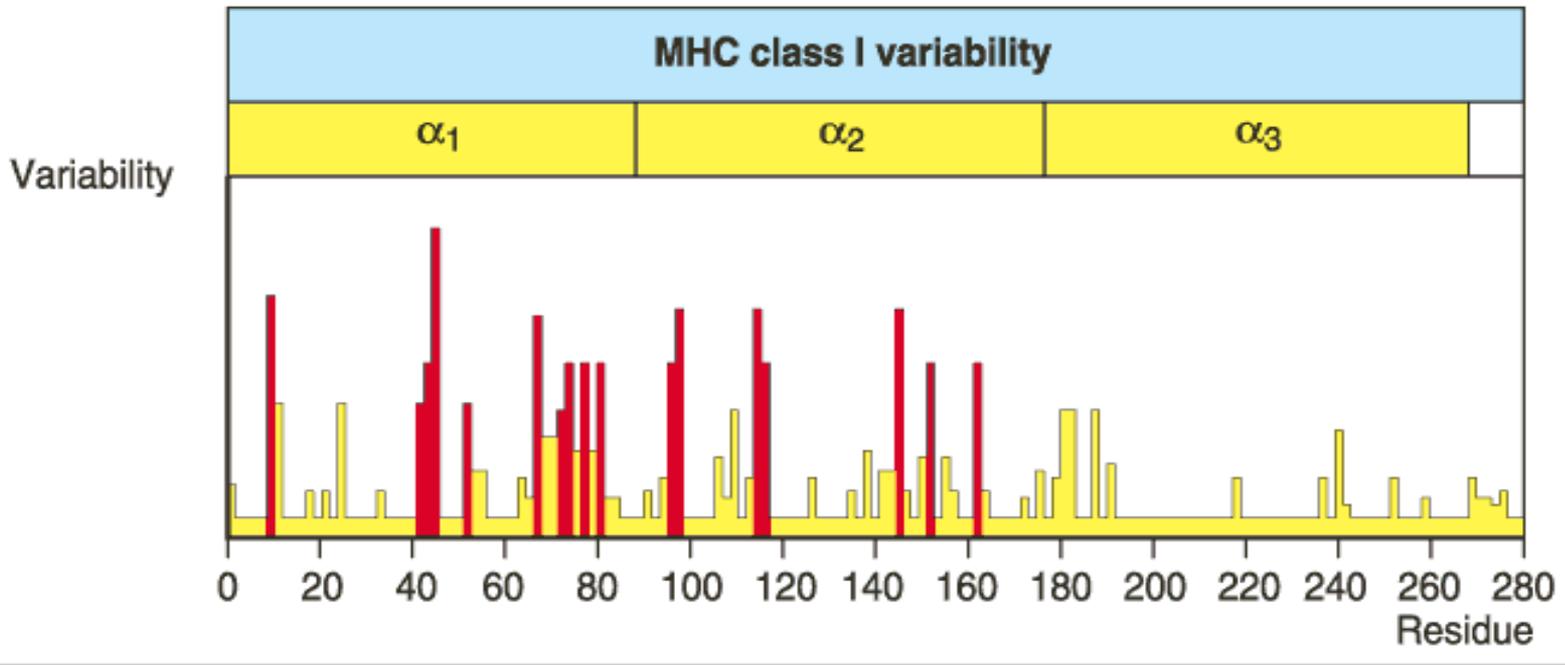
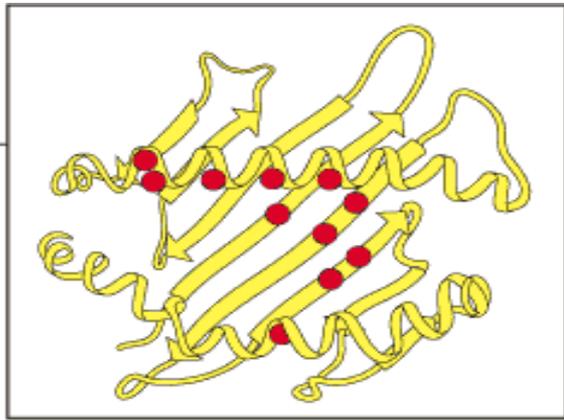
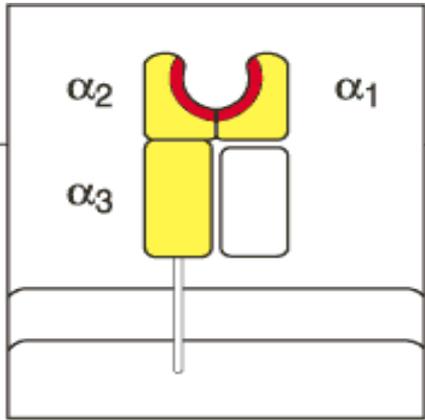


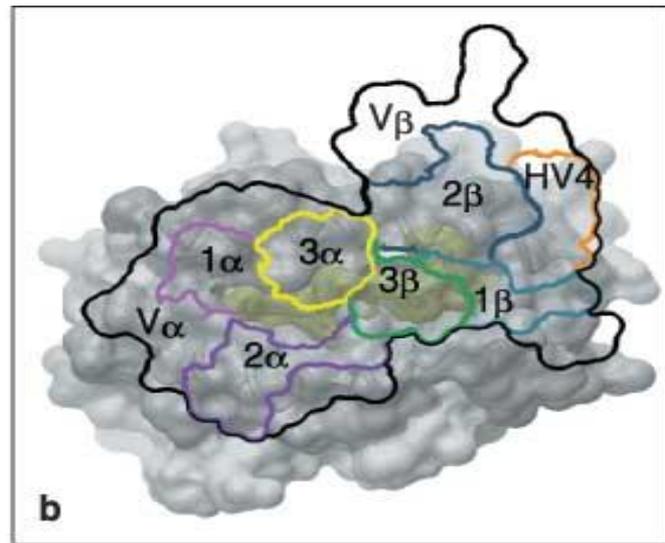
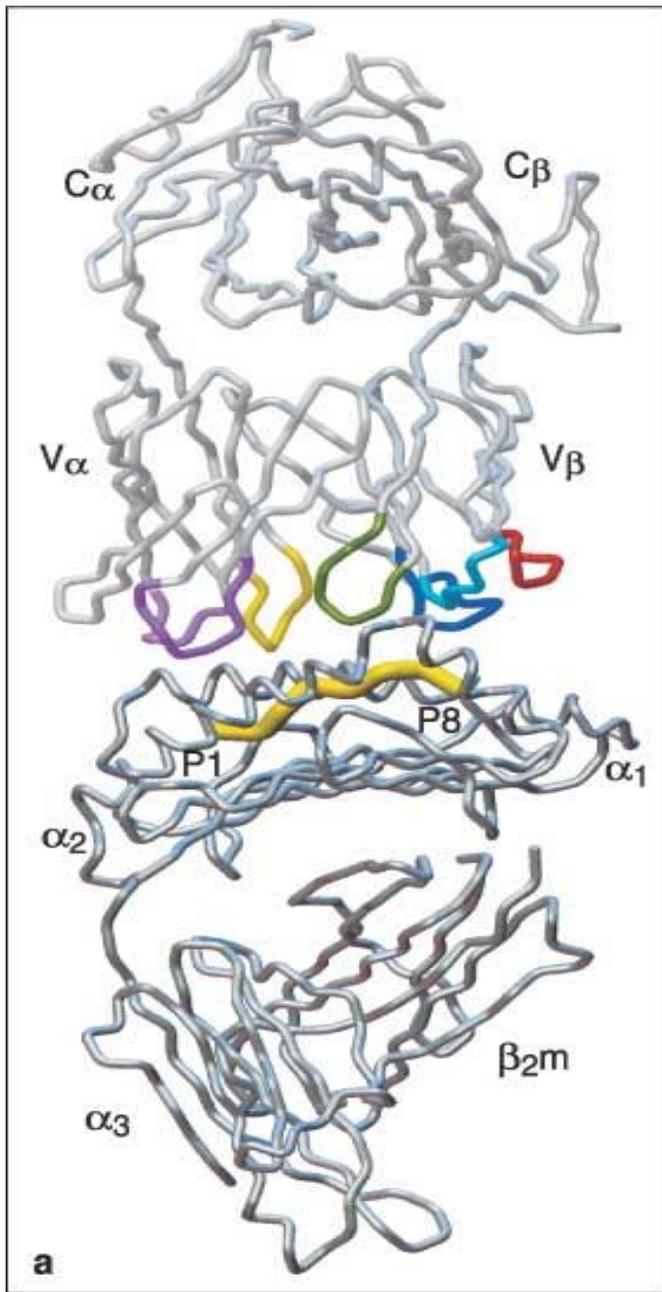
Polygeny



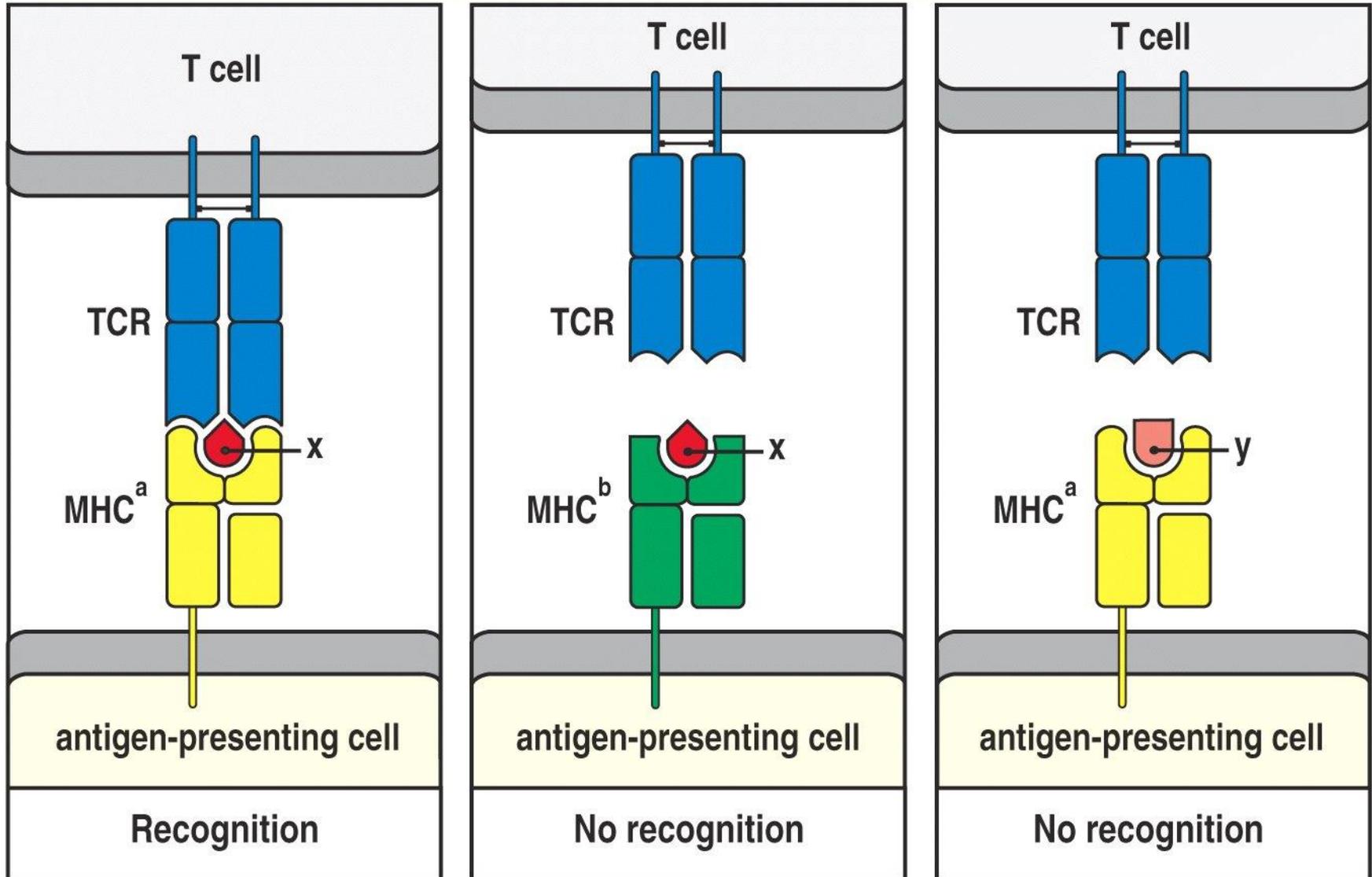
Polymorphism and polygeny







MHC restriction



altered 'self' recognition